

Ethnic Differences, Lung Cancer Risk, and Association of NRF2 Gene Polymorphism with Gemcitabine-Based Chemotherapy

Review began 07/07/2024

Review ended 07/11/2024

Published 07/18/2024

© Copyright 2024

Devika et al. This is an open access article distributed under the terms of the Creative Commons Attribution License CC-BY 4.0., which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

DOI: 10.7759/cureus.64849

Tirumalasetty Devika¹, Ganesapandian Mahalakshmi², K Mythili³, Katiboona Srinivasa Rao⁴, Suresh Kumar Srinivasamurthy⁵, Dubashi Biswajit⁶, Deepak Gopal Shewade⁷

1. Department of Pharmacology, Guntur Medical College, Guntur, IND 2. Department of Pharmacology, Nandha Medical College and Hospital, Erode, IND 3. Department of Physiology, Siddhartha Medical College, Vijayawada, IND 4. Department of Pharmacology, All India Institute of Medical Sciences, Mangalagiri, Mangalagiri, IND 5. Department of Pharmacology, RAK (Ras Al Khaimah) College of Medical Sciences, RAK Medical and Health Sciences University, Ras Al Khaimah, ARE 6. Department of Medical Oncology, Jawaharlal Institute of Postgraduate Medical Education & Research, Puducherry, IND 7. Department of Pharmacology, Jawaharlal Institute of Postgraduate Medical Education & Research, Puducherry, IND

Corresponding author: Tirumalasetty Devika, devikarani387@gmail.com

Abstract

Introduction: The cancer burden is rising every year. Lung cancer is one of the most common cancers and non-small cell lung cancer is the most common type. Chemotherapy based on platinum drugs and third-generation nucleoside anti-metabolites such as gemcitabine are used widely. Gemcitabine has a complex metabolic pathway, with many mechanisms contributing to its cytotoxicity. Derangements in the metabolic pathway genes contribute to drug resistance and toxicity with this drug. Association studies including these genetic polymorphisms in the metabolic pathway, clinical outcomes, and cancer risk reported inter-individual differences. Thus, the aim of this study was to ascertain the role of these genetic variants in South Indian cancer patients treated with gemcitabine-based therapy.

Methods: The study was done with 184 healthy volunteers for frequency establishment and 123 cancer patients were treated with gemcitabine-based chemotherapy for response and toxicity assessment. The participants were aged 18-65 years and resided in the southern states of India. DNA extraction was done from the leukocyte fraction of the blood by phenol-chloroform extraction procedures and genotyping was done by reverse transcription-polymerase chain reaction (RT-PCR) techniques to identify DNA repair gene polymorphisms. Tumor response was determined using Response evaluation criteria in solid tumors (RECIST) guidelines and toxicity using Common Terminology Criteria for Adverse Events (CTCAE), version 4.03. The patients were followed up for survival analysis.

Results: The minor allele frequency of the single nucleotide polymorphism (SNP) NRF2-617 C>A (rs6721961) in the healthy population was 12.8%. SNPs were in Hardy-Weinberg equilibrium ($p > 0.05$). Gender-based differences were not observed with the studied SNP in the healthy population and the lung cancer patients. These frequencies of NRF2 were found to be similar when compared to EUR (European) and all the South Asian subpopulations. They are significantly divergent compared to AFR (African), AMR (American), and EAS (East Asian) populations. The minor allele frequency in cancer patients was found to be 14.2% and the lung cancer risk with the SNP studied could not be detected. There was no association found with the response, toxicity, and survival among lung cancer patients.

Conclusion: NRF2, being a multifaced molecule, did not show a significant association with lung cancer risk, response, and toxicity in patients with gemcitabine-based chemotherapy.

Categories: Pharmacology, Genetics, Oncology

Keywords: south indians, rs6721961 c>a, chemotherapy, polymorphisms, gemcitabine, lung cancer, nrf2

Introduction

NRF2 is a transcriptional protein that activates numerous genes that are involved in cytoprotection [1-3]. In normal cells, this transcription factor is a cap n collar basic leucine zipper molecule. Chemical cancer susceptibility was found in NRF2 knocked-out mice. Interestingly, RNAi-mediated gene silencing of NRF2 expression inhibited abnormal cancerous growth in non-small cell lung cancer (NSCLC) [4,5]. Promoter polymorphisms in the *NRF2* gene have been identified and evaluated for their correlation with carcinogenesis [6]. Poor prognosis was found in squamous cell carcinoma (SCC) patients who were chronic smokers with *NRF2* mutation [7]. This confirms the role of NRF2 in cancer progression. The constitutive expression provides a survival advantage for the cancer cells to invade the distant organs and to undergo changes that lead to chemo-resistance under hypoxia. Increased DNA damage and decreased expression of drug transporters have been observed in cells transfected with NRF2 siRNA [8-10]. Another study showed that cisplatin sensitivity has been restored in human ovarian cancer cells by inhibiting NRF2 function [11].

How to cite this article

Devika T, Mahalakshmi G, Mythili K, et al. (July 18, 2024) Ethnic Differences, Lung Cancer Risk, and Association of NRF2 Gene Polymorphism with Gemcitabine-Based Chemotherapy. Cureus 16(7): e64849. DOI 10.7759/cureus.64849

Assay studies including transient transfection have shown that the polymorphism of NRF2 rs6721961 C>A has significantly altered its protein expression levels and thus the subsequent functions. It has been observed that this SNP is associated with a high risk of causing lung injury. This effect can probably be explained by the fact that rs6721961 C>A is involved in the self-induction of the *NRF2* gene by being present in the antioxidant response element (ARE) of the gene. To explain it further, this SNP initiates the regulation of the positive feedback loop of the activation of the *NRF2* gene and thus modifies its protein expression. As mentioned earlier, by being located at ARE-like loci of the *NRF2* gene, it reduces the binding affinity to target transcription factors. Thus, it is believed that the homozygous variant A/A can attenuate this positive feedback loop of NRF2 and decrease its transcriptional activation [12]. It was also found that lung cancer patients who harbor homozygous variant genotype of rs6721961 C>A showed high overall survival compared to the other genotypes [13].

Ethnic differences markedly influence the benefits of cancer chemotherapy [14]. India is home to a diverse population that comes from a range of ethnic backgrounds. Therefore, genetic differences across multiple ethnic groups are expected, and the southern region of India denotes a diverse population [15]. These differences form the main reasons for the varying results and have the potential to skew association study results [16]. Thus, ethnic indicators in addition to common genetic markers are employed as diagnostic and therapeutic tools to fulfill the goal of customized chemotherapy.

In this study, we aimed to establish the normative frequency of NRF2 rs6721961 C>A (-617C>A, 2q31.2 Upstream gene variant). The distribution of the aforementioned variant in the ethnic Asian population has been identified, along with its correlation with lung cancer susceptibility. Furthermore, our objectives included evaluating the commonalities and/or differences among different populations with 1000 genomes and investigating the relationship between NRF2 rs6721961 C>A and response and toxicity in patients with NSCLC.

Materials And Methods

This was a prospective, cohort study conducted at the Department of Pharmacology, Jawaharlal Institute of Postgraduate Medical Education and Research (JIPMER), Puducherry, in collaboration with the Department of Medical Oncology, Regional Cancer Centre (RCC), JIPMER, from June 2014 to June 2017. For survival analysis, the patients were followed up till August 2018 (37 months). The study was approved by the Institutional Ethics Committee (Human Studies), JIPMER, Puducherry, India (approval number: JIP/IEC/2014/4/310). Written informed consent was taken from all the participants.

A total of 307 individuals participated in the study and were divided into the control group which comprised 184 unrelated, healthy individuals aged 18–70 years, who had lived in the southern part of India for three or more generations, and the test group which comprised 123 patients with a confirmed diagnosis of metastatic stage IV lung cancer on gemcitabine-based chemotherapy, and attending the Regional Cancer Centre, JIPMER. The control group included 89 female and 95 male participants, with a mean age of 52.0±10.5 years and the test group included 47 female and 76 male patients with a mean age of 53.5±9.9. Of the 123 lung cancer patients, 47(38.2) were smokers.

Exclusion criteria for healthy volunteers were subjects with impaired liver or kidney function and pregnant and lactating women while the exclusion criteria for cancer patients were: (i) Pregnant and lactating women, (ii) Kidney dysfunction, (iii) Chronic liver diseases (serum creatinine ≥1.5 times normal value, serum transaminase, and serum bilirubin ≥2 times normal value).

Brief history, baseline data including the demographic features, Eastern Cooperative Oncology Group (ECOG) performance status, initial CT, stage of the tumor, biopsy report, and levels of biomarkers if any, and other routine investigations like hemogram and liver and renal function tests were documented. Patients received 1000 mg/m² of gemcitabine on day 1 and day 8 and carboplatin of area under the curve 5 was given in addition on day 1 for four to six cycles. Response Evaluation Criteria in Solid Tumors (RECIST) version 1.1 [17] and Common Terminology Criteria for Adverse Events (CTCAE) version 4.03 [18] guidelines were used to assess response and toxicity in lung cancer patients, respectively. Data from the 1000 Genomes Project from the International Genome Sample Resource (IGSR) (<https://www.internationalgenome.org/>) was used to compare the allele and genotype frequencies in the studied healthy population with global populations.

Statistical analysis was done using IBM SPSS Statistics for Windows, Version 19.0 (Released 2010; IBM Corp., Armonk, New York, United States) and GraphPad InStat version 3.06 (Dotmatics, Boston, Massachusetts, United States). The two-sided P-value of 0.05 was considered significant. Allele and genotype frequencies were analyzed using the direct gene count method. For the assessment of Hardy-Weinberg equilibrium and frequency comparison between the healthy South Indian volunteers and the 1000 genome populations, the Chi-Square (χ^2) test was used. Linkage disequilibrium analysis was done using the PLEM algorithm with HaploView software V 4.2 (Inform Technologies, Inc., Los Angeles, California). Association of *NRF2* gene polymorphism and response to gemcitabine-based chemotherapy as well as for case-control analysis, binary logistic regression was used. The association of *NRF2* gene polymorphism and occurrence of toxicities was analyzed using the Chi-Square (χ^2) test or Fisher's exact test. Survival analysis was done by Log-rank and

Cox regression test.

Results

The genotype and allele frequencies in the control group (n=184) were in Hardy-Weinberg equilibrium. The allele frequencies of NRF2 C>A (rs6721961) were 87.2% for C and 12.8% for A. Genotype frequencies were found to be CC 77.7%, CA 19.0%, and AA 3.3%. To establish the normative frequency, the healthy group was compared with the 1000 Genome Project populations which also included healthy participants from different ethnicities. These frequencies of NRF2 were found to be similar when compared to EUR (European) and all the South Asian subpopulations. They are significantly divergent compared to AFR (African), AMR (American), and EAS (East Asian) populations (Table 1). There were no gender-wise differences observed with this SNP (Table 2).

Present	N	Genotype frequency (%)			Allele frequency (%)		P value
		CC	CA	AA	C	A	
SI	184	77.7	19.0	3.3	87.2	12.8	Ref
AFR	661	88.7	11.2	0.1	94.3	5.7	<0.0001
AMR	347	67.4	27.4	5.2	81.1	18.9	0.011
EAS	504	57.7	35.9	6.3	75.7	24.3	<0.0001
EUR	503	76.3	22.3	1.4	87.5	12.5	0.902
SAS							
BEB	86	66.3	29.1	4.7	80.8	19.2	0.050
GIH	103	71.8	24.3	3.9	84.0	16.0	0.281
ITU	102	74.5	24.5	1.0	86.8	13.2	0.874
PJL	96	79.2	16.7	4.2	87.5	12.5	0.926
STU	102	70.6	27.5	1.9	84.3	15.7	0.333

TABLE 1: Comparison of the genotype and allele frequencies of NRF2 rs6721961 C>A polymorphism of the healthy subjects with the 1000 Genome Project populations*
N: number of subjects; SI, South Indian, AFR: African; AMR: American; EAS: East Asian; EUR: European; SAS: South Asian, BEB, Bengali in Bangladesh
GIH, Gujarat Indian in Houston; ITU; Indian Telugu in the United Kingdom; PJL: Punjabi in Lahore; STU, Srilankan Tamil in the United Kingdom
*Data from the 1000 Genomes Project from the International Genome Sample Resource (IGSR) (https://www.internationalgenome.org/)

NRF2 rs6721961	CC, n (%)	CA, n (%)	AA, n (%)	C, n (%)	A, n (%)	P value
Male (n=95)	72 (75.8)	20 (21.1)	3 (3.2)	164 (86.3)	26 (13.7)	0.587
Female (n=89)	71 (79.8)	15 (16.9)	3 (3.4)	157 (88.2)	21 (11.8)	

TABLE 2: Gender-wise genotype and allele frequency distribution of gene polymorphism in the healthy population (N = 184)

The frequencies of alleles in lung cancer patients (n=123) were found to be 85.8% for C and 14.2% for A. The genotype frequencies were CC 76.4%, CA 18.7%, and AA 4.9%. In addition, there were no gender-wise differences observed (Table 3).

Table with 7 columns: NRF2 rs6721961, CC, n (%), CA, n (%), AA, n (%), C, n (%), A, n (%), P value. Rows include Male (n=78) and Female (n=45) with their respective genotype counts and percentages.

TABLE 3: Gender-wise genotype and allele frequency distribution of gene polymorphism in the test (lung cancer) population (N= 123)

Case-control analysis did not find any risk of lung cancer susceptibility with NRF2 C>A (rs6721961) (Table 4).

Table with 6 columns: NRF2 rs6721961 C>A, Test group (N = 123), n (%), Control group (N = 184), n (%), p-value, Odds ratio, 95% CI. Rows show CC, CA, and AA genotypes with their frequencies in both groups and associated p-values and odds ratios.

TABLE 4: Case control analysis of studied polymorphism

Of the 123 lung cancer patients in the study, the response was evaluable for 95 patients. The median number of chemotherapy cycles was 4 (range, 1-6). Among them, we did not find any patients with complete response. Partial response (PR) was observed in 42 patients, stable disease (SD) in 24 patients, and progressive disease (PD) was observed in 29 patients. The objective response rate was 44.2% (PR) and the non-responders were 55.8% (SD and PD). The studied SNPs did not show any significant association with the response to gemcitabine therapy (Table 5).

Table with 7 columns: NRF2 rs6721961 C>A, Responders (n=42), Non-responders (n=53), Total (n=95), p-value, Odds ratio, 95% CI. Rows show CC, CA, and AA genotypes with their frequencies in responders, non-responders, and the total group, along with p-values and odds ratios.

TABLE 5: Association of NRF2 gene polymorphism with response to gemcitabine-based chemotherapy in lung cancer patients (N=95)

The toxicity profile was evaluated for all 123 patients in the test (lung cancer) group. They were categorized into hematological and nonhematological toxicities. Non-hematological toxicities were mild and manageable. Among hematological toxicities, anemia (62%) was found to be the highest followed by thrombocytopenia (29.2%). Grading of the hematological toxicities was done using CTCAE criteria. Except for anemia, not many patients had grade 3 and 4 toxicities. Few patients underwent blood transfusion and granulocyte-colony stimulating factor injection administration. There was no significant association observed with the studied SNP (Table 6).

Table with 9 columns: NRF2 rs6721961 C>A, Anemia (n=76), p-value, Thrombocytopenia (n=36), p-value, Leucopenia (n=20), p-value, Neutropenia (n=29), p-value. Rows show data for CC, CA, and AA genotypes.

TABLE 6: Association of NRF2 gene polymorphisms with toxicities of gemcitabine-based chemotherapy in South Indian cancer patients (N=123)

Survival analysis was performed, and the median overall survival was found to be 14 months (95%CI 11.0-16.9). Cox regression analysis was performed to find out the association of survival with the SNP. There was no significant association found between the studied SNP and survival in patients with gemcitabine-based chemotherapy. The survival curves for the studied SNP are given in Figure 1.

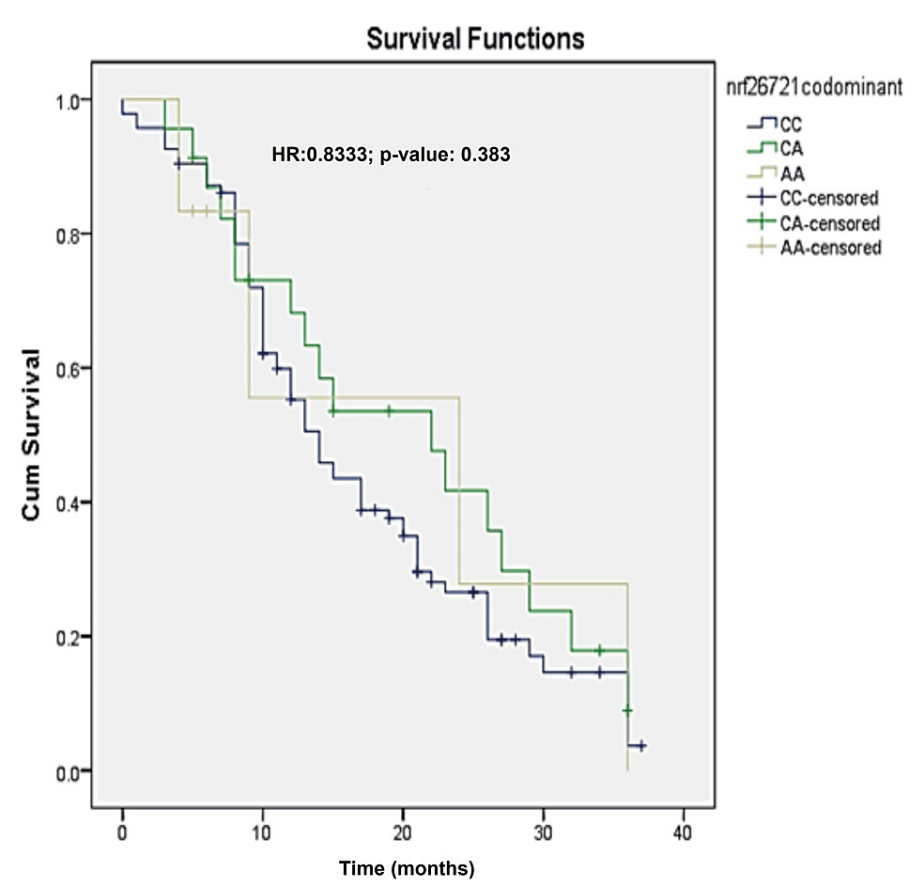


FIGURE 1: Kaplan-Meier curve of NRF2 rs 6721961 C>A for the lung cancer patients (N=123)
HR: hazard ratio; Cum Survival: cumulative survival

Discussion

The study results show that there were significant differences in allele and genotype frequencies between the study population and the 1000 Genome Project populations. Case-control analysis did not show any risk of the studied SNP with lung cancer. Additionally, there was no significant association found between the NRF2 genetic variant with response, toxicity, and survival in lung cancer patients.

Activation of NRF2 is a critical step in the process of cytoprotection as it is involved in the eradication

of reactive free radicals. At the genetic level, however, the regulatory pathways involved in activating NRF2 are not fully understood. It is well known that NRF2 activity is regulated by a protein called Keap1 (Kelch-like ECH-associated protein 1). During oxidative stress, this protein releases NRF2, which in turn causes activation of the antioxidant pathway. Thus, any alteration in this regulation can lead to increased oxidative stress that allows cancer cells to acquire different resistance mechanisms for constant proliferation. Thus, NRF2 determines the progression of cancer and resistance to chemotherapy [19].

Genotype frequencies of NRF2 rs6721961 C>A were found to be CC 77.7%, CA 19.0%, and AA 3.3%. These frequencies of NRF2 were found to be similar when compared to EUR and all the South Asian subpopulations. They are significantly divergent as compared to AFR, AMR, and EAS populations. The AFR and Taiwanese [13] population had a lower frequency of 5.7% and 7.1% compared to the present study (12.8%). In the AMR and EAS population, the frequencies are higher (18.9% and 24.3%).

Carcinogenicity may be elicited by either extrinsic or intrinsic activation of inflammatory pathways which results in immunosuppression, thereby creating a favorable background for progression [15]. NRF2 is an important factor that is involved in initiating the antioxidant pathway and protects the normal and also premalignant cells from free radical insults. Because of the continuous accumulation of damaged DNA, there is constant activation of NRF2 during tumorigenesis. Prolonged activation of NRF2 removes the reactive oxygen species along with activation of metabolic pathway genes leading to cellular reprogramming and enhanced proliferation of cancer cells. Constitutive expression of NRF2 also increases the activation of metabolizing enzymes of drugs and thus can confer resistance and poor prognosis in cancer patients [20]. SNPs in NRF2 at the molecular level affect the activation of NRF2. The SNP rs6721961 C>A has been found to be significantly associated with protein expression of NRF2 in vitro and in vivo [12]. This SNP was reported to act through antioxidant response element (ARE), activates NRF2, and thus affects the transcription of transporters and metabolizing enzymes as said earlier [21-23].

The present study could not find any association between genetic polymorphism of NRF2 with response and toxicity to gemcitabine in addition to the risk of developing lung cancer. A Japanese population-based study was done with 387 lung cancer patients to evaluate the influence of NRF2 rs6721961 C>A. Allele frequencies between males and females were found to be statistically significant (22.6% and 28.6%), and, the ratio of homozygous variant genotype in female patients was significantly higher than in males (10.8% and 2.7%, $p=0.004$). The authors further analyzed by including the smoking status as a confounding variable and found that nonsmokers were carrying a higher frequency of AA genotype (10.4% and 4.5%, respectively, $p=0.02$). In further analysis, the significance remained only for gender. Interestingly, out of 20 patients with adenocarcinoma, 16 were females and were nonsmokers. In contrast, in males, all were smoking patients [13].

Mutations in epidermal growth factor receptor (EGFR) have been found recently in their association with smoking status. The most common of them are exon 19 or 21 deletions. These derangements were observed frequently in female, nonsmoking lung cancer patients. In addition, they are most commonly observed in East Asians with adenocarcinoma [24,25]. In the current study, EGFR status was evaluated in 30 patients. Among them, 19 patients were positive for either exon19 or 21 deletions at the tyrosine kinase domain. Okano et al. found a correlation between EGFR mutations and NRF2 variant genotype AA in females with nonsmoking status and adenocarcinoma [13]. Another study has found similar results in evaluating the risk of lung cancer where the homozygous variant AA of SNP rs6721961 C>A was significantly associated with EGFR mutations [26]. Both EGFR and NRF2 follow the same pattern of expression in lung cancer patients. Thus, it can be of great value to investigate more and evaluate for any common signaling talk between them.

In the current study, the patients were followed up for 37 months and were analyzed for survival outcomes. The median overall survival was found to be 14 months (95%CI 11.0-16.9). Cox regression analysis was performed to find out the association of survival with SNPs. There was no significant association found between the studied SNP and survival in patients with gemcitabine-based chemotherapy.

NRF2 plays a double role in protecting and destroying the cells/body. Thus, its expression levels and derangements can determine the risk of cancer as well as the clinical outcomes of anticancer drugs by helping cancer cells proliferate and by causing degradation of the drugs. Our study results showed no significant association of NRF2 rs6721961 C>A with survival in lung cancer patients. In an extensive study including 304 lung specimens the immuno-histochemical analysis showed unusually high expression of NRF2, and was correlated with frequent relapse [19]. It has been found that lung cancer patients who harbor homozygous variant genotype rs6721961 C>A showed high overall survival compared to the other genotypes. Lung cancer patients from stages I to IV were included for analysis and it was found that the wild type and heterozygous genotypes were not correlated with survival. Further, stage I alone, adenocarcinoma, squamous, and large cell types were included for analysis which also resulted in a favorable prognosis. The authors reported that all the patients with variant genotype AA and adenocarcinoma had survived over 1000 days after surgical resection without any follow-up treatment [13].

The limitation of this study is that the sample size was small to extrapolate the results. The plasma concentration of drug and its metabolite were not estimated, which could give additional understanding in

evaluating the results with the studied polymorphism. NRF2 has various roles and lung cancer is a multifactorial disease with many facets; thus, future studies with multiple biomarkers may be considered in analyzing the better treatment option.

Conclusions

The present study findings showed remarkable heterogeneity with respect to the genotype frequency of the South Indian population compared to other global populations. We could not get a significant association of the studied polymorphism with lung cancer risk, response, toxicity, and overall survival in patients on gemcitabine-based chemotherapy. *NRF2* has multifarious roles, the polymorphisms of which need to be evaluated on a larger scale to understand the intricate functions in response to gemcitabine-based chemotherapy.

Additional Information

Author Contributions

All authors have reviewed the final version to be published and agreed to be accountable for all aspects of the work.

Concept and design: Tirumalasetty Devika, Katiboina Srinivasa Rao, Suresh Kumar Srinivasamurthy, Deepak Gopal Shewade, Dubashi Biswajit

Acquisition, analysis, or interpretation of data: Tirumalasetty Devika, Katiboina Srinivasa Rao, Ganesapandian Mahalakshmi, K Mythili, Deepak Gopal Shewade, Dubashi Biswajit

Drafting of the manuscript: Tirumalasetty Devika, Katiboina Srinivasa Rao, Ganesapandian Mahalakshmi, Suresh Kumar Srinivasamurthy, K Mythili, Deepak Gopal Shewade, Dubashi Biswajit

Critical review of the manuscript for important intellectual content: Tirumalasetty Devika, Katiboina Srinivasa Rao, Ganesapandian Mahalakshmi, Suresh Kumar Srinivasamurthy, K Mythili, Deepak Gopal Shewade, Dubashi Biswajit

Disclosures

Human subjects: Consent was obtained or waived by all participants in this study. Institute Ethics Committee (Human Studies), JIPMER, Puducherry, India issued approval JIP/IEC/2014/4/310 dated June 27, 2014. **Animal subjects:** All authors have confirmed that this study did not involve animal subjects or tissue.

Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

References

1. Itoh K, Chiba T, Takahashi S, et al.: An Nrf2/small Maf heterodimer mediates the induction of phase II detoxifying enzyme genes through antioxidant response elements. *Biochem Biophys Res Commun*. 1997, 236:313-22. [10.1006/bbrc.1997.6943](https://doi.org/10.1006/bbrc.1997.6943)
2. Rushmore TH, Kong AN: Pharmacogenomics, regulation and signaling pathways of phase I and II drug metabolizing enzymes. *Curr Drug Metab*. 2002, 3:481-90. [10.2174/1389200023337171](https://doi.org/10.2174/1389200023337171)
3. Nguyen T, Yang CS, Pickett CB: The pathways and molecular mechanisms regulating Nrf2 activation in response to chemical stress. *Free Radic Biol Med*. 2004, 37:433-41. [10.1016/j.freeradbiomed.2004.04.033](https://doi.org/10.1016/j.freeradbiomed.2004.04.033)
4. Fahey JW, Kensler TW: Role of dietary supplements/nutraceuticals in chemoprevention through induction of cytoprotective enzymes. *Chem Res Toxicol*. 2007, 20:572-6. [10.1021/tx7000459](https://doi.org/10.1021/tx7000459)
5. Kensler TW, Wakabayashi N, Biswal S: Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. *Annu Rev Pharmacol Toxicol*. 2007, 47:89-116. [10.1146/annurev.pharmtox.46.120604.141046](https://doi.org/10.1146/annurev.pharmtox.46.120604.141046)
6. Arisawa T, Tahara T, Shibata T, et al.: Nrf2 gene promoter polymorphism and gastric carcinogenesis. *Hepatogastroenterology*. 2008, 55:750-4.
7. Shibata T, Ohta T, Tong KI, et al.: Cancer related mutations in NRF2 impair its recognition by Keap1-Cul3 E3 ligase and promote malignancy. *Proc Natl Acad Sci U S A*. 2008, 9:13568-73. [10.1073/pnas.080626810](https://doi.org/10.1073/pnas.080626810)
8. Semenza GL: Targeting HIF-1 for cancer therapy. *Nat Rev Cancer*. 2003, 3:721-32. [10.1038/nrc1187](https://doi.org/10.1038/nrc1187)
9. Zhou J, Schmid T, Schnitzer S, Brüne B: Tumor hypoxia and cancer progression. *Cancer Lett*. 2006, 237:10-21. [10.1016/j.canlet.2005.05.028](https://doi.org/10.1016/j.canlet.2005.05.028)
10. Homma S, Ishii Y, Morishima Y, et al.: Nrf2 enhances cell proliferation and resistance to anticancer drugs in human lung cancer. *Clin Cancer Res*. 2009, 15:3423-32. [10.1158/1078-0432.CCR-08-2822](https://doi.org/10.1158/1078-0432.CCR-08-2822)
11. Cho JM, Manandhar S, Lee HR, Park HM, Kwak MK: Role of the Nrf2-antioxidant system in cytotoxicity mediated by anticancer cisplatin: implication to cancer cell resistance. *Cancer Lett*. 2008, 260:96-108. [10.1016/j.canlet.2007.10.022](https://doi.org/10.1016/j.canlet.2007.10.022)

12. Marzec JM, Christie JD, Reddy SP, et al.: Functional polymorphisms in the transcription factor NRF2 in humans increase the risk of acute lung injury. *FASEB J*. 2007, 21:2237-46. [10.1096/fj.06-7759com](https://doi.org/10.1096/fj.06-7759com)
13. Okano Y, Nezu U, Enokida Y, et al.: SNP (-617C>A) in ARE-like loci of the NRF2 gene: a new biomarker for prognosis of lung adenocarcinoma in Japanese non-smoking women. *PLoS One*. 2013, 8:e73794. [10.1371/journal.pone.0073794](https://doi.org/10.1371/journal.pone.0073794)
14. Watson J: Oxidants, antioxidants and the current incurability of metastatic cancers . *Open Biol*. 2013, 3:120144. [10.1098/rsob.120144](https://doi.org/10.1098/rsob.120144)
15. Todoric J, Antonucci L, Karin M: Targeting inflammation in cancer prevention and therapy . *Cancer Prev Res (Phila)*. 2016, 9:895-905. [10.1158/1940-6207.CAPR-16-0209](https://doi.org/10.1158/1940-6207.CAPR-16-0209)
16. Cheng H, Xu X, Costa DB, Powell CA, Halmos B: Molecular testing in lung cancer: the time is now . *Curr Oncol Rep*. 2010, 12:335-48. [10.1007/s11912-010-0118-z](https://doi.org/10.1007/s11912-010-0118-z)
17. Eisenhauer EA, Therasse P, Bogaerts J, et al.: New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur J Cancer*. 2009, 45:228-47. [10.1016/j.ejca.2008.10.026](https://doi.org/10.1016/j.ejca.2008.10.026)
18. Common Terminology Criteria for Adverse Events (CTCAE) Version 4.0 . U.S. Department of Health and Human Services, Washington, DC; 2009. https://evs.nci.nih.gov/ftp1/CTCAE/CTCAE_4.03/CTCAE_4.03_2010-06-14_QuickReference_8.5x11.pdf.
19. Solis LM, Behrens C, Dong W, et al.: Nrf2 and Keap1 abnormalities in non-small cell lung carcinoma and association with clinicopathologic features. *Clin Cancer Res*. 2010, 16:3743-53. [10.1158/1078-0432.CCR-09-3352](https://doi.org/10.1158/1078-0432.CCR-09-3352)
20. Sporn MB, Liby KT: NRF2 and cancer: the good, the bad and the importance of context . *Nat Rev Cancer*. 2012, 12:564-71. [10.1038/nrc3278](https://doi.org/10.1038/nrc3278)
21. Shen G, Kong AN: Nrf2 plays an important role in coordinated regulation of Phase II drug metabolism enzymes and Phase III drug transporters. *Biopharm Drug Dispos*. 2009, 30:345-55. [10.1002/bdd.680](https://doi.org/10.1002/bdd.680)
22. Singh A, Wu H, Zhang P, Happel C, Ma J, Biswal S: Expression of ABCG2 (BCRP) is regulated by Nrf2 in cancer cells that confers side population and chemoresistance phenotype. *Mol Cancer Ther*. 2010, 9:2365-76. [10.1158/1535-7163.MCT-10-0108](https://doi.org/10.1158/1535-7163.MCT-10-0108)
23. Ishikawa T, Shinkai M, Kaneko T: Nrf2 gene as a double-edged sword: clinical relevance of its genetic polymorphisms. *Integr Mol Med*. 2016, 3:688-93.
24. Toyooka S, Matsuo K, Shigematsu H, et al.: The impact of sex and smoking status on the mutational spectrum of epidermal growth factor receptor gene in non small cell lung cancer. *Clin Cancer Res*. 2007, 13:5763-8. [10.1158/1078-0432.CCR-07-0216](https://doi.org/10.1158/1078-0432.CCR-07-0216)
25. Ladanyi M, Pao W: Lung adenocarcinoma: guiding EGFR-targeted therapy and beyond . *Mod Pathol*. 2008, 21:S16-22. [10.1038/modpathol.3801018](https://doi.org/10.1038/modpathol.3801018)
26. Suzuki T, Shibata T, Takaya K, et al.: Regulatory nexus of synthesis and degradation deciphers cellular Nrf2 expression levels. *Mol Cell Biol*. 2013, 33:2402-12. [10.1128/MCB.00065-13](https://doi.org/10.1128/MCB.00065-13)