

Enamel Matrix Derivative (EMD) as an Adjunct to Non-surgical Periodontal Therapy: A Systematic Review

Review began 07/25/2023

Review ended 08/12/2023

Published 08/15/2023

© Copyright 2023

Abu-Ta'a et al. This is an open access article distributed under the terms of the Creative Commons Attribution License CC-BY 4.0., which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Mahmoud Abu-Ta'a¹, Dina Marzouka¹

1. Dental Sciences, Arab American University, Ramallah, PSE

Corresponding author: Mahmoud Abu-Ta'a, mahmoud.abutaa@aaup.edu

Abstract

If left untreated, periodontitis is a chronic, irreversible disease that can contribute to tooth loss. The primary objective of periodontal treatment is to arrest the progression of the disease and restore the supporting structures of the tooth. Scaling and root planing (SRP) is a common non-surgical periodontal therapy (NSPT) used to reduce inflammation, pocket depth, and clinical attachment loss. However, NSPT has limitations, notably in difficult-to-access deep pockets and molar furcations. Deep pockets (greater than 4 mm) frequently retain calculus following NSPT. To attain direct access, surgical periodontal therapy (SPT) is recommended, particularly for pockets deeper than 5 mm. Enamel matrix derivative (EMD) has emerged in recent years as a tool for periodontal regeneration when used in conjunction with NSPT for infrabony defects. EMD may also have advantageous effects when combined with NSPT. The purpose of this review is to provide a thorough understanding of the effects of EMD as an adjunct to NSPT. The databases Scopus, PubMed/MEDLINE, Google Scholar, Cochrane, and Embase were systematically searched to identify relevant studies on the benefits of EMD and its use as an adjunct to NSPT. Incorporating EMD into NSPT reduces chair time, and 60% of studies demonstrated considerable benefits compared to SRP alone, according to the findings. On the basis of research, it can be concluded that EMD can be used as an adjunct to NSPT, thereby reducing the amount of time spent in the operating chair. In some cases, it can, therefore, be regarded as an alternative to surgical treatment.

Categories: Dentistry

Keywords: root planing, scaling, regeneration, surgical therapy, non-surgical periodontal therapy, periodontitis, enamel matrix derivative

Introduction And Background

The primary goal of periodontal treatment is to eliminate inflammation and restore the periodontium's integrity, which includes vital components, such as cementum, bone, and the periodontal ligament. Periodontal pocket depth (PPD), which functions as a crucial clinical parameter, frequently guides the decision-making process for treatment. Surgical periodontal therapy (SPT) and non-SPT (NSPT), the latter being a less invasive option [1,2], comprise a spectrum of periodontitis management strategies.

The initial phase of periodontitis treatment necessitates effective control of subgingival biofilm, which necessitates collaboration between dental professionals and patients. Concurrently, periodontal disease risk factors must be addressed prior to proceeding with subsequent interventions [3,4]. Various agents have been investigated as potential adjuncts to NSPT in recent scientific studies. Enamel matrix derivative (EMD), Coe-PakTM dressings, and low-level lasers are notable examples [5-7].

EMD, a composite composed of enamel matrix proteins, has been used historically in SPT to enhance periodontal regeneration, particularly in the treatment of infrabony defects [8,9]. In addition, research has revealed the beneficial impacts of EMD when incorporated into SPT [10-12].

Due to the inherent biological properties of EMD, numerous attempts have been made to investigate its potential as an adjunct to NSPT. While some studies failed to identify additional benefits from incorporating EMD into NSPT when compared to scaling and root planing (SRP) alone [13-16], others have demonstrated that EMD can indeed serve as a beneficial adjunct to NSPT, resulting in positive outcomes [11,12,17-19].

In the context of this review, we will investigate the effects of adding EMD to NSPT. This investigation will be based on randomized clinical trials that compared the effects of integrating EMD into NSPT with those of NSPT alone. The objective is to assess whether EMD can enhance the efficacy of NSPT in a scientifically rigorous and comprehensive manner.

Review

Methodology

How to cite this article

Abu-Ta'a M, Marzouka D (August 15, 2023) Enamel Matrix Derivative (EMD) as an Adjunct to Non-surgical Periodontal Therapy: A Systematic Review. Cureus 15(8): e43530. DOI 10.7759/cureus.43530

Cochrane, Google Scholar, PubMed MEDLINE, and Embase databases were used for bibliographic search using the following search items: (“EMD” OR “enamel matrix derivative” OR “enamel matrix proteins”) and (“regeneration” OR “papilla preservation flap” OR “surgical periodontal therapy” OR “non-surgical periodontal therapy” OR “SRP” OR “deep pocket depth”). As well, a search was performed for all the reference lists of all the primary sources.

This review included all randomized clinical trials that evaluated the effect of EMD as an adjunct to NSPT in comparison to NSPT alone, which were published in the English language until September 2022. During the search process, no study based on publication date was restricted (Figure 1).

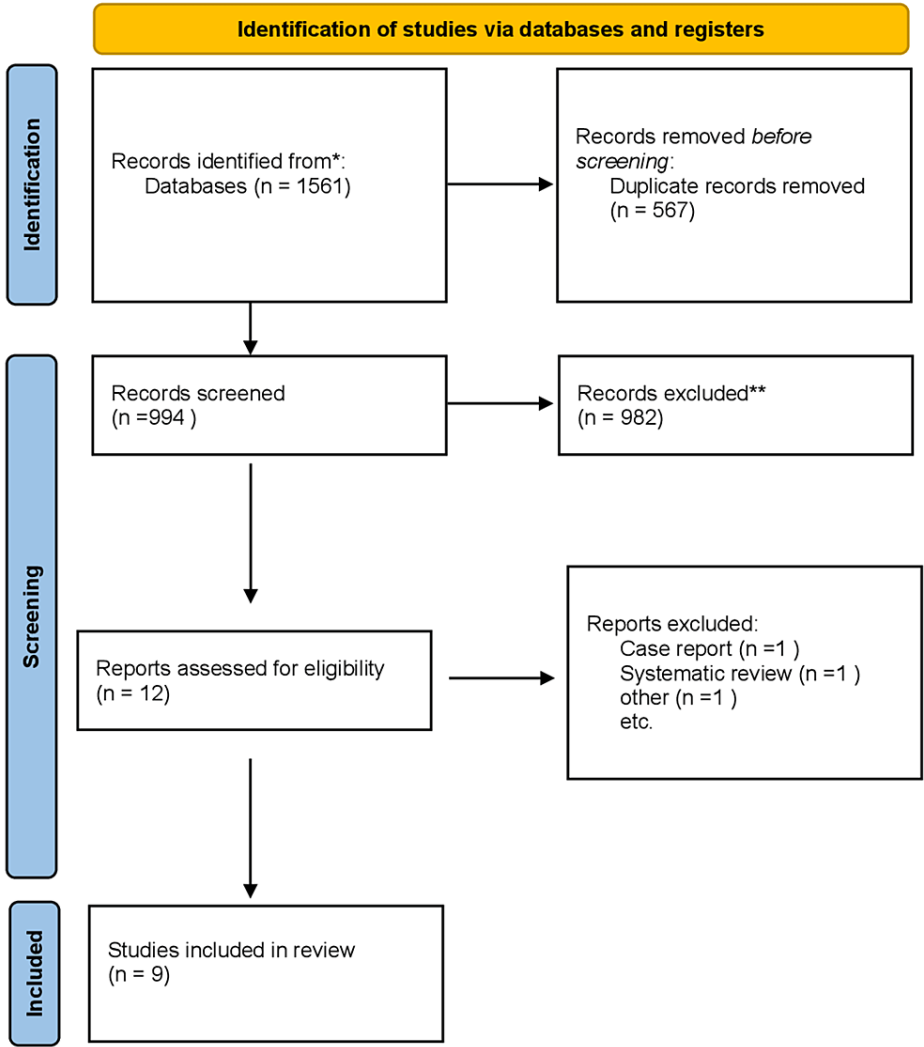


FIGURE 1: Flowchart diagram showing the inclusion and exclusion of studies

Results

According to the studies included in this review, there is a beneficial effect of using EMD as an adjunct to NSPT as it decreases chair time and improves early soft tissue healing. However, 60% of the studies showed an added benefit from using EMD as an adjunct to NSPT, whereas 40% of the studies showed no significant difference between EMD as an adjunct to NSPT compared to SRP alone.

The lack of definitive protocol and differences in the clinical parameters measured in each study limits the ability to conclude the result. However, the number of studies with no significant difference between using EMD as an adjunct to SRP is more than studies with an added beneficial effect from using EMD, when measuring the clinical parameters PPD (Figure 2) and clinical attachment level (CAL) (Figure 3).

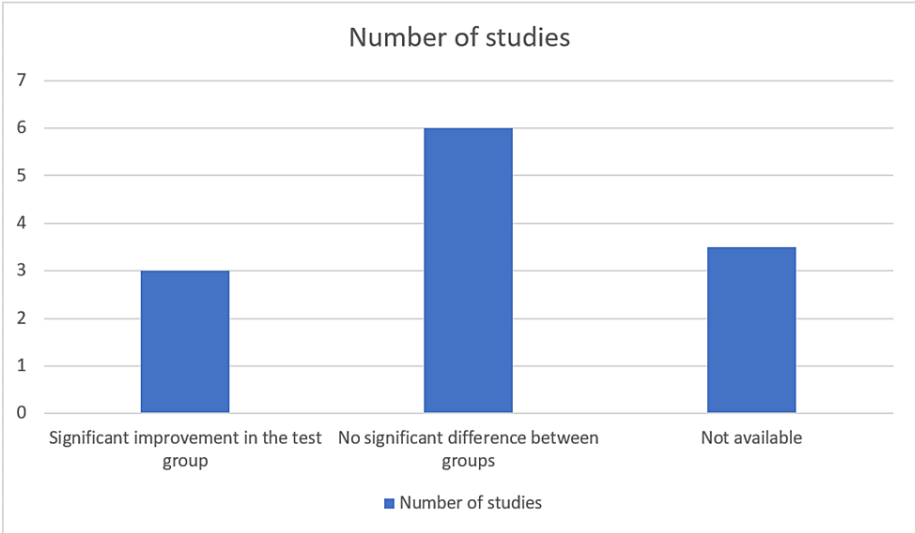


FIGURE 2: RCTs showing the effect of EMD on probing pocket depth

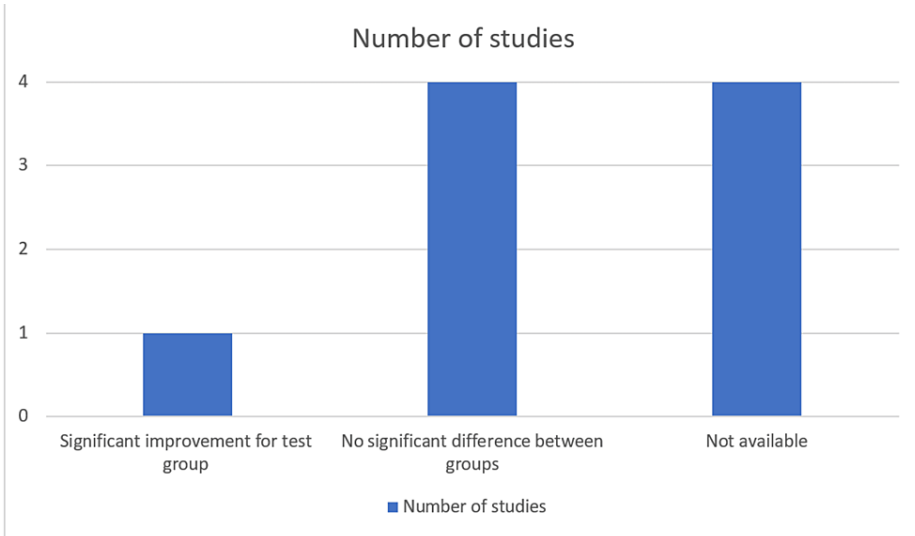


FIGURE 3: RCTs showing the effect of EMD on clinical attachment level

Discussion

Periodontitis

Periodontitis is one of the major chronic diseases worldwide [20]. Affecting the supporting apparatus of the tooth and causing irreversible damage, which is caused by the bacterial biofilm [21]. It is characterized by the imbalance between the host immune response and bacteria [22]. The risk factors for periodontitis are smoking and diabetes and other contributing factors, such as overhang restorations, anatomical factors, interproximal spaces, and occlusal trauma [23]. It may occur in adolescence or childhood, yet it is more common in patients aged more than 40 years old [23,24]. This disease can be controlled if intercepted in the early stage; nevertheless, it can cause damage and tooth loss if left untreated [25].

The main goal of periodontal treatment is the removal of the infection in order to arrest the disease, prevent its progression, and minimize symptoms. This resolution is manifested clinically by shallow PDs and the absence of bleeding on probing (BOP) [26]. However, when there are persistent deep pockets of more than 5 mm, there is a risk of disease progression, and this results in tooth loss whether there is bleeding on probing or not [27,28].

Treatment of periodontitis includes surgical and non-surgical options with behavioral changes, such as oral hygiene instructions (OHIs) and a smoking cessation program [29,30]. Daily supragingival plaque control

seems to be important in order to preserve periodontal health [31,32].

According to Graziani et al. [33], it was shown that no periodontal therapy has superiority over any other treatment option [33]. Moreover, it was concluded that NSPT seeks to reduce gingival inflammation (GI) and PD and facilitates tissue healing and patient hygiene [34]. NSPT should be done in the initial cause-related therapy, and SRP should be done in deep pockets 5 mm or greater using manual or ultrasonic scalers [23], while SPT provides accessibility for SRP, especially in sites with furcation and infrabony defect involvement, and provides bone and gingival contouring for facilitating self-oral hygiene [35,36]. Residual calculus accumulation was found in pockets more than 4 mm after SRP [37]; thus, SPT should be done after the hygienic phase in the remaining deep pockets (more than 5 mm) in order to avoid mechanical damage of the periodontium. In addition, surgical therapy should be delayed if there is no adequate biofilm removal [33,38-40].

Increased probing depth following treatment means that there is an infrabony defect (angular defects); thus, this angular defect seems to worsen the prognosis of the teeth [41].

Despite the fact that surgery and NSPT result in improvement of the clinical outcome, it is characterized by long junctional epithelium, which means repair and not regeneration (formation of new cementum, periodontal ligament (PDL) fibers, and new alveolar bone) [42]. Moreover, regeneration of periodontal tissue is the goal of periodontal treatment [43,44], and different surgical techniques include implantation of bone grafts or/and substitutes in order to achieve periodontal regeneration [2,45,46].

Some systematic reviews showed that these regenerative techniques result in better clinical outcomes in terms of clinical attachment level (CAL), PD, and hard tissue fill compared to open flap debridement (OFD) alone [2]. Meanwhile, other studies showed that these clinical outcomes can be maintained long-term in cases where plaque control is maintained [9,45,47-50].

EMD

In 1997, EMD was introduced [51], and it is produced from the enamel of a developing porcine tooth. Consisting of enamel matrix proteins (EMPs), amelogenin is the most important protein and propylene glycol alginate (PGA) [52]. PGA has antibacterial properties [5]. Cells of periodontal ligaments are favored by EMD over epithelial and gingival cells, which do not consist of cytotoxic but cytostasis (inhibition of cells), having the least effect on epithelial cells [9]. Moreover, EMD did not show any side effects, such as allergic or incompatibility [53,54].

EMPs are secreted from Hertwig's epithelial root sheet, which is able to promote periodontal regeneration [55]. It was shown that it induces bone cells (osteoblasts) and PDL cells and stimulates the proliferation of osteogenesis [35,56-59], and it was also shown that it stimulates the growth of new blood vessels [60,61].

EMD has been used in regenerative periodontal therapy; in some studies, it has been shown that there are better clinical outcomes using EMD in SPT compared to OFD alone [62].

A randomized clinical study by Heijl et al. showed that the CAL, when using EMD and surgery (test group), is 2.2 mm, while it is 1.7 mm when applying OFD alone (control group). The radiographs in the test group showed that there is a 2.6 mm bone fill, and 66% of the defect is filled, while the control group did not show any bone fill [63]. Furthermore, the study by Froum et al. [64] showed three times bone fill in the test group compared to the control group, and 74% fill of the defect filled versus 23% fill of the defect in the control group.

Some studies included that the application of EMD in conjunction with surgical treatment caused regeneration of new PDL, cementum, and bone in angular bone and class II furcation defects [9,52,65]. Moreover, it was concluded that EMD improves soft tissue healing in the SPT in the post-operative period [66].

Application of the EMD in SPT

The key elements for successful regeneration are space provision and clot stabilization in infrabony defects [67-69]. Although EMD has limited space-making potential due to its gel-like consistency, this will not provide sufficient soft tissue support [9,67]. Thus, root instrumentation should be done after flap elevation using a simplified papilla preservation flap (SPPT) or modified papilla preservation flap (MPPT) in order to obtain primary closure and clot stabilization, thus obtaining better clinical results [70,71].

Ethylenediaminetetraacetic acid (EDTA) is always used before the application of EMD as a root surface bio modifier. It has a natural PH; thus, the vitality of the tissue is preserved [72]. It was shown that collagen fibers are exposed, the smear layer is removed, and dentinal tubules are demineralized by EDTA [73]. Thus, the root is conditioned with EDTA 24% for two minutes, followed by the application of EMD [74].

EMD as an Adjunct to NSPT

In order to preserve the soft tissue and improve wound healing, a minimally invasive technique is preferred over surgical therapy [75]. EMD showed a beneficial effect when used in SPT. Thus, this effect may have benefits when using EMD with NSPT.

Some clinical studies evaluated the effect of the EMD as an adjunct to NSPT but failed to find additional benefits when compared to NSPT alone [13,14].

Other randomized clinical studies showed that there are no additional benefits from using EMD with NSPT when compared to SRP alone in terms of CAL and PD, but it was revealed that using EMD with NSPT has less postoperative pain [16]. While in another randomized clinical trial, it was revealed that, in the test group, two of 10 teeth showed insignificant new cementum and new bone formation; however, it failed to show a significant benefit from using EMD as an adjunct to NSPT [52]. Furthermore, a significant improvement after 12 months of treating infrabony defects, less than 8 mm, was observed with minimally invasive non-surgical techniques, but failed to show significant improvement in radiographic outcomes when using additional EMD [5]. By incorporating both non-surgical and minimally invasive surgical perspectives, the systematic review is better equipped to meet the diverse requirements of patients and clinicians. This strategy is consistent with the dynamic nature of periodontal therapy and the recognition that there is no single modality that is universally superior. This study contributes a more holistic comprehension of how EMD can enhance various periodontal treatment strategies and thus fits well within the existing literature.

In a case report of four cases, three cases had new cementum and new bone formation; thus, it was found, in the case report, that EMD has a beneficial effect and can be used after NSPT in deep pockets. According to the study, it was concluded that EMD as an adjunct to NSPT is used in patients who refuse surgical treatment [18]. In addition to this, it was concluded in a randomized clinical study that there is no change in the clinical parameters when comparing EMD with SRP versus SRP alone (Table 1). But using EMD inhibits the development of *Prevotella* and *Porphyromonas gingivalis* [17].

Authors, year, and study design	Age	Sample size	Test group	Control group	Follow-up	Clinical and histological parameters	Clinical evidence
Anoixiadou et al. [5], 2022, RCT	More than 18 years	36 patients with infrabony defects	18 patients, minimally invasive surgical technique (MINST)+ EMD	18 patients, MINST alone	6 and 12 months	CAL, BOP, and PPD	EMD has no added benefit over MINST alone. EMD significantly decreases PPD with BOP in sites with a baseline pocket depth of 8 mm or less (92% in the test group and 69% in the control group).
Schallhorn et al. [12], 2021, Split mouth RCT	18-85 years old	51 patients with moderate to severe periodontitis	SRP+EMD, second application 2-3 weeks later	SRP alone	12 months	CAL, PPD, and BOP	A significant improvement for both groups in PPD and CAL. A significant improvement in BOP, CAL, and PPD in test sites compared to control sites. Sites that are converted to sites that no longer needs treatment: 65% for control sites and 79% for test sites.
Gutierrez et al. [13], 2003, Split mouth RCT	More than 18 years old	22 patients	22 sites with pocket depth of 5 mm or greater and angular bone defect, with more than 3 mm. EMD application with SRP	22 sites SRP without additional treatment	3 months	CAL, PPD, PI, and bleeding index (BI)	No significant difference in all clinical parameters between two groups. Mean PPD 1.47±0.3 mm for test sites, and 1.87±0.4 mm for control sites.
Mombelli et al. [14], 2005, Split mouth RCT	25–65 years old	16 patients with moderate to advanced periodontitis	16 sites EMD +SRP	16 sites, SRP alone	2, 6, and 12 months	GI, PI, BOP, and PPD	No added benefit from EMD as an adjunct to non-surgical periodontal therapy.
Sculean et al. [15], 2003, RCT	NA.	16 patients with one intrabony defect	1. SRP (hand)+EMD, 2. SRP (ultrasonic)+EMD	SRP alone	6 months	Histological evaluation, CAL, and PPD	PPD reduction and CAL gain in all groups, but no true connective tissue attachment in all three modalities.
Wennström et al. [16], 2002, Split mouth RCT	NA	84 patients with moderate to severe periodontitis	84 sites, EMD +SRP	28 sites, SRP alone	1, 2, and 3 weeks	PPD, BOP, GI, post-treatment discomfort	EMD enhanced early soft tissue healing (at 1 week proportion of people having minimal discomfort were 54% for test group and 31% for control group, but no significant difference at a three-week follow-up. No significant difference in the clinical parameters between groups at a three-week follow-up.

TABLE 1: Summary of the randomized clinical trial (RCTs) comparing EMD as an adjunct to non-surgical periodontal therapy versus non-surgical periodontal alone

According to Schallhorn et al., different results in the studies are caused by a lack of standardized protocol. The application of EMD for a second time on the root during healing will prolong the presence of EMD on the root surface, and this decrease the potential of washing out the material. This double-blinded randomized clinical study concluded that EMD can be used as an adjunctive to non-surgical periodontal treatment in implant-associated diseases. Furthermore, a randomized clinical trial of 51 patients found that the control and the test groups have significant improvement in the clinical attachment level and PPD. Nonetheless, the use of the EMD improves bleeding on probing and increases the number of healthy pockets (less than 5 mm); these pockets were converted to sites that no longer need surgery, a standardized workflow in this study [12].

In summary, the study demonstrated that a flapless technique employing EMD in infrabony defects reduces chair time compared to the minimally invasive technique (MIST) [11]. Additionally, when EMD is used as an adjunct to NSPT for peri-implant mucositis, it provides some additional advantages in terms of PPD and CAL, but it may not result in a full recovery, and its efficacy compared to NSPT alone may not be particularly

significant [19]. Only one study investigated the effectiveness of EMD application as an adjunct to non-surgical retreatment therapy after three months after the initial NSPT and concluded that the application of EMD in the re-instrumentation session increases significantly the frequencies of pocket closure, with no bleeding on probing compared to re-instrumentation alone (Table 1) [76].

Limitations

The lack of a definitive protocol for the EMD application between the studies, variability in follow-ups, and the parameters measured will limit the review from having a definitive conclusion. Moreover, due to heterogeneity in lifestyle in each study because of different countries, more evidence is required to confirm the beneficial effect of EMD as an adjunct to NSPT.

Conclusions

While some studies did not observe additional benefits from using EMD as an adjunct to NSPT compared to non-surgical therapy alone, this may be attributable to the techniques used. These investigations may have employed EMD in a manner that increased the possibility of material washout by the blood. However, when EMD is utilized in a less invasive manner during NSPT, it can substantially reduce chair time and provide greater patient comfort than SPT. Consequently, EMD can still be considered as an adjunct to NSPT, particularly in deep pockets (greater than 5 mm), where surgical alternatives are unavailable or refused by the patient.

In conclusion, EMD can be a valuable instrument when combined with NSPT. It reduces chair time and provides a more comfortable experience for patients, especially those who are ineligible for or refuse surgical treatment.

Additional Information

Disclosures

Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

References

- Heitz-Mayfield LJ, Lang NP: Surgical and nonsurgical periodontal therapy. Learned and unlearned concepts. *Periodontol* 2000. 2013, 62:218-31. [10.1111/prd.12008](#)
- Kao RT, Nares S, Reynolds MA: Periodontal regeneration - intrabony defects: a systematic review from the AAP Regeneration Workshop. *J Periodontol*. 2015, 86:S77-104. [10.1902/jop.2015.130685](#)
- Sanz-Sánchez I, Montero E, Citterio F, Romano F, Molina A, Aimetti M: Efficacy of access flap procedures compared to subgingival debridement in the treatment of periodontitis. A systematic review and meta-analysis. *J Clin Periodontol*. 2020, 47 Suppl 22:282-302. [10.1111/jcpe.13259](#)
- Bouchard P, Carra MC, Boillot A, Mora F, Rangé H: Risk factors in periodontology: a conceptual framework. *J Clin Periodontol*. 2017, 44:125-31. [10.1111/jcpe.12650](#)
- Anoixiadou S, Parashis A, Vouros I: Enamel matrix derivative as an adjunct to minimally invasive non-surgical treatment of intrabony defects: a randomized clinical trial. *J Clin Periodontol*. 2022, 49:134-43. [10.1111/jcpe.13567](#)
- Monje A, Kramp AR, Criado E, Suárez-López Del Amo F, Garaicoa-Pazmiño C, Gargallo-Albiol J, Wang HL: Effect of periodontal dressing on non-surgical periodontal treatment outcomes: a systematic review. *Int J Dent Hyg*. 2016, 14:161-7. [10.1111/idh.12130](#)
- Ren C, McGrath C, Jin L, Zhang C, Yang Y: The effectiveness of low-level laser therapy as an adjunct to non-surgical periodontal treatment: a meta-analysis. *J Periodontol Res*. 2017, 52:8-20. [10.1111/jre.12361](#)
- Esposito M, Grusovin MG, Papanikolaou N, Coulthard P, Worthington HV: Enamel matrix derivative (Emdogain(R)) for periodontal tissue regeneration in intrabony defects. *Cochrane Database Syst Rev*. 2009, 2009:CD003875. [10.1002/14651858.CD003875.pub3](#)
- Sculean A, Alessandri R, Miron R, Salvi GE, Bosshardt DD: Enamel matrix proteins and periodontal wound healing and regeneration. *Clin Adv Periodontics*. 2011, 1:101-17. [10.1902/cap.2011.110047](#)
- Miron RJ, Sculean A, Cochran DL, et al.: Twenty years of enamel matrix derivative: the past, the present and the future. *J Clin Periodontol*. 2016, 43:668-83. [10.1111/jcpe.12546](#)
- Aimetti M, Ferrarotti F, Mariani GM, Romano F: A novel flapless approach versus minimally invasive surgery in periodontal regeneration with enamel matrix derivative proteins: a 24-month randomized controlled clinical trial. *Clin Oral Investig*. 2017, 21:327-37. [10.1007/s00784-016-1795-2](#)
- Schallhorn RA, McClain PK, Benhamou V, Doobrow JH, Grandin HM, Kasaj A: Application of enamel matrix derivative in conjunction with non-surgical therapy for treatment of moderate to severe periodontitis: a 12-month, randomized prospective, multicenter study. *J Periodontol*. 2021, 92:619-28. [10.1002/JPER.19-0579](#)
- Gutierrez MA, Mellonig JT, Cochran DL: Evaluation of enamel matrix derivative as an adjunct to non-surgical periodontal therapy. *J Clin Periodontol*. 2003, 30:739-45. [10.1034/j.1600-051x.2003.00374.x](#)
- Mombelli A, Brochut P, Plagnat D, Casagni F, Giannopoulou C: Enamel matrix proteins and systemic

- antibiotics as adjuncts to non-surgical periodontal treatment: clinical effects. *J Clin Periodontol*. 2005, 32:225-30. [10.1111/j.1600-051X.2005.00664.x](#)
15. Sculean A, Windisch P, Keglévich T, Gera I: Histologic evaluation of human intrabony defects following non-surgical periodontal therapy with and without application of an enamel matrix protein derivative. *J Periodontol*. 2003, 74:153-60. [10.1902/jop.2003.74.2.153](#)
 16. Wennström JL, Lindhe J: Some effects of enamel matrix proteins on wound healing in the dento-gingival region. *J Clin Periodontol*. 2002, 29:9-14. [10.1034/j.1600-051x.2002.290102.x](#)
 17. Wyganowska-Świątkowska M, Szkaradkiewicz AK, Karpiński TM, Marcinkowski JT: The evaluation of enamel matrix derivative on subgingival microbial environment in non-surgical periodontal therapy. *Ann Agric Environ Med*. 2013, 30:431-5.
 18. Mellonig JT, Valderrama P, Gregory HJ, Cochran DL: Clinical and histologic evaluation of non-surgical periodontal therapy with enamel matrix derivative: a report of four cases. *J Periodontol*. 2009, 80:1534-40. [10.1902/jop.2009.090160](#)
 19. Kashefimehr A, Pourabbas R, Faramarzi M, Zarandi A, Moradi A, Tenenbaum HC, Azarpazhooh A: Effects of enamel matrix derivative on non-surgical management of peri-implant mucositis: a double-blind randomized clinical trial. *Clin Oral Investig*. 2017, 21:2379-88. [10.1007/s00784-016-2033-7](#)
 20. Petersen PE, Baehni PC: Periodontal health and global public health. *Periodontol* 2000. 2012, 60:7-14. [10.1111/j.1600-0757.2012.00452.x](#)
 21. Savage A, Eaton KA, Moles DR, Needleman I: A systematic review of definitions of periodontitis and methods that have been used to identify this disease. *J Clin Periodontol*. 2009, 36:458-67. [10.1111/j.1600-051X.2009.01408.x](#)
 22. Yucel-Lindberg T, Båge T: Inflammatory mediators in the pathogenesis of periodontitis. *Expert Rev Mol Med*. 2013, 15:e7. [10.1017/erm.2013.8](#)
 23. Kwon T, Lamster IB, Levin L: Current concepts in the management of periodontitis. *Int Dent J*. 2021, 71:462-76. [10.1111/idj.12630](#)
 24. Botero JE, Rösing CK, Duque A, Jaramillo A, Contreras A: Periodontal disease in children and adolescents of Latin America. *Periodontol* 2000. 2015, 67:34-57. [10.1111/prd.12072](#)
 25. Slots J: Periodontitis: facts, fallacies and the future. *Periodontol* 2000. 2017, 75:7-23. [10.1111/prd.12221](#)
 26. Ganeles J, Listgarten MA, Evian CI: Ultrastructure of durapatite-periodontal tissue interface in human intrabony defects. *J Periodontol*. 1986, 57:133-40. [10.1902/jop.1986.57.3.133](#)
 27. Claffey N, Egelberg J: Clinical indicators of probing attachment loss following initial periodontal treatment in advanced periodontitis patients. *J Clin Periodontol*. 1995, 22:690-6. [10.1111/j.1600-051x.1995.tb00828.x](#)
 28. Matulienė G, Pjetursson BE, Salvi GE, Schmidlin K, Brägger U, Zwahlen M, Lang NP: Influence of residual pockets on progression of periodontitis and tooth loss: results after 11 years of maintenance. *J Clin Periodontol*. 2008, 35:685-95. [10.1111/j.1600-051X.2008.01245.x](#)
 29. Jönsson B, Baker SR, Lindberg P, Oscarson N, Ohn K: Factors influencing oral hygiene behaviour and gingival outcomes 3 and 12 months after initial periodontal treatment: an exploratory test of an extended theory of reasoned action. *J Clin Periodontol*. 2012, 39:138-44. [10.1111/j.1600-051X.2011.01822.x](#)
 30. Jönsson B, Ohn K, Lindberg P, Oscarson N: Evaluation of an individually tailored oral health educational programme on periodontal health. *J Clin Periodontol*. 2010, 37:912-9. [10.1111/j.1600-051X.2010.01590.x](#)
 31. Serino G, Rosling B, Ramberg P, Socransky SS, Lindhe J: Initial outcome and long-term effect of surgical and non-surgical treatment of advanced periodontal disease. *J Clin Periodontol*. 2001, 28:910-6. [10.1034/j.1600-051x.2001.028010910.x](#)
 32. Tonetti MS, Chapple IL: Biological approaches to the development of novel periodontal therapies - consensus of the Seventh European Workshop on Periodontology. *J Clin Periodontol*. 2011, 38:114-8. [10.1111/j.1600-051X.2010.01675.x](#)
 33. Graziani F, Karapetsa D, Alonso B, Herrera D: Nonsurgical and surgical treatment of periodontitis: how many options for one disease?. *Periodontol* 2000. 2017, 75:152-88. [10.1111/prd.12201](#)
 34. Ramfjord SP, Caffesse RG, Morrison EC, et al.: 4 modalities of periodontal treatment compared over 5 years. *J Clin Periodontol*. 1987, 14:445-52. [10.1111/j.1600-051x.1987.tb02249.x](#)
 35. Vivekananda MR, Vandana KL, Bhat KG: Effect of the probiotic *Lactobacilli reuteri* (Prodentis) in the management of periodontal disease: a preliminary randomized clinical trial. *J Oral Microbiol*. 2010, 2:5344. [10.3402/jom.v2i0.5344](#)
 36. Wachtel H, Schenk G, Böhm S, Weng D, Zuh O, Hürzeler MB: Microsurgical access flap and enamel matrix derivative for the treatment of periodontal intrabony defects: a controlled clinical study. *J Clin Periodontol*. 2003, 30:496-504. [10.1034/j.1600-051x.2003.00013.x](#)
 37. Waerhaug J: Healing of the dento-epithelial junction following subgingival plaque control. II: As observed on extracted teeth. *J Periodontol*. 1978, 49:119-34. [10.1902/jop.1978.49.3.119](#)
 38. Kwon T, Salem DM, Levin L: Nonsurgical periodontal therapy based on the principles of cause-related therapy: rationale and case series. *Quintessence Int*. 2019, 50:370-6. [10.3290/j.qi.a42292](#)
 39. Labriola A, Needleman I, Moles DR: Systematic review of the effect of smoking on nonsurgical periodontal therapy. *Periodontol* 2000. 2005, 37:124-37. [10.1111/j.1600-0757.2004.03793.x](#)
 40. Renvert S, Persson GR: Supportive periodontal therapy. *Periodontol* 2000. 2004, 36:179-95. [10.1111/j.1600-0757.2004.03680.x](#)
 41. Papapanou PN, Tonetti MS: Diagnosis and epidemiology of periodontal osseous lesions. *Periodontol* 2000. 2000, 22:8-21. [10.1034/j.1600-0757.2000.2220102.x](#)
 42. Caton JG, Greenstein G: Factors related to periodontal regeneration. *Periodontol* 2000. 2003, 1:9-15.
 43. Wang HL, Greenwell H, Fiorellini J, et al.: Position paper: periodontal regeneration. *J Periodontol*. 2005, 76:1601-22. [10.1902/jop.2005.76.9.1601](#)
 44. Wikesjö UM, Selvig KA: Periodontal wound healing and regeneration. *Periodontol* 2000. 1999, 19:21-39. [10.1111/j.1600-0757.1999.tb00145.x](#)
 45. Cortellini P, Tonetti MS: Long-term tooth survival following regenerative treatment of intrabony defects. *J Periodontol*. 2004, 75:672-8. [10.1902/jop.2004.75.5.672](#)
 46. Stavropoulos A, Windisch P, Gera I, Capsius B, Sculean A, Wikesjö UM: A phase IIa randomized controlled

- clinical and histological pilot study evaluating rhGDF-5/ β -TCP for periodontal regeneration. *J Clin Periodontol.* 2011, 38:1044-54. [10.1111/j.1600-051X.2011.01778.x](#)
47. Sculean A, Kiss A, Miliauskaitė A, Schwarz F, Arweiler NB, Hannig M: Ten-year results following treatment of intra-bony defects with enamel matrix proteins and guided tissue regeneration. *J Clin Periodontol.* 2008, 35:817-24. [10.1111/j.1600-051X.2008.01295.x](#)
 48. Sculean A, Chiantella GC, Arweiler NB, Becker J, Schwarz F, Stavropoulos A: Five-year clinical and histologic results following treatment of human intrabony defects with an enamel matrix derivative combined with a natural bone mineral. *Int J Periodontics Restorative Dent.* 2008, 28:153-61.
 49. Stavropoulos A, Karring T: Long-term stability of periodontal conditions achieved following guided tissue regeneration with bioresorbable membranes: case series results after 6-7 years. *J Clin Periodontol.* 2004, 31:939-44. [10.1111/j.1600-051X.2004.00586.x](#)
 50. Silvestri M, Rasperini G, Milani S: 120 infrabony defects treated with regenerative therapy: long-term results. *J Periodontol.* 2011, 82:668-75. [10.1902/jop.2010.100297](#)
 51. Heijl L: Periodontal regeneration with enamel matrix derivative in one human experimental defect. A case report. *J Clin Periodontol.* 1997, 24:693-6.
 52. Kalpidis CD, Ruben MP: Treatment of intrabony periodontal defects with enamel matrix derivative: a literature review. *J Periodontol.* 2002, 73:1360-76. [10.1902/jop.2002.73.11.1360](#)
 53. Froum S, Weinberg M, Novak J, et al.: A multicenter study evaluating the sensitization potential of enamel matrix derivative after treatment of two infrabony defects. *J Periodontol.* 2004, 75:1001-8. [10.1902/jop.2004.75.7.1001](#)
 54. Petinaki E, Nikolopoulos S, Castanas E: Low stimulation of peripheral lymphocytes, following in vitro application of Emdogain. *J Clin Periodontol.* 1998, 25:715-20. [10.1111/j.1600-051x.1998.tb02512.x](#)
 55. Bosshardt DD, Nanci A: Hertwig's epithelial root sheath, enamel matrix proteins, and initiation of cementogenesis in porcine teeth. *J Clin Periodontol.* 2004, 31:184-92. [10.1111/j.0303-6979.2004.00473.x](#)
 56. Lossdörfer S, Sun M, Götz W, Dard M, Jäger A: Enamel matrix derivative promotes human periodontal ligament cell differentiation and osteoprotegerin production in vitro. *J Dent Res.* 2007, 86:980-5. [10.1177/154405910708601012](#)
 57. Miron RJ, Bosshardt DD, Laugisch O, et al.: In vitro evaluation of demineralized freeze-dried bone allograft in combination with enamel matrix derivative. *J Periodontol.* 2013, 84:1646-54. [10.1902/jop.2013.120574](#)
 58. Miron RJ, Chandad F, Buser D, Sculean A, Cochran DL, Zhang Y: Effect of enamel matrix derivative liquid on osteoblast and periodontal ligament cell proliferation and differentiation. *J Periodontol.* 2016, 87:91-9. [10.1902/jop.2015.150389](#)
 59. Zeldich E, Koren R, Nemcovsky C, Weinreb M: Enamel matrix derivative stimulates human gingival fibroblast proliferation via ERK. *J Dent Res.* 2007, 86:41-6. [10.1177/154405910708600106](#)
 60. Bond E, Barrett S, Pragnell J: Successful treatment of non-healing wounds with Xelma(R). *Br J Nurs.* 2009, 18:1404-9. [10.12968/bjon.2009.18.22.45571](#)
 61. Bosshardt DD: Biological mediators and periodontal regeneration: a review of enamel matrix proteins at the cellular and molecular levels. *J Clin Periodontol.* 2008, 35:87-105. [10.1111/j.1600-051X.2008.01264.x](#)
 62. Koop R, Merheb J, Quirynen M: Periodontal regeneration with enamel matrix derivative in reconstructive periodontal therapy: a systematic review. *J Periodontol.* 2012, 83:707-20. [10.1902/jop.2011.110266](#)
 63. Heijl L, Heden G, Svärström G, Ostgren A: Enamel matrix derivative (EMDOGAIN) in the treatment of intrabony periodontal defects. *J Clin Periodontol.* 1997, 24:705-14. [10.1111/j.1600-051x.1997.tb00253.x](#)
 64. Froum SJ, Weinberg MA, Rosenberg E, Tarnow D: A comparative study utilizing open flap debridement with and without enamel matrix derivative in the treatment of periodontal intrabony defects: a 12-month re-entry study. *J Periodontol.* 2001, 72:25-34. [10.1902/jop.2001.72.1.25](#)
 65. Sculean A, Rathe F, Junker R, Becker J, Schwarz F, Arweiler N: [The use of Emdogain in periodontal and osseous regeneration]. *Schweiz Monatsschr Zahnmed.* 2007, 117:598-606.
 66. Heden G: A case report study of 72 consecutive Emdogain-treated intrabony periodontal defects: Clinical and radiographic findings after 1 year. *Int J Periodontics Restorative Dent.* 2000, 20:127-39.
 67. Cortellini P, Tonetti MS: Improved wound stability with a modified minimally invasive surgical technique in the regenerative treatment of isolated interdental intrabony defects. *J Clin Periodontol.* 2009, 36:157-63. [10.1111/j.1600-051X.2008.01352.x](#)
 68. Ramseier CA, Rasperini G, Batia S, Giannobile WV: Advanced reconstructive technologies for periodontal tissue repair. *Periodontol.* 2000. 2012, 59:185-202. [10.1111/j.1600-0757.2011.00432.x](#)
 69. Wikesjö UM, Nilvéus R: Periodontal repair in dogs: effect of wound stabilization on healing. *J Periodontol.* 1990, 61:719-24. [10.1902/jop.1990.61.12.719](#)
 70. Farina R, Simonelli A, Minenna L, Rasperini G, Trombelli L: Single-flap approach in combination with enamel matrix derivative in the treatment of periodontal intraosseous defects. *Int J Periodontics Restorative Dent.* 2014, 34:497-506. [10.11607/prd.2050](#)
 71. Francetti L, Del Fabbro M, Basso M, Testori T, Weinstein R: Enamel matrix proteins in the treatment of intra-bony defects. A prospective 24-month clinical trial. *J Clin Periodontol.* 2004, 31:52-9. [10.1111/j.0303-6979.2004.00437.x](#)
 72. Blomlöf J, Lindskog S: Periodontal tissue-vitality after different etching modalities. *J Clin Periodontol.* 1995, 22:464-8.
 73. Register AA, Burdick FA: Accelerated reattachment with cementogenesis to dentin, demineralized in situ. II. Defect repair. *J Periodontol.* 1976, 47:497-505. [10.1902/jop.1976.47.9.497](#)
 74. Bueno AC, Ferreira RC, Cota LO, Silva GC, Magalhães CS, Moreira AN: Comparison of different criteria for periodontitis case definition in head and neck cancer individuals. *Support Care Cancer.* 2015, 23:2599-604. [10.1007/s00520-015-2618-8](#)
 75. Cortellini P, Tonetti MS: A minimally invasive surgical technique with an enamel matrix derivative in the regenerative treatment of intra-bony defects: a novel approach to limit morbidity. *J Clin Periodontol.* 2007, 34:87-93. [10.1111/j.1600-051X.2006.01020.x](#)
 76. Jentsch HF, Rocuzzo M, Pilloni A, Kasaj A, Fimmers R, Jepsen S: Flapless application of enamel matrix derivative in periodontal retreatment: a multicentre randomized feasibility trial. *J Clin Periodontol.* 2021,

