

Role of Nonalcoholic Fatty Liver Disease in Periodontitis: A Bidirectional Relationship

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Abstract

Nonalcoholic fatty liver disease (NAFLD) and periodontitis share common risk factors such as obesity, insulin resistance (IR), and dyslipidemia, which contribute to systemic inflammation. It has been suggested that a bidirectional relationship exists between NAFLD and periodontitis, indicating that one condition may exacerbate the other. NAFLD is characterized by excessive fat deposition in the liver and is associated with low-grade chronic inflammation. There are several risk factors for the development of NAFLD, including gender, geriatric community, race, ethnicity, poor sleep quality and sleep deprivation, physical activity, nutritional status, dysbiosis gut microbiota, increased oxidative stress, overweight, obesity, higher body mass index (BMI), IR, type 2 diabetes mellitus (T2DM), metabolic syndrome (MetS), dyslipidemia (hypercholesterolemia), and sarcopenia (decreased skeletal muscle mass). This systemic inflammation can contribute to the progression of periodontitis by impairing immune responses and exacerbating the inflammatory processes in the periodontal tissues.

Furthermore, individuals with NAFLD often exhibit altered lipid metabolism, which may affect oral microbiota composition, leading to dysbiosis and increased susceptibility to periodontal disease. Conversely, periodontitis has been linked to the progression of NAFLD through mechanisms involving systemic inflammation and oxidative stress. Chronic periodontal inflammation can release pro-inflammatory cytokines and bacterial toxins into the bloodstream, contributing to liver inflammation and exacerbating hepatic steatosis. Moreover, periodontitis-induced oxidative stress may promote hepatic lipid accumulation and IR, further aggravating NAFLD. The interplay between NAFLD and periodontitis underscores the importance of comprehensive management strategies targeting both conditions. Lifestyle modifications such as regular exercise, a healthy diet, and proper oral hygiene practices are crucial for preventing and managing these interconnected diseases. Additionally, interdisciplinary collaboration between hepatologists and periodontists is essential for optimizing patient care and improving outcomes in individuals with NAFLD and periodontitis.

Categories: Public Health, Nutrition, Dentistry

Keywords: nonalcoholic fatty liver disease, inflammatory bowel disease, low socioeconomic status, oral hygiene status, distorted lipid metabolism, dyslipidemia, insulin resistance, obesity, chronic generalized periodontitis, naflid

Introduction And Background

Periodontitis is an inflammatory chronic tooth disorder initiated by poly-pathogenic microbes remaining in plaque biofilm [1-3]. Periodontitis is also known as grave gum infectious disorder. Chronic periodontitis is influenced by various factors like genetic predisposition, age, alcohol, smoking, obesity, calculus, low socioeconomic status, systemic diseases (e.g., diabetes mellitus, cardiovascular disease, osteoporosis, etc.), and the availability of pathogenic microbes [4-6]. This leads to the destruction of supporting tissues and alveolar bone [7], which, if the patient does not receive appropriate healthcare, can, in turn, lead to tooth loss [8]. It is the sixth most prevalent human disease, which affects approximately 11.2% of the population [9]. The mouth cavity's homeostatic equilibrium is lost because the oral microbiome is disintegrated [10-13]. Pathobionts (opportunistic pathogenic microbes) enter a healthy microbiome, instigating the development of a dysbiotic plaque and biofilm [10-13]. Henceforth, oral pathological issues, such as caries and periodontic diseases, are recognized because of the upset equipoise between the microbiota and the human host [14,15]. Dysbiosis of the oral microbiota gives rise to a probability of pathogenic infection among internal organs through gut microbiota dysbiosis [16-18]. It arouses poor mucosal wall physiology, leading to steatohepatitis through the enterohepatic circulation [19]. Another study revealed that the oral microbiome has supervisory authority over the gut microbiome [17]. The Human Oral Microbiome Database (HOMD) makes it web accessible (www.homd.org). The HOMD includes 619 taxa in 13 phyla, as follows: *Actinobacteria*, *Bacteroidetes*, *Chlamydiae*, *Chloroflexi*, *Euryarchaeota*, *Firmicutes*, *Fusobacteria*, *Proteobacteria*, *Spirochaetes*, *SR1*, *Synergistetes*, *Tenericutes*, and *Saccharibacteria* (TM7) [20]. Furthermore, the major bacterial genera in the healthy rima oris are depicted in Table 1 [21-26].

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Types	Cocci	Rods
Gram-positive	<i>Abiotrophia</i> , <i>Peptostreptococcus</i> , <i>Streptococcus</i> , <i>Stomatococcus</i>	<i>Actinomyces</i> , <i>Bifidobacterium</i> , <i>Corynebacterium</i> , <i>Eubacterium</i> , <i>Lactobacillus</i> , <i>Propionibacterium</i> , <i>Pseudoramibacter</i> , <i>Rothia</i>
Gram-negative	<i>Moraxella</i> , <i>Neisseria</i> , <i>Veillonella</i>	<i>Bacteroidales</i> , <i>Campylobacter</i> , <i>Capnocytophaga</i> , <i>Desulfobacter</i> , <i>Dialister</i> , <i>Fusobacterium</i> , <i>Prevotella</i> , <i>Firmicutes</i> , <i>Lautropia</i> , <i>Desulfovibrio</i> , <i>Eikenella</i> , <i>Fusobacterium</i> , <i>Hemophilus</i> , <i>Leptotrichia</i> , <i>Seimonas</i> , <i>Simonsiella</i> , <i>Treponema</i> , <i>Wolinella</i>

TABLE 1: List of microbiota presence in the healthy oral cavity

Antony van Leeuwenhoek first reported the presence of microbes' oral cavity in the late 16th century (1684) [27-29]. An American dentist and the first oral microbiologist, Willoughby Dayton Miller, earliest (1891), generated the concept that oral pathogens and infection affect other body parts, causing multiple systemic disorders [30,31]. Billings in 1913 reported that dental infection instigated several systemic diseases, for example, endocarditis, nephritis, rheumatoid arthritis, etc. [32]. A considerable portion of healthcare professionals who trust dental infection, biofilm, plaque, and their breakdown products enter the blood circulation and instigate several retrogressive, wasting, or systemic diseases. Consequently, tooth extraction became a very accepted therapeutic intervention for multiple systemic disorders [31]. The discovery of molecular procedures for microbiology, such as DNA sequencing, DNA-DNA hybridization, and polymerase chain reaction, has led to an opportunity to build a wide-ranging depiction of the assortment and symphony of the oral microbiome [29]. Our mouth orifice has the second most prevalent and assorted microbiota after the colon, embracing over 700 different microbes. The oral cavity fosters abundant microorganisms that comprise viruses, bacteria, protozoa, and fungi [28]. Dysbiosis of oral microbiota causes oral (chronic periodontitis) and multiple long-lasting inflammatory or degenerative systemic health disorders such as inflammatory bowel disease [33], squamous cell carcinoma of mouth [34,35], obesity [36], rheumatoid arthritis [37,38], type 2 diabetes mellitus (T2DM) [27,39], atherosclerosis [40], cardiovascular diseases [41], preterm birth [42], Alzheimer's disease [43], and many more [44-47].

Globally, nonalcoholic fatty liver disease (NAFLD) is a frequent instigating factor of long-lasting hepatic illness [48], with a worldwide prevalence of around 30% [49]. Schaffner and Thaler first coined the term NAFLD [50]. Ludwig et al. reported that a persistent type of fatty hepatic ailment histologically looks like alcoholic steatohepatitis, causing hepatocellular injury. Nevertheless, these liver disorder cases possess no history of alcohol consumption. This research group termed this condition nonalcoholic steatohepatitis (NASH) [51]. Two types of NAFLD have been recognized: NASH and nonalcoholic fatty liver (NAFL) [52]. NAFL is identified by the buildup of lipids in hepatic cells without causing any cellular destruction and with trivial or no inflammatory process in the hepatic lobe [53-55]. There is another hepatic disorder named metabolic dysfunction-associated fatty liver disease (MAFLD). Readers often get confused with a similar hepatic disease. MAFLD was anticipated as a more suitable name than NAFLD since this terminology preferably delineates the pathophysiology of this hepatic disorder and its related metabolic anomalies in 2020 [56,57]. MAFLD has appeared as the most frequent persistent liver disorder in both rich and poor economic nations because of the mushrooming in the frequency of overweightness, obesity, and metabolic syndrome (MetS) [58]. One meta-analysis was conducted regarding study evaluations of periodontal disease in NAFLD. "The odds ratio (1.91; 95% CI 1.21-3.02; p = 0.006) indicates that the chance of presenting periodontal disease is 91%" greater among patients with NAFLD than patients without NAFLD [59]. Chen et al. reported that affirmative relations link periodontal disorder and hepatic cirrhosis (OR = 2.28; 95% CI = 1.50-3.48) and NAFLD (OR = 1.19; 95% CI = 1.06-1.33) [60]. Aguiar et al. revealed that patients with periodontal disorders instigate the progression of NAFLD because of distorted sphingolipid metabolism that headed to intensified insulin resistance, hepatic inflammation [59], and mitochondrial dysfunction [61].

Problem statement of the study

This paper aims to investigate the complex interplay between NAFLD and periodontitis. Despite the established association between these two conditions and their shared factors, such as obesity, insulin resistance, and chronic inflammation, there remains a gap in understanding the bidirectional nature of their relationship. This study also aims to address. Questions like the extent of the influence of NAFLD on the development and progression of periodontitis. How does periodontitis contribute to the pathogenesis and exacerbation of NAFLD? What underlying mechanisms link NAFLD and periodontitis, including systemic inflammation, dysbiosis, and oxidative stress? What are the clinical implications of the bidirectional relationship between NAFLD and periodontitis regarding disease management and treatment outcomes? This research will also contribute to advancing our understanding of the systemic consequences of periodontal disease and highlight the importance of interdisciplinary collaboration between hepatologists and periodontists in optimizing patient care.

Objective of the study

The primary objective of this review is to evaluate the possible connection or association between periodontal disease and NAFLD.

Review

Materials and methods

This narrative review paper explores the prevalence and severity of periodontitis in NAFLD patients, the connection between these two disorders, and their relationship to other body systems. These were all examined through a thorough literature assessment to compile relevant data. Databases such as PubMed, Google Scholar, and Google were hand-searched for articles published up to 2023. Keywords included were "periodontal disease," AND "nonalcoholic fatty liver disease," AND "nonalcoholic steatohepatitis," AND "dysbiosis," AND "oral-gut-liver axis." The articles included were based on their relevance to the connection between NAFLD and periodontal disease. Studies focusing on the relationship between periodontitis and NAFLD were selected. The quality of the evidence, study design, and methods for evaluating the rapport regarding the association between both conditions received particular attention (Figure 1). Almost all papers were downloaded from PubMed; only one utilized the website BioRender (<https://www.biorender.com/>).

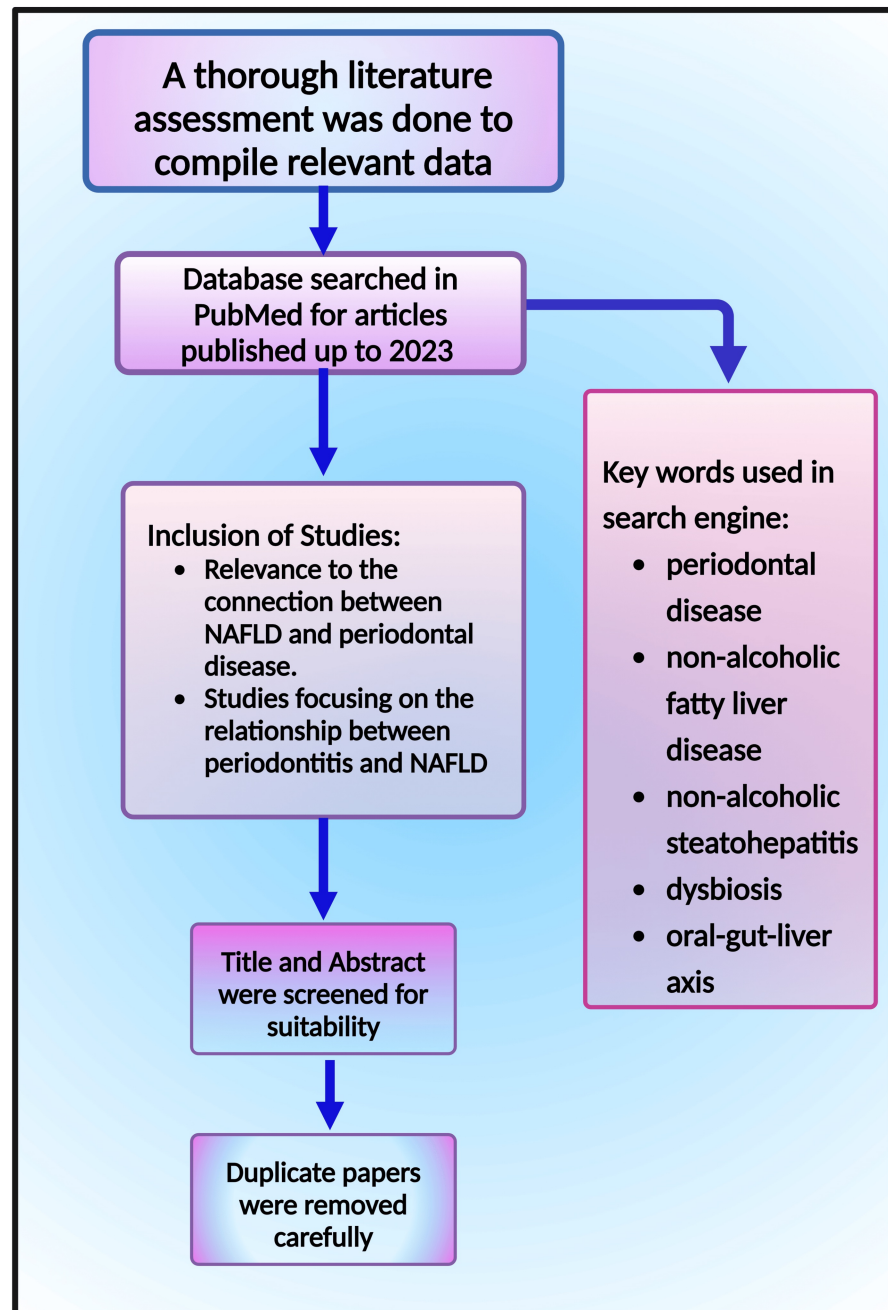


FIGURE 1: A flow chart illustrating the methodology of the study

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Review of literatures

The liver is the largest solid organ in the human body [63]. The liver engages in a dominant physiological function in metabolic homeostasis and is a paramount area for the creation, metabolism, stowage, and redistribution of carbohydrates, proteins, and lipids [64]. The liver detoxifies potentially toxic chemicals in exposed human beings [65]. The liver also processes substances like hemoglobin, bile salts, iron, vitamins, copper, and medications by absorbing, transforming, and releasing these products [66]. The liver, the largest parenchymal organ, obtains blood from the gastrointestinal tract (GIT) and systemic circulation due to its distinctive anatomical positioning. The enteric portal vein, which supplies the liver with 80% of its blood supply, is rich in nutrients, bacterial metabolites, and dietary antigens. The hepatic artery, which branches

off the abdominal aorta, provides the remaining 20% and serves as the liver's nutrient vessel [67]. Hepatocytes, comprising roughly 70%-85% of the liver's volume, are the primary functional cells within the organ. They play a pivotal role in controlling a portion of its immune capabilities (innate immunity) [68,69]. Hepatocytes are the primary contributors to the creation of the liver's extracellular matrix (ECM) [70,71]. Kupffer cells represent the principal resident macrophages within the liver and the maximum number of mononuclear phagocytes in the human body, constituting the primary cell population regarding the innate immune system [72-74]. The liver possesses its regenerative ability; henceforth, it can often recover from significant damage, yet continuous or chronic damage eventually leads to various chronic liver conditions [75].

The most reliable method (gold standard) for diagnosing and staging NAFLD is through liver biopsy and histopathology [76]. Iwasaki et al. conducted a cross-sectional study to determine the relationship between severe periodontitis and NAFLD in Japan. This research does abdominal ultrasonography to detect NAFLD. Among the research participants, around 28% had NAFLD with extreme cases of periodontal diseases. NAFLD cases had a significantly ($p < 0.05$) greater incidence of probing pocket depth (PPD) ≥ 4 mm than those cases who do not have NAFLD. There was a statistically significant ($p < 0.01$) difference observed in the adjusted odds ratios of having PPD ≥ 4 mm for NAFLD (odds ratio = 1.881; 95% CI 1.184-2.9870); after adjusting for body mass index (BMI), regular exercise habits, number of teeth present, presence of periodontitis, Brinkman index, raised serum C-reactive protein (CRP), blood pressure, sex, and age [77]. Abdominal ultrasound is highly effective at detecting both moderate and high levels of fat accumulation, making it a valuable diagnostic tool for identifying NAFLD, whether the deposits are present or absent [78,79]. Again, if distinguishing or diagnosing NAFLD from other chronic hepatic diseases becomes challenging or there's suspicion of nonalcoholic steatohepatitis, contemplating a hepatic biopsy and histopathological is the most recommended examination for appropriate diagnosis [80,81]. Indicators such as serum alanine aminotransferase (ALT), aspartate aminotransferase (AST), gamma-glutamyl transpeptidase, platelet count, albumin levels, triglycerides, cholinesterase, fasting plasma insulin, homeostasis model of assessment of IR, and various other biomarkers have been utilized to assess liver conditions (Figure 2) [82-84]. The blood indicators GGT, ALT, and AST are measured in liver function tests [85]. Key enzymes in the metabolism of amino acids, AST and ALT, are found in the cells of essential organs such as the liver, kidneys, and heart [86-88].

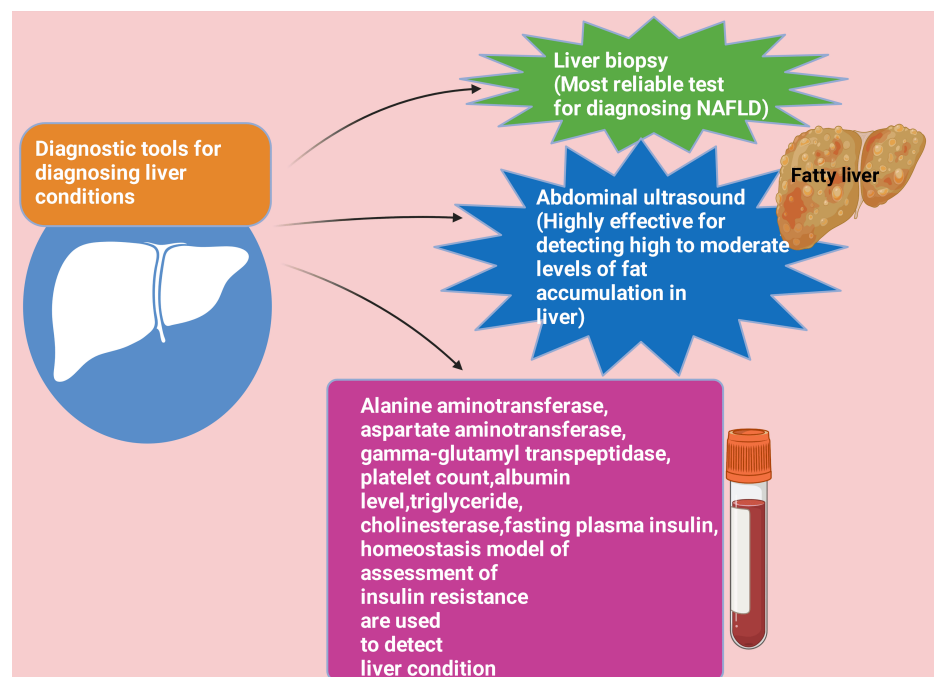


FIGURE 2: Various diagnostic tools for detecting liver conditions

NAFLD: Nonalcoholic fatty liver disease

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Common risk factors for NAFLD

Genetic tendencies, gender, geriatric community, race, ethnicity, poor sleep quality and sleep deprivation, physical activity, nutritional status, dysbiosis of the gut microbiota, increased oxidative stress, overweight, obesity, higher BMI, insulin resistance (IR), T2DM, MetS, dyslipidemia (hypercholesterolemia), and sarcopenia (decreased skeletal muscle mass) are common risk factors for developing NAFLD (Figure 3) [89,90].

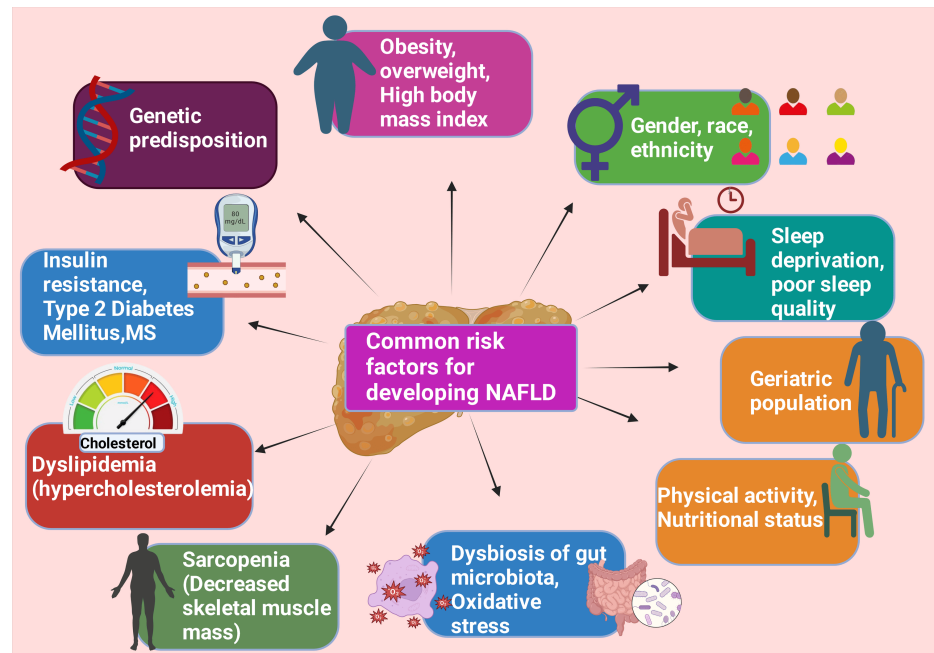


FIGURE 3: Illustrates the common risk factors for developing NAFLD

NAFLD: Nonalcoholic fatty liver disease; MS: metabolic syndrome; O₂: super oxide

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Oral-gut-liver axis

"The oral-gut-liver axis is a concept that aims to understand the potential relationships between different organs, particularly the oral cavity, gut, and liver" [91]. Individuals suffering from periodontitis possess a distinctive dysbiotic oral microbiome [92], leading to the continuous, unwitting ingestion of pathogens and noxious metabolites from the saliva and dental plaque to the gut through blood circulation or enteral paths [93,94]. Dysbiotic oral pathogens, when translocating to the gastrointestinal tract, similarly cause gut dysbiosis [95]. These pathogenic microbes and lethal metabolites enter the liver through the portal vein, causing adverse hepatic consequences [96]. These metabolites are derived from dysbiotic oral pathogens, often known as pathogen- and damage-associated molecular patterns (PAMPs; DAMPs) [92]. They activate a predisposed host immune system in the periodontal disease setting, which could influence the escalation of a range of systemic pro-inflammatory cytokines and chemokines [97,98], increasing the possibility of impairing the hepatic physiology function [92,99]. It has been reported that the gut microbiota is reflected as a biological barricade, defending against settling and expanding the population of pathogenic microbes within the GIT [100]. An impaired gut microbiota disrupts the integrity of intestinal tight junctions, leading to heightened mucosal permeability [82,100,101]. Analogously, hepatic poor performance function also worsens the existing disease process and clinical sequels within the buccal cavity [102].

NAFLD and periodontitis

Kobayashi et al. reported that the way of living associated with various severe illnesses such as hepatic disorders, NASH, and NAFLD also has a potential connection with periodontal diseases [103]. The manner of living remains the independent risk factor that has the potential to be alterable for several noncommunicable diseases (NCDs) [104-106]. The pathogenesis of NAFLD remains not definitively known, although this disease is quite common around the globe [107]. Nevertheless, several precipitating factors are common for developing periodontitis and NAFLD [108]. Furthermore, the progression of both these diseases leads to the excess synthesis of systemic pro-inflammatory signaling proteins and reactive oxygen species

(ROS); IR intensifies the process and is mainly observed among T2DM and obese cases [108-111]. Kuroe et al. reported that those cases of moderate to severe periodontitis augmented the probability of emerging liver fibrosis with an odds ratio of 2.06 (0.89-4.76) among the Japanese population [112]. Additionally, NAFLD-related advanced fibrosis usually observes reduced microbial multiplicity, frequently complemented by the high level of Gram-negative microbes [113,114]. Helenius-Hietala et al. revealed that advanced cases of severe periodontitis when concurrently diagnosed with NAFLD among the Finnish population require immediate hospitalization with fatal clinical outcomes and hepatic carcinoma [115].

Red complex pathogenic microbes most frequently instigate periodontal disease, including *P. gingivalis*, *Treponema denticola*, and *Tannerella forsythia* (*Bacteroides forsythias*) [116,117]. *Porphyromonas gingivalis* (a Gram-negative anaerobic bacteria) is one of the principal pathogenic microbes extensively found among patients with periodontal disorders [107,118]. *P. gingivalis* infection has been revealed to be responsible for the deterring feature for sprouting IR, NAFLD, NASH, and MetS [107,119]. Elevated *P. gingivalis* levels in the oral cavity, elevated endotoxin levels, and elevated antibody titers for *P. gingivalis* are linked to amplified hepatic rigourousness, which can be detected by MRE [120,121]. Periodontopathic microbes such as *P. gingivalis* flow down from the mouth to the gut and directly enter the blood through a bleeding gum, a typical feature of periodontitis [122,123]. Periodontopathic *P. gingivalis* later instigates gut dysbiosis [124]. *P. gingivalis* provokes intestinal microenvironmental disorder and increases the permeability of the intestinal wall, thus facilitating the direct passage of pathogenic microbes and their metabolites to the bloodstream and hepatic tissues [125-127]. Thus, periodontal pathogenic microbes and their metabolites in hepatic tissue instigate hepatic inflammation and the progression of fatty liver disease [19,82,103,128].

Periodontal-disease-producing pathogens can synthesize inflammatory mediators such as tumor necrosis factor-alpha (TNF- α), interleukin 6 (IL-6), and interleukin-1 family (IL-1), which results in inflammation-triggering vascular endothelial damage and when entering the blood instigates systemic inflammation [44,129]. Periodontal pathogens trigger neutrophils and monocytes, producing excessive pro-inflammatory cytokines [129]. Hence, systemic inflammations are frequently linked with periodontitis and the development of IR [129,130] by increasing the levels of serum adipocytokines (TNF- α , IL-6, and IL-1) [44,129]. Additionally, these Gram-negative anaerobic periodontal pathogens produce lipopolysaccharides (LPS). LPS "is a virulence factor of Gram-negative bacteria with a crucial importance to the bacterial surface integrity" [131]. Furthermore, circulating LPS can potentiate ongoing inflammation and oxidative stress [132]. LPS binds activated Toll-like receptors (TLRs) such as TLR-4, which increases oxidative stress and chronic inflammation [108,133,134]. Therefore, all these pathogenic difficulties further potentiate additional harm to the liver's parenchymal tissue (Figure 4).

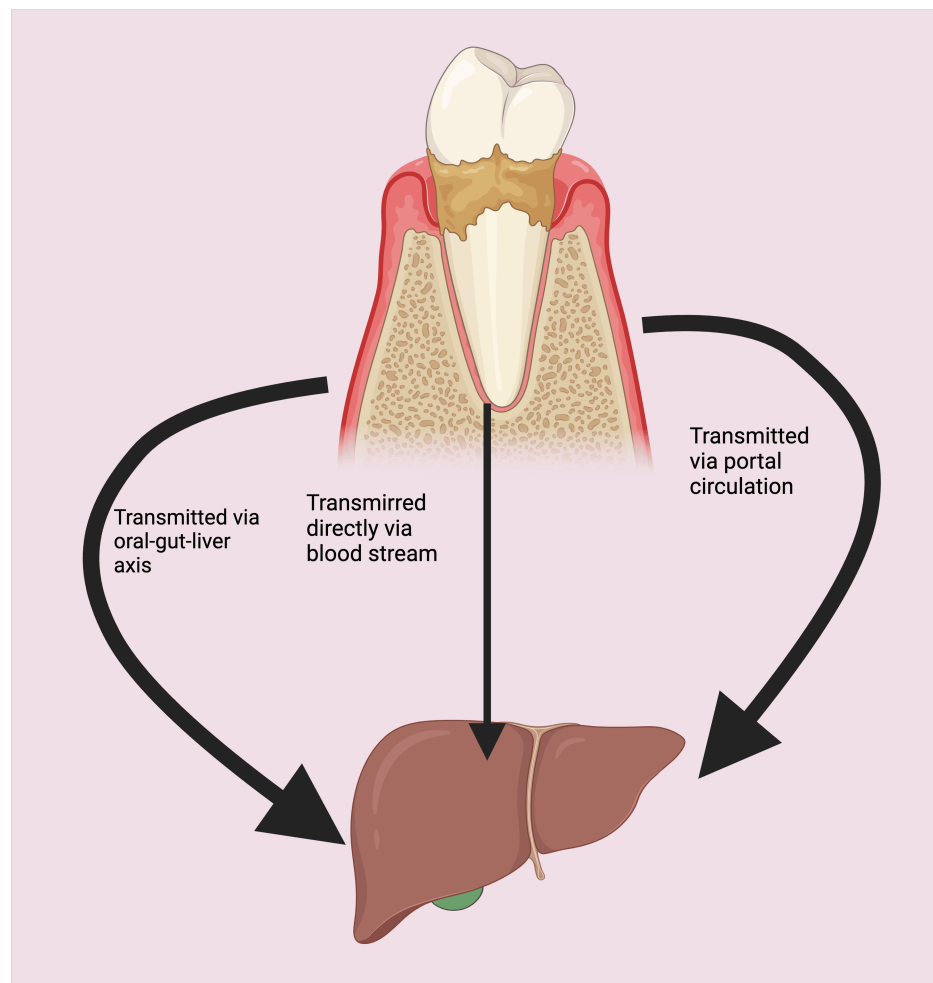


FIGURE 4: Pathways showing the connection of oral microbiota with the liver

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It has been determined that oral and intestinal dysbiosis are considerable risk factors for chronic liver disorders. Subsequently, the inflammatory mediators promote adverse impacts on the liver and bring forward an essential clinical and public health issue [91]. The earliest medical and surgical intervention of chronic periodontal diseases can potentially decrease endotoxin levels and enhance the clinical outcome of NAFLD [44].

Patients with NAFLD had considerably higher rates of *P. gingivalis* in their saliva (detected through polymerase chain reaction (PCR)) and hepatic tissue (determined through biopsy and histopathology) than in non-NAFLD healthy subjects [103,135,136]. Multiple studies similarly reported that a higher number of *P. gingivalis* consider magnetic resonance elastography (MRE)-appraised hepatic tautness [41,120,136,137]. MRE-determined hepatic stiffness was observed more among NAFLD patients with higher levels of *P. gingivalis* FDC381 and SU63 antibody levels [120,138]. Multiple research projects reported that periodontal pockets of 10 or more and a depth of 4 mm or more are considerably associated with hepatic stiffness [103,120].

However, *P. gingivalis* is not the solitary contributor to NAFLD pathogenesis [139]. *Selenomonas noxia* and *Streptococcus oralis* serum antibodies possess a substantial relationship with the development of severe periodontal fatty liver disease [82,139]. *Tannerella forsythia* and *Treponema denticola* antibody titers possess a strong relationship in causing severe periodontitis, nevertheless having a weak association with the advancement of hepatic steatosis [139]. *T. denticola* was notably more frequently detected in NAFLD patients compared to the frequency observed in control subjects [135].

Aggregatibacter actinomycetemcomitans is often called “a tooth killer” [140]. *A. actinomycetemcomitans* is another pathogen that can persuade gastrointestinal microbiome commotion and disrupt glucose metabolism [141,142]. This pathogen synthesizes both endotoxin and exotoxin to trigger an inflammatory response by collaborating with the TLR4 [143-145]. Various research demonstrates that a high-fat diet is linked to dysbiosis in the gastrointestinal microbiome of humans [146,147]. Genus *Turicibacter* of *Firmicutes* phylum is a usual microbiota component of the human gastrointestinal tract as normal flora [148,149]. It is involved in host lipids, bile acids, and steroid metabolism [150,151]. Its richness is inverse as microbiota in the human intestine are strongly associated with inflammatory bowel diseases, e.g., Crohn’s disease [151] and tryptophane/serotonin metabolism [152]. *Turicibacter* has been reported to increase butyric acid synthesis (a short-chain fatty acid) in human cases [153,154]. An increase in butyric acid is associated with improved insulin secretion and sensitivity and possesses substantial anti-obesity and anti-inflammatory effects [154-157]. It has been reported that oral periodontal pathogen *A. actinomycetemcomitans* causes dysbiosis of the gut microbiota that includes the genus *Turicibacter* [141,158,159]. *A. actinomycetemcomitans* may modify the gut microbiota to influence IR but reduce butyric acid production (Figure 5). Multiple studies have examined the impact of treating *A. actinomycetemcomitans* with antibiotics on the gut microbiota and glucose metabolism, resulting in enhanced glucose tolerance, reduced liver lipid accumulation, and alterations in the gut microbiota [160-163].

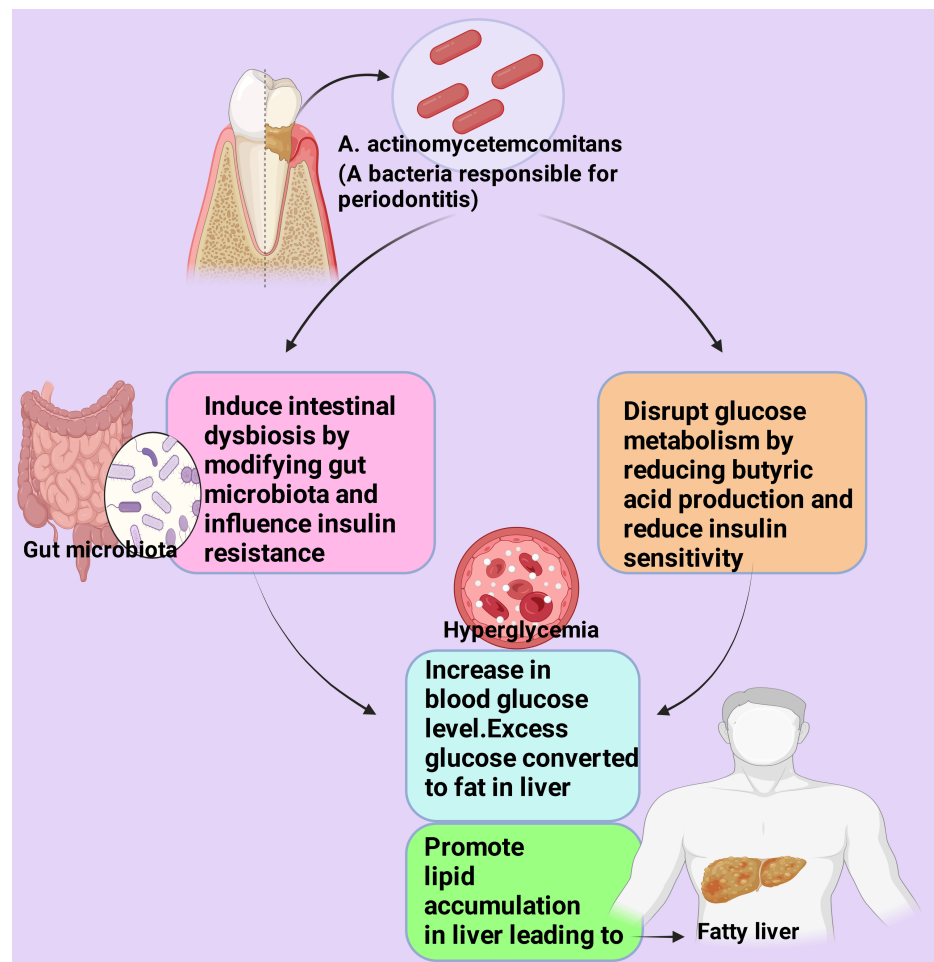


FIGURE 5: The mechanism by which *A. actinomycetemcomitans* causes fatty liver

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Patients with periodontitis exhibited elevated CRP levels [164], whereas those with gingivitis demonstrated elevated gamma-glutamyl transferase (GGT) levels [83]. GGT is a hepatic plasma-membrane-bound enzyme primarily produced in liver cells. It is also synthesized or found on the plasma membranes of almost all vital organ epithelial tissues with secretory or absorbing functions and cells [165-168]. The principal physiological function of GGT is the extracellular synthesis of the thiol antioxidant, glutathione [169].

Glutathione shields cells antagonistic toward oxidants constructed during regular metabolic processes [170]. GGT, therefore, plays an essential role in cellular defense. Whitfield [164] reported that GGT is a conventional biomarker of hepatic diseases, bile duct disorders, alcohol drinking-related hepatic illness, T2DM, coronary heart disease, and stroke [171]. Additionally, raised GGT was also observed in heart failure, hyperthyroidism, MetS, and pancreatitis [172-175].

Alcohol consumption is known to reduce GGT production [171,176]. Explicitly, for every one-unit escalation in “GGT/high-density lipoprotein cholesterol (HDL-C) ratio, the prevalence of NAFLD will increase by 0.3%. As GGT/HDL-C ratio quartiles increased, the prevalence of NAFLD/MetS in Q4 (highest GGT/HDL-C ratio quartile) was 6.362/3.968 times higher than that in Q1 (lowest GGT/HDL-C ratio quartile)” [177]. Individuals with chronic periodontitis exhibit elevated protein carbonyl (PC) levels in serum and the gingival crevicular fluid (GCF) [178]. Chronic severe periodontitis could potentially result in heightened inflammatory biomarkers including GGT levels in the GCF [179,180]. Furthermore, high periodontal inflamed surface area (PISA) had raised GGT levels. PISA is currently considered a new marker for chronic destructive periodontal disease, hepatic disorders, and related systemic illness [83,181,182]. It has been reported that therapeutic intervention, e.g., scaling and root planning (SRP), considerably cut down hepatic enzyme (AST, ALT) and endotoxin among cases of chronic destructive periodontal disease and NAFLD [103,138,183,184]. SRP’s primary goals are removing biofilm calculus and “to create a biologically compatible root surface and reduce the inflammatory burden” [185]. SRP treatment suggestively minimized liver fat content (LFC). Endotoxin and liver enzyme levels in patients with NAFLD and periodontal disease were mainly well-accepted in these patients [138]. One pharmacological agent, lubiprostone, a laxative, minimizes hepatic enzymes in patients with NAFLD and constipation by blocking endotoxin leakage [186,187]. Lubiprostone (the CIC-2 chloride channel activator) is well-accepted among patients. It reduces intestinal penetrability by synthesizing additional colonic mucus through its prostaglandin-like properties on the intestine and improves repairing tight junction proteins (TJPs) of the intestine [187,188]. Lubiprostone alters lipid metabolism and improves NAFLD [187].

Multiple Japanese studies revealed a substantial association between nonalcohol drinkers, the presence of probing pocket depths of 4 mm or greater, and higher serum ALT levels [77,120,189]. Ahmad et al. also demonstrated that the coexistence of MetS and elevated serum ALT was positively associated with pocket depth in adult males with minimal alcohol consumption [190].

Multiple research studies revealed that timely medical and surgical intervention regarding chronic periodontal diseases improves serum lipid profiles toward positive, controls blood glucose, increases insulin sensitivity, decreases IR, decreases inflammatory markers, including endotoxin, and restores overall microbiota [128,138,191-194]. Thus, chronic periodontitis and NAFLD cases are improved. Figure 6 reveals the principal findings of this narrative review.

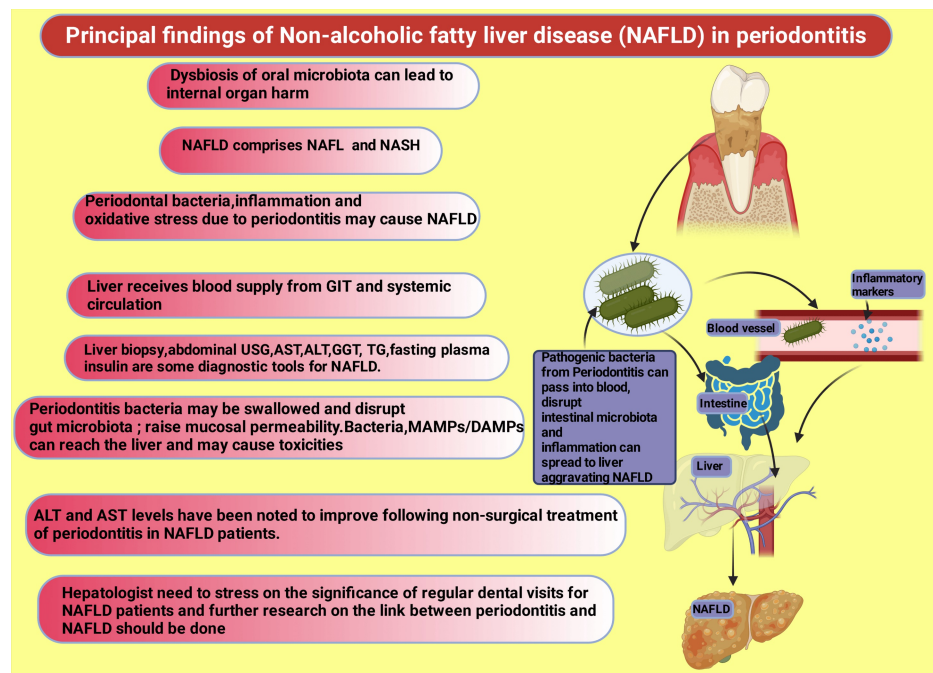


FIGURE 6: Principal findings of nonalcoholic fatty liver disease in periodontitis

ALT: Alanine aminotransferase; AST: aspartate aminotransferase; GGT: gamma-glutamyl transferase; MAMP: microbe-associated molecular patterns; DAMPs: damage-associated molecular pattern; *P. gingivalis*: *Porphyromonas gingivalis*; *T. denticola*: *Treponema denticola*; *T. forsythia*: *Tannerella forsythia*; *A. actinomycetemcomitans*: *Aggregatibacter actinomycetemcomitans*

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Limitations of this paper

This review's drawback was its exclusive dependence on the previously published documents from explorations of “computerized databases, hand searches, and authoritative texts” [195]. This is a narrative review; therefore, no patient data was collected. Articles only written in English were included. No attempt was made to conduct a systematic review and meta-analysis. Narrative reviews have built-in confines in footings of neutrality, comprehensiveness of literature exploration, and clarification of findings [195-199]. Narrative reviews cannot comprise based on science for dedicated issues, nor do they propose conclusive assertions to develop guidelines [196]. In opposing earlier findings, Greenhalgh et al. reported that “narrative reviews provide interpretation and critique; their key contribution is deepening understanding” [200]. Henceforth, the narrative review is considered a different way of disseminating science, knowledge, wisdom, sagacity, and many other issues; nevertheless, it is not an unfortunate colleague of the systematic review [201,202].

Future recommendations regarding management of NAFLD

Extensive research is needed to elucidate the pathophysiological mechanisms linking NAFLD and periodontitis. Understanding these connections on a molecular and systemic level is crucial for developing targeted interventions. We should focus on creating effective pharmacological treatments that can interrupt the various pathways through which NAFLD and periodontitis interact. This includes identifying specific biomarkers and therapeutic targets. Healthcare professionals must spearhead policy and public health initiatives. We need to develop and enforce policies that educate healthcare professionals and the public about the profound impact of poor oral health on overall health, particularly its role in exacerbating conditions like NAFLD.

Public awareness campaigns should be launched to raise awareness about the importance of oral hygiene. These campaigns should emphasize the connection between oral health and systemic conditions, including liver diseases like NAFLD. Integrate oral health education into existing health promotion programs. Educate the public on effective oral hygiene practices, the significance of regular dental check-ups, and the broader

health implications of maintaining good oral health.

Encourage a collaborative approach between dental and medical healthcare providers to manage and prevent NAFLD and related conditions. This includes regular screenings for periodontal disease in patients with NAFLD and vice versa. Implement routine oral health assessments as part of the standard care protocol for patients at risk of or diagnosed with NAFLD. Early detection and management of periodontal disease can mitigate its impact on liver health. By advancing research, promoting public awareness, and integrating healthcare services, we can develop a more comprehensive strategy to manage and potentially reduce the incidence of NAFLD and its associated complications.

Conclusions

Growing evidence from scholastic scientific works exhibits that there is a correlation between NAFLD and periodontitis. There was a noteworthy and statistically significant drop in blood albumin levels when comparing NASH/NAFLD individuals who tested positive for *P. gingivalis* to those who did not. These findings suggest that a decline in liver function could progress more rapidly in patients positive for *P. gingivalis*. Considering these findings, chronic inflammation, IR associated with obesity, and changes brought on by periodontal disease in the gut microbiota might contribute to the development of NAFLD. Other risk factors for NAFLD development include genetic tendencies, gender, geriatric population, race, ethnicity, metabolic syndrome, oxidative stress, physical activity, sleep quality, and nutritional status. Therefore, this study concludes that hepatologists should remind patients with NAFLD of the significance of routine dental visits. As new findings suggest a potential connection between NAFLD and periodontitis, forthcoming research is crucial to ascertain the strength of these links and understand the biological pathways that interconnect these conditions. Additionally, there's a possibility of a two-way relationship where NAFLD could impact periodontitis outcomes, necessitating further exploration.

Additional Information

Author Contributions

All authors have reviewed the final version to be published and agreed to be accountable for all aspects of the work.

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References

1. Lasserre JF, Brex MC, Toma S: Oral microbes, biofilms and their role in periodontal and peri-implant diseases. *Materials (Basel)*. 2018, 11:1802. [10.3390/ma11101802](https://doi.org/10.3390/ma11101802)
2. Abdulkareem AA, Al-Taweel FB, Al-Sharqi AJ, Gul SS, Sha A, Chapple IL: Current concepts in the pathogenesis of periodontitis: from symbiosis to dysbiosis. *J Oral Microbiol*. 2023, 15:2197779. [10.1080/20002297.2023.2197779](https://doi.org/10.1080/20002297.2023.2197779)
3. Bui FQ, Almeida-da-Silva CL, Huynh B, et al.: Association between periodontal pathogens and systemic disease. *Biomed J*. 2019, 42:27-35. [10.1016/j.bj.2018.12.001](https://doi.org/10.1016/j.bj.2018.12.001)
4. Păunică I, Giurgiu M, Dumitriu AS, Păunică S, Pantea Stoian AM, Martu MA, Serafinceanu C: The bidirectional relationship between periodontal disease and diabetes mellitus-a review. *Diagnostics (Basel)*. 2023, 13:681. [10.3390/diagnostics13040681](https://doi.org/10.3390/diagnostics13040681)

5. Hung M, Kelly R, Mohajeri A, et al.: Factors associated with periodontitis in younger individuals: a scoping review. *J Clin Med.* 2023, 12:6442. [10.3390/jcm12206442](#)
6. Lenk M, Noack B, Weidner K, Lorenz K: Psychopathologies and socioeconomic status as risk indicators for periodontitis: a survey-based investigation in German dental practices. *Clin Oral Investig.* 2022, 26:2853-62. [10.1007/s00784-021-04263-2](#)
7. Usui M, Onizuka S, Sato T, Kokabu S, Ariyoshi W, Nakashima K: Mechanism of alveolar bone destruction in periodontitis-periodontal bacteria and inflammation. *Jpn Dent Sci Rev.* 2021, 57:201-8. [10.1016/j.jdsr.2021.09.005](#)
8. Rana H, Warnes B, Davies M, West NX: Patient-reported understanding and dentist-reported management of periodontal diseases-a survey: do you know what gum disease is?. *Br Dent J.* 2023, 235:127-51. [10.1038/s41415-023-6055-7](#)
9. Sanz M, Marco Del Castillo A, Jepsen S, et al.: Periodontitis and cardiovascular diseases: consensus report. *J Clin Periodontol.* 2020, 47:268-88. [10.1111/jcpe.13189](#)
10. Cugini C, Ramasubbu N, Tsiagbe VK, Fine DH: Dysbiosis from a microbial and host perspective relative to oral health and disease. *Front Microbiol.* 2021, 12:617485. [10.3389/fmicb.2021.617485](#)
11. Chandra H, Sharma KK, Tuovinen OH, Sun X, Shukla P: Pathobionts: mechanisms of survival, expansion, and interaction with host with a focus on *Clostridioides difficile*. *Gut Microbes.* 2021, 13:1979882. [10.1080/19490976.2021.1979882](#)
12. Scannapieco FA, Dongari-Bagtzoglou A: Dysbiosis revisited: understanding the role of the oral microbiome in the pathogenesis of gingivitis and periodontitis: a critical assessment. *J Periodontol.* 2021, 92:1071-8. [10.1002/JPER.21-0120](#)
13. Jochum L, Stecher B: Label or concept-what is a pathobiont?. *Trends Microbiol.* 2020, 28:789-92. [10.1016/j.tim.2020.04.011](#)
14. Martínez M, Postolache TT, García-Bueno B, Leza JC, Figuero E, Lowry CA, Malan-Müller S: The role of the oral microbiota related to periodontal diseases in anxiety, mood and trauma- and stress-related disorders. *Front Psychiatry.* 2021, 12:814177. [10.3389/fpsy.2021.814177](#)
15. Santacrose L, Passarelli PC, Azzolino D, et al.: Oral microbiota in human health and disease: a perspective. *Exp Biol Med (Maywood).* 2023, 248:1288-301. [10.1177/15353702231187645](#)
16. Khor B, Snow M, Herrman E, et al.: Interconnections between the oral and gut microbiomes: reversal of microbial dysbiosis and the balance between systemic health and disease. *Microorganisms.* 2021, 9:496. [10.3390/microorganisms9030496](#)
17. Lu Y, Li Z, Peng X: Regulatory effects of oral microbe on intestinal microbiota and the illness. *Front Cell Infect Microbiol.* 2023, 13:1093967. [10.3389/fcimb.2023.1093967](#)
18. Schamarek I, Anders L, Chakaroun RM, Kovacs P, Rohde-Zimmermann K: The role of the oral microbiome in obesity and metabolic disease: potential systemic implications and effects on taste perception. *Nutr J.* 2023, 22:28. [10.1186/s12937-023-00856-7](#)
19. Kuraji R, Ye C, Zhao C, et al.: Nisin lantibiotic prevents NAFLD liver steatosis and mitochondrial oxidative stress following periodontal disease by abrogating oral, gut and liver dysbiosis. *NPJ Biofilms Microbiomes.* 2024, 10:3. [10.1038/s41522-024-00476-x](#)
20. Dewhirst FE, Chen T, Izard J, et al.: The human oral microbiome. *J Bacteriol.* 2010, 192: [10.1128/JB.00542-10](#)
21. Aas JA, Paster BJ, Stokes LN, Olsen I, Dewhirst FE: Defining the normal bacterial flora of the oral cavity. *J Clin Microbiol.* 2005, 43:5721-32. [10.1128/JCM.43.11.5721-5732.2005](#)
22. Alghamdi S: Isolation and identification of the oral bacteria and their characterization for bacteriocin production in the oral cavity. *Saudi J Biol Sci.* 2022, 29:318-23. [10.1016/j.sjbs.2021.08.096](#)
23. Amaroli A, Ravera S, Zekiy A, Benedicenti S, Pasquale C: A narrative review on oral and periodontal bacteria microbiota photobiomodulation, through visible and near-infrared light: from the origins to modern therapies. *Int J Mol Sci.* 2022, 23:1372. [10.3390/ijms23031372](#)
24. Caselli E, Fabbri C, D'Accolti M, Soffritti I, Bassi C, Mazzacane S, Franchi M: Defining the oral microbiome by whole-genome sequencing and resistome analysis: the complexity of the healthy picture. *BMC Microbiol.* 2020, 20:120. [10.1186/s12866-020-01801-y](#)
25. Marsh PD: Role of the oral microflora in health. *Microb Ecol Health Dis.* 2000, 12:130-7. [10.1080/089106000750051800](#)
26. Nagakubo D, Kaibori Y: Oral microbiota: the influences and interactions of saliva, IgA, and dietary factors in health and disease. *Microorganisms.* 2023, 11:2307. [10.3390/microorganisms11092307](#)
27. Radaic A, Kapila YL: The oralome and its dysbiosis: new insights into oral microbiome-host interactions. *Comput Struct Biotechnol J.* 2021, 19:1335-60. [10.1016/j.csbj.2021.02.010](#)
28. Deo PN, Deshmukh R: Oral microbiome: unveiling the fundamentals. *J Oral Maxillofac Pathol.* 2019, 23:122-8. [10.4103/jomfp.JOMFP_304_18](#)
29. Benn A, Heng N, Broadbent JM, Thomson WM: Studying the human oral microbiome: challenges and the evolution of solutions. *Aust Dent J.* 2018, 63:14-24. [10.1111/adj.12565](#)
30. Miller WD: The human mouth as a focus of infection. *Lancet.* 1891, 138:340-342. [10.1016/S0140-6736\(02\)01387-9](#)
31. Miller WD: The micro-organisms of the human mouth: the local and general diseases which are caused by them. Karger, Basel, Switzerland; 1890.
32. Billings F: Chronic focal infection as a causative factor in chronic arthritis. *JAMA.* 1913, 61:819-22. [10.1001/jama.1913.04350110001001](#)
33. Elmaghrawy K, Hussey S, Moran GP: The oral microbiome in pediatric IBD: a source of pathobionts or biomarkers?. *Front Pediatr.* 2020, 8:620254. [10.3389/fped.2020.620254](#)
34. Cai L, Zhu H, Mou Q, et al.: Integrative analysis reveals associations between oral microbiota dysbiosis and host genetic and epigenetic aberrations in oral cavity squamous cell carcinoma. *NPJ Biofilms Microbiomes.* 2024, 10:39. [10.1038/s41522-024-00511-x](#)
35. Sarkar P, Malik S, Laha S, et al.: Dysbiosis of oral microbiota during oral squamous cell carcinoma development. *Front Oncol.* 2021, 11:614448. [10.3389/fonc.2021.614448](#)
36. de Lemos GM, Resende CM, Campello CP, et al.: Is oral microbiota associated with overweight and obesity

- in children and adolescents? A systematic review. *Crit Rev Food Sci Nutr.* 2024, 64:4275-85. [10.1080/10408398.2022.2140330](https://doi.org/10.1080/10408398.2022.2140330)
37. Cheng Z, Do T, Mankia K, et al.: Dysbiosis in the oral microbiomes of anti-CCP positive individuals at risk of developing rheumatoid arthritis. *Ann Rheum Dis.* 2021, 80:162-8. [10.1136/annrheumdis-2020-216972](https://doi.org/10.1136/annrheumdis-2020-216972)
 38. Bellando-Randone S, Russo E, Venerito V, Matucci-Cerinic M, Iannone F, Tangaro S, Amedei A: Exploring the oral microbiome in rheumatic diseases, state of art and future prospective in personalized medicine with an AI approach. *J Pers Med.* 2021, 11:625. [10.3390/jpm11070625](https://doi.org/10.3390/jpm11070625)
 39. Di Stefano M, Polizzi A, Santonocito S, Romano A, Lombardi T, Isola G: Impact of oral microbiome in periodontal health and periodontitis: a critical review on prevention and treatment. *Int J Mol Sci.* 2022, 23:5142. [10.3390/ijms23095142](https://doi.org/10.3390/ijms23095142)
 40. Pietiäinen M, Liljestrand JM, Kopra E, Pussinen PJ: Mediators between oral dysbiosis and cardiovascular diseases. *Eur J Oral Sci.* 2018, 126:26-36. [10.1111/eos.12423](https://doi.org/10.1111/eos.12423)
 41. Li Y, Zhu M, Liu Y, et al.: The oral microbiota and cardiometabolic health: a comprehensive review and emerging insights. *Front Immunol.* 2022, 13:1010368. [10.3389/fimmu.2022.1010368](https://doi.org/10.3389/fimmu.2022.1010368)
 42. Yin C, Chen J, Wu X, et al.: Preterm birth is correlated with increased oral originated microbiome in the gut. *Front Cell Infect Microbiol.* 2021, 11:579766. [10.3389/fcimb.2021.579766](https://doi.org/10.3389/fcimb.2021.579766)
 43. Wan J, Fan H: Oral microbiome and Alzheimer's disease. *Microorganisms.* 2023, 11:2550. [10.3390/microorganisms11102550](https://doi.org/10.3390/microorganisms11102550)
 44. Peng X, Cheng L, You Y, et al.: Oral microbiota in human systematic diseases. *Int J Oral Sci.* 2022, 14:14. [10.1038/s41368-022-00163-7](https://doi.org/10.1038/s41368-022-00163-7)
 45. Thomas C, Minty M, Vinel A, et al.: Oral microbiota: a major player in the diagnosis of systemic diseases. *Diagnostics (Basel).* 2021, 11:1376. [10.3390/diagnostics11081376](https://doi.org/10.3390/diagnostics11081376)
 46. Li Q, Wang H, Tan L, Zhang S, Lin L, Tang X, Pan Y: Oral pathogen *Fusobacterium nucleatum* coaggregates with *Pseudomonas aeruginosa* to modulate the inflammatory cytotoxicity of pulmonary epithelial cells. *Front Cell Infect Microbiol.* 2021, 11:643913. [10.3389/fcimb.2021.643913](https://doi.org/10.3389/fcimb.2021.643913)
 47. Boccia G, Di Spirito F, D'Ambrosio F, Di Palo MP, Giordano F, Amato M: Local and systemic antibiotics in peri-implantitis management: an umbrella review. *Antibiotics (Basel).* 2023, 12:114. [10.3390/antibiotics12010114](https://doi.org/10.3390/antibiotics12010114)
 48. Teng ML, Ng CH, Huang DQ, et al.: Global incidence and prevalence of nonalcoholic fatty liver disease. *Clin Mol Hepatol.* 2023, 29:S32-42. [10.3350/cmh.2022.0365](https://doi.org/10.3350/cmh.2022.0365)
 49. Han SK, Baik SK, Kim MY: Non-alcoholic fatty liver disease: definition and subtypes. *Clin Mol Hepatol.* 2023, 29:S5-16. [10.3350/cmh.2022.0424](https://doi.org/10.3350/cmh.2022.0424)
 50. Schaffner F, Thaler H: Nonalcoholic fatty liver disease. *Prog Liver Dis.* 1986, 8:283-98.
 51. Ludwig J, Viggiano TR, McGill DB, Oh BJ: Nonalcoholic steatohepatitis: Mayo Clinic experiences with a hitherto unnamed disease. *Mayo Clin Proc.* 1980, 55:434-8.
 52. Feng Y, Sun W, Sun F, et al.: Biological mechanisms and related natural inhibitors of CD36 in nonalcoholic fatty liver. *Drug Des Devel Ther.* 2022, 16:3829-45. [10.2147/DDDT.S386982](https://doi.org/10.2147/DDDT.S386982)
 53. Bessone F, Razori MV, Roma MG: Molecular pathways of nonalcoholic fatty liver disease development and progression. *Cell Mol Life Sci.* 2019, 76:99-128. [10.1007/s00018-018-2947-0](https://doi.org/10.1007/s00018-018-2947-0)
 54. Machado MV, Diehl AM: Pathogenesis of nonalcoholic steatohepatitis. *Gastroenterology.* 2016, 150:1769-77. [10.1053/j.gastro.2016.02.066](https://doi.org/10.1053/j.gastro.2016.02.066)
 55. Suppli MP, Rigbolt KT, Veidal SS, et al.: Hepatic transcriptome signatures in patients with varying degrees of nonalcoholic fatty liver disease compared with healthy normal-weight individuals. *Am J Physiol Gastrointest Liver Physiol.* 2019, 316:G462-72. [10.1152/ajpgi.00358.2018](https://doi.org/10.1152/ajpgi.00358.2018)
 56. Eslam M, Newsome PN, Sarin SK, et al.: A new definition for metabolic dysfunction-associated fatty liver disease: an international expert consensus statement. *J Hepatol.* 2020, 73:202-9. [10.1016/j.jhep.2020.03.039](https://doi.org/10.1016/j.jhep.2020.03.039)
 57. Zhang XL, Fan JG, Wei L, Shi JP, Zheng MH; Chinese MAFLD Clinical Research Network: Promoting the term MAFLD: China in action. *Lancet Gastroenterol Hepatol.* 2022, 7:598. [10.1016/S2468-1253\(22\)00127-3](https://doi.org/10.1016/S2468-1253(22)00127-3)
 58. Adams LA, Roberts SK, Strasser SI, et al.: Nonalcoholic fatty liver disease burden: Australia, 2019-2030. *J Gastroenterol Hepatol.* 2020, 35:1628-35. [10.1111/jgh.15009](https://doi.org/10.1111/jgh.15009)
 59. Aguiar IL, Santos-Lins LS, Brasil-Oliveira R, Cotrim HP, Lins-Kusterer L: Non-alcoholic fatty liver disease and periodontal disease: a systematic review and meta-analysis of cross-sectional studies. *Arab J Gastroenterol.* 2023, 24:198-203. [10.1016/j.ajg.2023.09.005](https://doi.org/10.1016/j.ajg.2023.09.005)
 60. Chen Y, Yang YC, Zhu BL, Wu CC, Lin RF, Zhang X: Association between periodontal disease, tooth loss and liver diseases risk. *J Clin Periodontol.* 2020, 47:1053-63. [10.1111/jcpe.13341](https://doi.org/10.1111/jcpe.13341)
 61. Zhang J, Wang L, Jiang M: Diagnostic value of sphingolipid metabolism-related genes CD37 and CXCL9 in nonalcoholic fatty liver disease. *Medicine (Baltimore).* 2024, 103:e37185. [10.1097/MD.00000000000037185](https://doi.org/10.1097/MD.00000000000037185)
 62. BioRender. (2024). Accessed: June 20, 2024: <https://www.biorender.com/>.
 63. MacParland SA, Liu JC, Ma XZ, et al.: Single cell RNA sequencing of human liver reveals distinct intrahepatic macrophage populations. *Nat Commun.* 2018, 9:4383. [10.1038/s41467-018-06318-7](https://doi.org/10.1038/s41467-018-06318-7)
 64. Bechmann LP, Hannivoort RA, Gerken G, Hotamisligil GS, Trauner M, Canbay A: The interaction of hepatic lipid and glucose metabolism in liver diseases. *J Hepatol.* 2012, 56:952-64. [10.1016/j.jhep.2011.08.025](https://doi.org/10.1016/j.jhep.2011.08.025)
 65. Grant DM: Detoxification pathways in the liver. *J Inher Metab Dis.* 1991, 14:421-30. [10.1007/BF01797915](https://doi.org/10.1007/BF01797915)
 66. Trefts E, Gannon M, Wasserman DH: The liver. *Curr Biol.* 2017, 27:R1147-51. [10.1016/j.cub.2017.09.019](https://doi.org/10.1016/j.cub.2017.09.019)
 67. Chaudhry S, Emond J, Griesemer A: Immune cell trafficking to the liver. *Transplantation.* 2019, 103:1323-37. [10.1097/TP.0000000000002690](https://doi.org/10.1097/TP.0000000000002690)
 68. Hastings KL, Green MD, Gao B, Ganey PE, Roth RA, Burleson GR: Beyond metabolism: role of the immune system in hepatic toxicity. *Int J Toxicol.* 2020, 39:151-64. [10.1177/1091581819898399](https://doi.org/10.1177/1091581819898399)
 69. Zhou Z, Xu MJ, Gao B: Hepatocytes: a key cell type for innate immunity. *Cell Mol Immunol.* 2016, 13:301-15. [10.1038/cmi.2015.97](https://doi.org/10.1038/cmi.2015.97)
 70. Ortiz C, Schierwagen R, Schaefer L, Klein S, Treppe X, Trebicka J: Extracellular matrix remodeling in chronic liver disease. *Curr Tissue Microenviron Rep.* 2021, 2:41-52. [10.1007/s43152-021-00030-3](https://doi.org/10.1007/s43152-021-00030-3)
 71. Baiocchi A, Montaldo C, Conigliaro A, et al.: Extracellular matrix molecular remodeling in human liver fibrosis evolution. *PLoS One.* 2016, 11:e0151736. [10.1371/journal.pone.0151736](https://doi.org/10.1371/journal.pone.0151736)

72. Nguyen-Lefebvre AT, Horuzsko A: Kupffer cell metabolism and function . *J Enzymol Metab.* 2015, 1:
73. Woltman AM, Boonstra A, Naito M, Leenen PJ: Kupffer cells in health and disease . *Macrophages: Biology and Role in the Pathology of Diseases.* Biswas S, Mantovani A (ed): Springer, New York (NY); 2014. 217-47. [10.1007/978-1-4939-1311-4_10](https://doi.org/10.1007/978-1-4939-1311-4_10)
74. Yuan F, Zhang W, Mu D, Gong J: Kupffer cells in immune activation and tolerance toward HBV/HCV infection. *Adv Clin Exp Med.* 2017, 26:739-45. [10.17219/acem/62759](https://doi.org/10.17219/acem/62759)
75. Hora S, Wuestefeld T: Liver injury and regeneration: current understanding, new approaches, and future perspectives. *Cells.* 2023, 12:2129. [10.3390/cells12172129](https://doi.org/10.3390/cells12172129)
76. Dyson JK, Anstee QM, McPherson S: Non-alcoholic fatty liver disease: a practical approach to diagnosis and staging. *Frontline Gastroenterol.* 2014, 5:211-8. [10.1136/flgastro-2013-100403](https://doi.org/10.1136/flgastro-2013-100403)
77. Iwasaki T, Hirose A, Azuma T, et al.: Correlation between ultrasound-diagnosed non-alcoholic fatty liver and periodontal condition in a cross-sectional study in Japan. *Sci Rep.* 2018, 8:7496. [10.1038/s41598-018-25857-z](https://doi.org/10.1038/s41598-018-25857-z)
78. Mahale AR, Prabhu SD, Nachiappan M, Fernandes M, Ullal S: Clinical relevance of reporting fatty liver on ultrasound in asymptomatic patients during routine health checkups. *J Int Med Res.* 2018, 46:4447-54. [10.1177/0300060518793039](https://doi.org/10.1177/0300060518793039)
79. Tantanavipas S, Vallibhakara O, Sobhonslidsuk A, Phongkitkarun S, Vallibhakara SA, Promson K, Sophonsritsuk A: Abdominal obesity as a predictive factor of nonalcoholic fatty liver disease assessed by ultrasonography and transient elastography in polycystic ovary syndrome and healthy women. *Biomed Res Int.* 2019, 2019:9047324. [10.1155/2019/9047324](https://doi.org/10.1155/2019/9047324)
80. Takahashi Y, Fukusato T: Histopathology of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis. *World J Gastroenterol.* 2014, 20:15539-48. [10.3748/wjg.v20.i42.15539](https://doi.org/10.3748/wjg.v20.i42.15539)
81. Brown GT, Kleiner DE: Histopathology of nonalcoholic fatty liver disease and nonalcoholic steatohepatitis . *Metabolism.* 2016, 65:1080-6. [10.1016/j.metabol.2015.11.008](https://doi.org/10.1016/j.metabol.2015.11.008)
82. Kuraji R, Sekino S, Kapila Y, Numabe Y: Periodontal disease-related nonalcoholic fatty liver disease and nonalcoholic steatohepatitis: an emerging concept of oral-liver axis. *Periodontol 2000.* 2021, 87:204-40. [10.1111/prd.12387](https://doi.org/10.1111/prd.12387)
83. Fujii T, Aoyama N, Kida S, Taniguchi K, Yata T, Minabe M, Komaki M: Associations between periodontal status and liver function in the Japanese population: a cross-sectional study. *J Clin Med.* 2023, 12:4759. [10.3390/jcm12144759](https://doi.org/10.3390/jcm12144759)
84. Jennison E, Patel J, Scorletti E, Byrne CD: Diagnosis and management of non-alcoholic fatty liver disease . *Postgrad Med J.* 2019, 95:314-22. [10.1136/postgradmedj-2018-136316](https://doi.org/10.1136/postgradmedj-2018-136316)
85. Hall P, Cash J: What is the real function of the liver 'function' tests? . *Ulster Med J.* 2012, 81:30-6.
86. Kwo PY, Cohen SM, Lim JK: ACG clinical guideline: evaluation of abnormal liver chemistries . *Am J Gastroenterol.* 2017, 112:18-35. [10.1038/ajg.2016.517](https://doi.org/10.1038/ajg.2016.517)
87. Huang XJ, Choi YK, Im HS, Yarimaga O, Yoon E, Kim HS: Aspartate aminotransferase (AST/GOT) and alanine aminotransferase (ALT/GPT) detection techniques. *Sensors (Basel).* 2006, 6:756-82.
88. Moriles KE, Zubair M, Azer SA: Alanine aminotransferase (ALT). *StatPearls Publishing, Treasure Island (FL);* 2024.
89. Huh Y, Cho YJ, Nam GE: Recent epidemiology and risk factors of nonalcoholic fatty liver disease . *J Obes Metab Syndr.* 2022, 31:17-27. [10.7570/jomes22021](https://doi.org/10.7570/jomes22021)
90. Sarkar S, Lipworth L, Kabagambe EK, et al.: A description of risk factors for non-alcoholic fatty liver disease in the southern community cohort study: a nested case-control study. *Front Nutr.* 2020, 7:71. [10.3389/fnut.2020.00071](https://doi.org/10.3389/fnut.2020.00071)
91. Chen TP, Yu HC, Lin WY, Chang YC: The role of microbiome in the pathogenesis of oral-gut-liver axis between periodontitis and nonalcoholic fatty liver disease. *J Dent Sci.* 2023, 18:972-5. [10.1016/j.jds.2023.03.012](https://doi.org/10.1016/j.jds.2023.03.012)
92. Hajishengallis G: Periodontitis: from microbial immune subversion to systemic inflammation . *Nat Rev Immunol.* 2015, 15:30-44. [10.1038/nri3785](https://doi.org/10.1038/nri3785)
93. Elghannam MT, Hassanien MH, Ameen YA, et al.: Oral microbiome dysbiosis and gastrointestinal diseases: a narrative review. *Egypt Liver J.* 2024, 14:32. [10.1186/s43066-024-00340-9](https://doi.org/10.1186/s43066-024-00340-9)
94. Costa CF, Sampaio-Maia B, Araujo R, et al.: Gut microbiome and organ fibrosis . *Nutrients.* 2022, 14:352. [10.3390/nu14020352](https://doi.org/10.3390/nu14020352)
95. Olsen I, Yamazaki K: Can oral bacteria affect the microbiome of the gut? . *J Oral Microbiol.* 2019, 11:1586422. [10.1080/20002297.2019.1586422](https://doi.org/10.1080/20002297.2019.1586422)
96. Lei Y, Li S, He M, Ao Z, Wang J, Wu Q, Wang Q: Oral pathogenic bacteria and the oral-gut-liver axis: a new understanding of chronic liver diseases. *Diagnostics (Basel).* 2023, 13:3324. [10.3390/diagnostics13213324](https://doi.org/10.3390/diagnostics13213324)
97. Korkmaz FT, Traber KE: Innate immune responses in pneumonia . *Pneumonia (Nathan).* 2023, 15:4. [10.1186/s41479-023-00106-8](https://doi.org/10.1186/s41479-023-00106-8)
98. Ramadan DE, Hariyani N, Indrawati R, Ridwan RD, Diyatri I: Cytokines and chemokines in periodontitis . *Eur J Dent.* 2020, 14:483-95. [10.1055/s-0040-1712718](https://doi.org/10.1055/s-0040-1712718)
99. Zhu X, Huang H, Zhao L: PAMPs and DAMPs as the bridge between periodontitis and atherosclerosis: the potential therapeutic targets. *Front Cell Dev Biol.* 2022, 10:856118. [10.3389/fcell.2022.856118](https://doi.org/10.3389/fcell.2022.856118)
100. Brandl K, Kumar V, Eckmann L: Gut-liver axis at the frontier of host-microbial interactions . *Am J Physiol Gastrointest Liver Physiol.* 2017, 312:G413-9. [10.1152/ajpgi.00361.2016](https://doi.org/10.1152/ajpgi.00361.2016)
101. Brandl K, Schnabl B: Is intestinal inflammation linking dysbiosis to gut barrier dysfunction during liver disease?. *Expert Rev Gastroenterol Hepatol.* 2015, 9:1069-76. [10.1586/17474124.2015.1057122](https://doi.org/10.1586/17474124.2015.1057122)
102. Qin N, Yang F, Li A, et al.: Alterations of the human gut microbiome in liver cirrhosis . *Nature.* 2014, 513:59-64. [10.1038/nature13568](https://doi.org/10.1038/nature13568)
103. Kobayashi T, Iwaki M, Nogami A, et al.: Involvement of periodontal disease in the pathogenesis and exacerbation of nonalcoholic fatty liver disease/nonalcoholic steatohepatitis: a review. *Nutrients.* 2023, 15:1269. [10.3390/nu15051269](https://doi.org/10.3390/nu15051269)
104. Park JH, Moon JH, Kim HJ, Kong MH, Oh YH: Sedentary lifestyle: overview of updated evidence of potential health risks. *Korean J Fam Med.* 2020, 41:365-73. [10.4082/kjfm.20.0165](https://doi.org/10.4082/kjfm.20.0165)

105. Oliveira NC, Balikian Júnior P, Júnior AT, et al.: Environmental planning and non-communicable diseases: a systematic review on the role of the metabolomic profile. *Int J Environ Res Public Health*. 2023, 20:6433. [10.3390/ijerph20146433](https://doi.org/10.3390/ijerph20146433)
106. Li Y, Fan X, Wei L, Yang K, Jiao M: The impact of high-risk lifestyle factors on all-cause mortality in the US non-communicable disease population. *BMC Public Health*. 2023, 23:422. [10.1186/s12889-023-15319-1](https://doi.org/10.1186/s12889-023-15319-1)
107. Liu L, Geng Y, Xiong C: Impact of Porphyromonas gingivalis-odontogenic infection on the pathogenesis of non-alcoholic fatty liver disease. *Ann Med*. 2023, 55: [10.1080/07853890.2023.2255825](https://doi.org/10.1080/07853890.2023.2255825)
108. Albuquerque-Souza E, Sahingur SE: Periodontitis, chronic liver diseases, and the emerging oral-gut-liver axis. *Periodontol 2000*. 2022, 89:125-41. [10.1111/prd.12427](https://doi.org/10.1111/prd.12427)
109. Sun Y, Yin Y, Yang S, et al.: Lipotoxicity: the missing link between diabetes and periodontitis? . *J Periodontol Res*. 2024, 59:431-45. [10.1111/jre.13242](https://doi.org/10.1111/jre.13242)
110. Weintraub JA, Lopez Mitnik G, Dye BA: Oral diseases associated with nonalcoholic fatty liver disease in the United States. *J Dent Res*. 2019, 98:1219-26. [10.1177/0022034519866442](https://doi.org/10.1177/0022034519866442)
111. Le Chatelier E, Nielsen T, Qin J, et al.: Richness of human gut microbiome correlates with metabolic markers. *Nature*. 2013, 500:541-6. [10.1038/nature12506](https://doi.org/10.1038/nature12506)
112. Kuroe K, Furuta M, Takeuchi K, et al.: Association between periodontitis and fibrotic progression of non-alcoholic fatty liver among Japanese adults. *J Clin Periodontol*. 2021, 48:368-77. [10.1111/jcpe.13415](https://doi.org/10.1111/jcpe.13415)
113. Koning M, Herrema H, Nieuwdorp M, Meijnikman AS: Targeting nonalcoholic fatty liver disease via gut microbiome-centered therapies. *Gut Microbes*. 2023, 15:2226922. [10.1080/19490976.2023.2226922](https://doi.org/10.1080/19490976.2023.2226922)
114. Safari Z, Gérard P: The links between the gut microbiome and non-alcoholic fatty liver disease (NAFLD) . *Cell Mol Life Sci*. 2019, 76:1541-58. [10.1007/s00018-019-03011-w](https://doi.org/10.1007/s00018-019-03011-w)
115. Helenius-Hietala J, Suominen AL, Ruokonen H, et al.: Periodontitis is associated with incident chronic liver disease-a population-based cohort study. *Liver Int*. 2019, 39:583-91. [10.1111/liv.13985](https://doi.org/10.1111/liv.13985)
116. Suzuki N, Yoneda M, Hirofujii T: Mixed red-complex bacterial infection in periodontitis . *Int J Dent*. 2013, 2013:587279. [10.1155/2013/587279](https://doi.org/10.1155/2013/587279)
117. Mohanty R, Asopa SJ, Joseph MD, Singh B, Rajguru JP, Saidath K, Sharma U: Red complex: polymicrobial conglomerate in oral flora: a review. *J Family Med Prim Care*. 2019, 8:3480-6. [10.4103/jfmpc.jfmpc_759_19](https://doi.org/10.4103/jfmpc.jfmpc_759_19)
118. How KY, Song KP, Chan KG: Porphyromonas gingivalis: an overview of periodontopathic pathogen below the gum line. *Front Microbiol*. 2016, 7:53. [10.3389/fmicb.2016.00053](https://doi.org/10.3389/fmicb.2016.00053)
119. Wu L, Shi R, Bai H, Wang X, Wei J, Liu C, Wu Y: Porphyromonas gingivalis induces increases in branched-chain amino acid levels and exacerbates liver injury through livh/livk. *Front Cell Infect Microbiol*. 2022, 12:776996. [10.3389/fcimb.2022.776996](https://doi.org/10.3389/fcimb.2022.776996)
120. Sato S, Kamata Y, Kessoku T, et al.: A cross-sectional study assessing the relationship between non-alcoholic fatty liver disease and periodontal disease. *Sci Rep*. 2022, 12:13621. [10.1038/s41598-022-17917-2](https://doi.org/10.1038/s41598-022-17917-2)
121. Selvaraj EA, Mózes FE, Jayaswal AN, et al.: Diagnostic accuracy of elastography and magnetic resonance imaging in patients with NAFLD: a systematic review and meta-analysis. *J Hepatol*. 2021, 75:770-85. [10.1016/j.jhep.2021.04.044](https://doi.org/10.1016/j.jhep.2021.04.044)
122. de Jongh CA, de Vries TJ, Bikker FJ, Gibbs S, Krom BP: Mechanisms of Porphyromonas gingivalis to translocate over the oral mucosa and other tissue barriers. *J Oral Microbiol*. 2023, 15:2205291. [10.1080/20002297.2023.2205291](https://doi.org/10.1080/20002297.2023.2205291)
123. Olsen I, Singhrao SK: Can oral infection be a risk factor for Alzheimer's disease? . *J Oral Microbiol*. 2015, 7:29143. [10.3402/jom.v7.29143](https://doi.org/10.3402/jom.v7.29143)
124. Yamazaki K: Oral-gut axis as a novel biological mechanism linking periodontal disease and systemic diseases: a review. *Jpn Dent Sci Rev*. 2023, 59:273-80. [10.1016/j.jdsr.2023.08.003](https://doi.org/10.1016/j.jdsr.2023.08.003)
125. Di Vincenzo F, Del Gaudio A, Petito V, Lopetuso LR, Scaldaferri F: Gut microbiota, intestinal permeability, and systemic inflammation: a narrative review. *Intern Emerg Med*. 2024, 19:275-93. [10.1007/s11739-023-03374-w](https://doi.org/10.1007/s11739-023-03374-w)
126. Poto R, Fusco W, Rinninella E, et al.: The role of gut microbiota and leaky gut in the pathogenesis of food allergy. *Nutrients*. 2023, 16:92. [10.3390/nu16010092](https://doi.org/10.3390/nu16010092)
127. Yao C, Lan D, Li X, Wang Y, Qi S, Liu Y: Porphyromonas gingivalis is a risk factor for the development of nonalcoholic fatty liver disease via ferroptosis. *Microbes Infect*. 2023, 25:105040. [10.1016/j.micinf.2022.105040](https://doi.org/10.1016/j.micinf.2022.105040)
128. Rinčić G, Gačina P, Virović Jukić L, Rinčić N, Božić D, Badovinac A: Association between periodontitis and liver disease. *Acta Clin Croat*. 2022, 60:510-8. [10.20471/acc.2021.60.03.22](https://doi.org/10.20471/acc.2021.60.03.22)
129. Martínez-García M, Hernández-Lemus E: Periodontal inflammation and systemic diseases: an overview . *Front Physiol*. 2021, 12:709438. [10.3389/fphys.2021.709438](https://doi.org/10.3389/fphys.2021.709438)
130. Kiryowa HM, Munabi IG, Buwembo W, Rwenyonyi CM, Mwaka ES, Kaddumukasa M: Periodontitis is associated with insulin resistance in adults living with diabetes mellitus in Uganda: a cross- sectional study. *BMC Res Notes*. 2023, 16:217. [10.1186/s13104-023-06473-1](https://doi.org/10.1186/s13104-023-06473-1)
131. Pussinen PJ, Kopra E, Pietiäinen M, Lehto M, Zaric S, Paju S, Salminen A: Periodontitis and cardiometabolic disorders: the role of lipopolysaccharide and endotoxemia. *Periodontol 2000*. 2022, 89:19-40. [10.1111/prd.12433](https://doi.org/10.1111/prd.12433)
132. Xu T, Liu R, Zhu H, Zhou Y, Pei T, Yang Z: The inhibition of LPS-induced oxidative stress and inflammatory responses is associated with the protective effect of (-)-epigallocatechin-3-gallate on bovine hepatocytes and murine liver. *Antioxidants (Basel)*. 2022, 11:914. [10.3390/antiox11050914](https://doi.org/10.3390/antiox11050914)
133. Kalyan M, Tousif AH, Sonali S, et al.: Role of endogenous lipopolysaccharides in neurological disorders . *Cells*. 2022, 11:4038. [10.3390/cells11244038](https://doi.org/10.3390/cells11244038)
134. Zanoni I, Ostuni R, Marek LR, et al.: CD14 controls the LPS-induced endocytosis of Toll-like receptor 4 . *Cell*. 2011, 147:868-80. [10.1016/j.cell.2011.09.051](https://doi.org/10.1016/j.cell.2011.09.051)
135. Yoneda M, Naka S, Nakano K, et al.: Involvement of a periodontal pathogen, Porphyromonas gingivalis on the pathogenesis of non-alcoholic fatty liver disease. *BMC Gastroenterol*. 2012, 12:16. [10.1186/1471-230X-12-16](https://doi.org/10.1186/1471-230X-12-16)
136. Nakahara T, Hyogo H, Ono A, et al.: Involvement of Porphyromonas gingivalis in the progression of non-alcoholic fatty liver disease. *J Gastroenterol*. 2018, 53:269-80. [10.1007/s00535-017-1368-4](https://doi.org/10.1007/s00535-017-1368-4)

137. Taylor GW, Burt BA, Becker MP, Genco RJ, Shlossman M, Knowler WC, Pettitt DJ: Severe periodontitis and risk for poor glycemic control in patients with non-insulin-dependent diabetes mellitus. *J Periodontol.* 1996, 67:1085-93. [10.1902/jop.1996.67.10s.1085](#)
138. Kamata Y, Kessoku T, Shimizu T, et al.: Periodontal treatment and usual care for nonalcoholic fatty liver disease: a multicenter, randomized controlled trial. *Clin Transl Gastroenterol.* 2022, 13:e00520. [10.14309/ctg.0000000000000520](#)
139. Hatasa M, Yoshida S, Takahashi H, et al.: Relationship between NAFLD and periodontal disease from the view of clinical and basic research, and immunological response. *Int J Mol Sci.* 2021, 22:3728. [10.3390/ijms22073728](#)
140. Raja M, Ummer F, Dhivakar CP: Aggregatibacter actinomycetemcomitans-a tooth killer? . *J Clin Diagn Res.* 2014, 8:ZE13-6. [10.7860/JCDR/2014/9845.4766](#)
141. Komazaki R, Katagiri S, Takahashi H, et al.: Periodontal pathogenic bacteria, Aggregatibacter actinomycetemcomitans affect non-alcoholic fatty liver disease by altering gut microbiota and glucose metabolism. *Sci Rep.* 2017, 7:13950. [10.1038/s41598-017-14260-9](#)
142. Wang T, Ishikawa T, Sasaki M, Chiba T: Oral and gut microbial dysbiosis and non-alcoholic fatty liver disease: the central role of Porphyromonas gingivalis. *Front Med (Lausanne).* 2022, 9:822190. [10.3389/fmed.2022.822190](#)
143. Ozuna H, Snider I, Belibasakis GN, Oscarsson J, Johansson A, Uriarte SM: Aggregatibacter actinomycetemcomitans and Filifactor alocis: two exotoxin-producing oral pathogens. *Front Oral Health.* 2022, 3:981343. [10.3389/froh.2022.981343](#)
144. Åberg CH, Kelk P, Johansson A: Aggregatibacter actinomycetemcomitans: virulence of its leukotoxin and association with aggressive periodontitis. *Virulence.* 2015, 6:188-95. [10.4161/21505594.2014.982428](#)
145. Li X, Zhou L, Takai H, et al.: Aggregatibacter actinomycetemcomitans lipopolysaccharide regulates bone sialoprotein gene transcription. *J Cell Biochem.* 2012, 113:2822-34. [10.1002/jcb.24157](#)
146. Kang GG, Trevaskis NL, Murphy AJ, Febbraio MA: Diet-induced gut dysbiosis and inflammation: key drivers of obesity-driven NASH. *iScience.* 2023, 26:105905. [10.1016/j.isci.2022.105905](#)
147. Kociszewska D, Chan J, Thorne PR, Vljakovic SM: The link between gut dysbiosis caused by a high-fat diet and hearing loss. *Int J Mol Sci.* 2021, 22:13177. [10.3390/ijms222413177](#)
148. Auchtung TA, Holder ME, Gesell JR, et al.: Complete genome sequence of Turicibacter sp. strain H121, isolated from the feces of a contaminated germ-free mouse. *Genome Announc.* 2016, 4:e00114-16. [10.1128/genomeA.00114-16](#)
149. Lynch JB, Gonzalez EL, Choy K, et al.: Gut microbiota Turicibacter strains differentially modify bile acids and host lipids. *Nat Commun.* 2023, 14:3669. [10.1038/s41467-023-39403-7](#)
150. Hoffman JM, Margolis KG: Building community in the gut: a role for mucosal serotonin . *Nat Rev Gastroenterol Hepatol.* 2020, 17:6-8. [10.1038/s41575-019-0227-6](#)
151. Bernstein CN, Forbes JD: Gut microbiome in inflammatory bowel disease and other chronic immune-mediated inflammatory diseases. *Inflamm Intest Dis.* 2017, 2:116-23. [10.1159/000481401](#)
152. Renga G, D'Onofrio F, Pariano M, et al.: Bridging of host-microbiota tryptophan partitioning by the serotonin pathway in fungal pneumonia. *Nat Commun.* 2023, 14:5753. [10.1038/s41467-023-41536-8](#)
153. Lin TC, Soorneedi A, Guan Y, Tang Y, Shi E, Moore MD, Liu Z: Turicibacter fermentation enhances the inhibitory effects of Antrodia camphorata supplementation on tumorigenic serotonin and Wnt pathways and promotes ROS-mediated apoptosis of Caco-2 cells. *Front Pharmacol.* 2023, 14:1203087. [10.3389/fphar.2023.1203087](#)
154. Schulthess J, Pandey S, Capitani M, et al.: The short chain fatty acid butyrate imprints an antimicrobial program in macrophages. *Immunity.* 2019, 50:432-45.e7. [10.1016/j.immuni.2018.12.018](#)
155. Pant K, Venugopal SK, Lorenzo Pisarello MJ, Gradilone SA: The role of gut microbiome-derived short-chain fatty acid butyrate in hepatobiliary diseases. *Am J Pathol.* 2023, 193:1455-67. [10.1016/j.ajpath.2023.06.007](#)
156. Peng K, Dong W, Luo T, Tang H, Zhu W, Huang Y, Yang X: Butyrate and obesity: current research status and future prospect. *Front Endocrinol (Lausanne).* 2023, 14:1098881. [10.3389/fendo.2023.1098881](#)
157. van Deuren T, Blaak EE, Canfora EE: Butyrate to combat obesity and obesity-associated metabolic disorders: current status and future implications for therapeutic use. *Obes Rev.* 2022, 23:e13498. [10.1111/obr.13498](#)
158. Liu Y, Huang W, Wang J, et al.: Multifaceted impacts of periodontal pathogens in disorders of the intestinal barrier. *Front Immunol.* 2021, 12:693479. [10.3389/fimmu.2021.693479](#)
159. Tan X, Wang Y, Gong T: The interplay between oral microbiota, gut microbiota and systematic diseases. *J Oral Microbiol.* 2023, 15:2213112. [10.1080/20002297.2023.2213112](#)
160. Vliex LM, Penders J, Nauta A, Zoetendal EG, Blaak EE: The individual response to antibiotics and diet-insights into gut microbial resilience and host metabolism. *Nat Rev Endocrinol.* 2024, 20:387-98. [10.1038/s41574-024-00966-0](#)
161. Hwang I, Park YJ, Kim YR, et al.: Alteration of gut microbiota by vancomycin and bacitracin improves insulin resistance via glucagon-like peptide 1 in diet-induced obesity. *FASEB J.* 2015, 29:2397-411. [10.1096/fj.14-265983](#)
162. Oettinger-Barak O, Dashper SG, Catmull DV, Adams GG, Sela MN, Machtei EE, Reynolds EC: Antibiotic susceptibility of Aggregatibacter actinomycetemcomitans JP2 in a biofilm. *J Oral Microbiol.* 2013, 5:20320. [10.3402/jom.v5i0.20320](#)
163. Bhat KG, Khot P, Patil S, Pattar G, Majukar S: Antimicrobial susceptibility pattern of oral isolates of Aggregatibacter actinomycetemcomitans. *J Oral Maxillofac Pathol.* 2019, 23:231-5. [10.4103/jomfp.JOMFP_123_19](#)
164. Machado V, Botelho J, Escalda C, et al.: Serum C-reactive protein and periodontitis: a systematic review and meta-analysis. *Front Immunol.* 2021, 12:706432. [10.3389/fimmu.2021.706432](#)
165. Kunutsor SK: Gamma-glutamyltransferase-friend or foe within?. *Liver Int.* 2016, 36:1723-34. [10.1111/liv.13221](#)
166. Akaydin SY, Salihoğlu EM, Güngör DG, Karanlık H, Demokan S: Correlation between gamma-glutamyl transferase activity and glutathione levels in molecular subgroups of breast cancer. *Eur J Breast Health.* 2020, 16:72-6.

167. Fornaciari I, Fierabracci V, Corti A, et al.: Gamma-glutamyltransferase fractions in human plasma and bile: characteristic and biogenesis. *PLoS One*. 2014, 9:e88532. [10.1371/journal.pone.0088532](https://doi.org/10.1371/journal.pone.0088532)
168. Xing M, Gao M, Li J, Han P, Mei L, Zhao L: Characteristics of peripheral blood gamma-glutamyl transferase in different liver diseases. *Medicine (Baltimore)*. 2022, 101:e28443. [10.1097/MD.00000000000028443](https://doi.org/10.1097/MD.00000000000028443)
169. Hanigan MH: Gamma-glutamyl transpeptidase: redox regulation and drug resistance. *Adv Cancer Res*. 2014, 122:103-41. [10.1016/B978-0-12-420117-0.00003-7](https://doi.org/10.1016/B978-0-12-420117-0.00003-7)
170. Pizzorno J: Glutathione!. *Integr Med (Encinitas)*. 2014, 13:8-12.
171. Whitfield JB: Gamma glutamyl transferase. *Crit Rev Clin Lab Sci*. 2001, 38:263-355. [10.1080/20014091084227](https://doi.org/10.1080/20014091084227)
172. Jiang S, Jiang D, Tao Y: Role of gamma-glutamyltransferase in cardiovascular diseases. *Exp Clin Cardiol*. 2013, 18:53-6.
173. Sung Y, Lee YJ, Jung DH, Park B: Potential association of isolated γ -glutamyltransferase elevation with incident ischemic heart disease in lean Koreans. *J Pers Med*. 2022, 12:1966. [10.3390/jpm12121966](https://doi.org/10.3390/jpm12121966)
174. Azizi F: Gamma-glutamyl transpeptidase levels in thyroid disease. *Arch Intern Med*. 1982, 142:79-81.
175. Yorke E: Hyperthyroidism and liver dysfunction: a review of a common comorbidity. *Clin Med Insights Endocrinol Diabetes*. 2022, 15:[10.1177/11795514221074672](https://doi.org/10.1177/11795514221074672)
176. van Beek JH, de Moor MH, Geels LM, et al.: The association of alcohol intake with γ -glutamyl transferase (GGT) levels: evidence for correlated genetic effects. *Drug Alcohol Depend*. 2014, 134:99-105. [10.1016/j.drugalcdep.2013.09.016](https://doi.org/10.1016/j.drugalcdep.2013.09.016)
177. Feng G, Feng L, Zhao Y: Association between ratio of γ -glutamyl transpeptidase to high-density lipoprotein cholesterol and prevalence of nonalcoholic fatty liver disease and metabolic syndrome: a cross-sectional study. *Ann Transl Med*. 2020, 8:634. [10.21037/atm-19-4516](https://doi.org/10.21037/atm-19-4516)
178. Baltacıoğlu E, Akalin FA, Alver A, Değer O, Karabulut E: Protein carbonyl levels in serum and gingival crevicular fluid in patients with chronic periodontitis. *Arch Oral Biol*. 2008, 53:716-22. [10.1016/j.archoralbio.2008.02.002](https://doi.org/10.1016/j.archoralbio.2008.02.002)
179. G K, Shankar SM, Nagesh U, Gururaj SB, Chidambar CK, Bhushan K: Assessment of gingival crevicular fluid levels of gamma glutamyl transferase in chronic periodontitis patients before and after non-surgical periodontal therapy: a clinico-biochemical study. *J Oral Biol Craniofac Res*. 2022, 12:481-5. [10.1016/j.jobcr.2022.03.002](https://doi.org/10.1016/j.jobcr.2022.03.002)
180. Morita T, Yamazaki Y, Fujiharu C, et al.: Serum γ -glutamyltransferase level is associated with periodontal disease independent of drinking habits in Japanese adults. *Med Sci Monit*. 2014, 20:2109-16. [10.12659/MSM.891204](https://doi.org/10.12659/MSM.891204)
181. Pietropaoli D, Del Pinto R, Ferri C, Marzo G, Giannoni M, Ortu E, Monaco A: Association between periodontal inflammation and hypertension using periodontal inflamed surface area and bleeding on probing. *J Clin Periodontol*. 2020, 47:160-72. [10.1111/jcpe.13216](https://doi.org/10.1111/jcpe.13216)
182. Nesse W, Linde A, Abbas F, et al.: Dose-response relationship between periodontal inflamed surface area and HbA1c in type 2 diabetics. *J Clin Periodontol*. 2009, 36:295-300. [10.1111/j.1600-051X.2009.01377.x](https://doi.org/10.1111/j.1600-051X.2009.01377.x)
183. Åberg F, Helenius-Hietala J: Oral health and liver disease: bidirectional associations-a narrative review. *Dent J (Basel)*. 2022, 10:16. [10.3390/dj10020016](https://doi.org/10.3390/dj10020016)
184. Kamata Y, Kessoku T, Shimizu T, et al.: Efficacy and safety of PERIODontal treatment versus usual care for Nonalcoholic liver disease: protocol of the PERION multicenter, two-arm, open-label, randomized trial. *Trials*. 2020, 21:291. [10.1186/s13063-020-4201-y](https://doi.org/10.1186/s13063-020-4201-y)
185. Cobb CM, Sottosanti JS: A re-evaluation of scaling and root planing. *J Periodontol*. 2021, 92:1370-8. [10.1002/JPER.20-0839](https://doi.org/10.1002/JPER.20-0839)
186. Kessoku T, Imajo K, Kobayashi T, et al.: Lubiprostone in patients with non-alcoholic fatty liver disease: a randomized, double-blind, placebo-controlled, phase 2a trial. *Lancet Gastroenterol Hepatol*. 2020, 5:996-1007. [10.1016/S2468-1253\(20\)30216-8](https://doi.org/10.1016/S2468-1253(20)30216-8)
187. Kessoku T, Imajo K, Kobayashi T, et al.: Efficacy, safety, and tolerability of lubiprostone for the treatment of non-alcoholic fatty liver disease in adult patients with constipation: the LUBIPRONE, double-blind, randomised, placebo-controlled study design. *Contemp Clin Trials*. 2018, 69:40-7. [10.1016/j.cct.2018.04.002](https://doi.org/10.1016/j.cct.2018.04.002)
188. Park YS, Kang SB, Marchelletta RR, et al.: The CIC-2 chloride channel activator, lubiprostone, improves intestinal barrier function in biopsies from Crohn's disease but not ulcerative colitis patients. *Pharmaceutics*. 2023, 15:811. [10.3390/pharmaceutics15030811](https://doi.org/10.3390/pharmaceutics15030811)
189. Ito Y, Yoshioka K, Hayashi K, et al.: Prevalence of non-alcoholic fatty liver disease detected by computed tomography in the general population compared with ultrasonography. *Intern Med*. 2024, 63:159-67. [10.2169/internalmedicine.1861-23](https://doi.org/10.2169/internalmedicine.1861-23)
190. Ahmad A, Furuta M, Shinagawa T, Takeuchi K, Takeshita T, Shimazaki Y, Yamashita Y: Association of periodontal status with liver abnormalities and metabolic syndrome. *J Oral Sci*. 2015, 57:335-43. [10.2334/josnusd.57.335](https://doi.org/10.2334/josnusd.57.335)
191. Nana Nana AR, Tsobgny Tsague NF, Lontchi-Yimagou E, et al.: Effects of non-surgical treatment of chronic periodontitis on insulin resistance and glucose tolerance in subjects without diabetes (PARODIA 2 study). *J Investig Med*. 2021, 69:1377-81. [10.1136/jim-2021-001831](https://doi.org/10.1136/jim-2021-001831)
192. Sun WL, Chen LL, Zhang SZ, Wu YM, Ren YZ, Qin GM: Inflammatory cytokines, adiponectin, insulin resistance and metabolic control after periodontal intervention in patients with type 2 diabetes and chronic periodontitis. *Intern Med*. 2011, 50:1569-74. [10.2169/internalmedicine.50.5166](https://doi.org/10.2169/internalmedicine.50.5166)
193. Mohan M, Jhingran R, Bains VK, Gupta V, Madan R, Rizvi I, Mani K: Impact of scaling and root planing on C-reactive protein levels in gingival crevicular fluid and serum in chronic periodontitis patients with or without diabetes mellitus. *J Periodontal Implant Sci*. 2014, 44:158-68. [10.5051/jpis.2014.44.4.158](https://doi.org/10.5051/jpis.2014.44.4.158)
194. Bian Y, Liu C, Fu Z: Application value of combination therapy of periodontal curettage and root planing on moderate-to-severe chronic periodontitis in patients with type 2 diabetes. *Head Face Med*. 2021, 17:12.
195. Green BN, Johnson CD, Adams A: Writing narrative literature reviews for peer-reviewed journals: secrets of the trade. *J Chiropr Med*. 2006, 5:101-17. [10.1016/S0899-3467\(07\)60142-6](https://doi.org/10.1016/S0899-3467(07)60142-6)
196. Vidal EI, Fukushima FB: The art and science of writing a scientific review article. *Cad Saude Publica*. 2021, 37:e00063121. [10.1590/0102-311X00063121](https://doi.org/10.1590/0102-311X00063121)

197. Sukhera J: Narrative reviews: flexible, rigorous, and practical. *J Grad Med Educ.* 2022, 14:414-7. [10.4300/JGME-D-22-00480.1](https://doi.org/10.4300/JGME-D-22-00480.1)
198. Jahan N, Naveed S, Zeshan M, Tahir MA: How to conduct a systematic review: a narrative literature review . *Cureus.* 2016, 8:e864. [10.7759/cureus.864](https://doi.org/10.7759/cureus.864)
199. Basheer A: The art and science of writing narrative reviews . *Int J Adv Med Health Res.* 2022, 9:124-6. [10.4103/ijamr.ijamr_234_22](https://doi.org/10.4103/ijamr.ijamr_234_22)
200. Greenhalgh T, Thorne S, Malterud K: Time to challenge the spurious hierarchy of systematic over narrative reviews?. *Eur J Clin Invest.* 2018, 48:e12931. [10.1111/eci.12931](https://doi.org/10.1111/eci.12931)
201. Petticrew M, Rehfuess E, Noyes J, et al.: Synthesizing evidence on complex interventions: how meta-analytical, qualitative, and mixed-method approaches can contribute. *J Clin Epidemiol.* 2013, 66:1230-43. [10.1016/j.jclinepi.2013.06.005](https://doi.org/10.1016/j.jclinepi.2013.06.005)
202. Dixon-Woods M, Agarwal S, Jones D, Young B, Sutton A: Synthesising qualitative and quantitative evidence: a review of possible methods. *J Health Serv Res Policy.* 2005, 10:45-53. [10.1177/135581960501000110](https://doi.org/10.1177/135581960501000110)