Cureus

Received 09/06/2023 Review began 10/01/2023 Review ended 11/13/2023 Published 11/17/2023

#### © Copyright 2023

Nair et al. This is an open access article distributed under the terms of the Creative Commons Attribution License CC-BY 4.0., which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

# Aldose Reductase and Protein Glycation Inhibitory Activity of Dark Chocolate-Assisted Zinc Oxide Nanoparticles

Vedha R. Nair $^1$ , Geetha R V Sr. $^1$ , Parameswari R P $^2$ 

1. Microbiology, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai, IND 2. Pharmacology, Centre for Transdisciplinary Research, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technial Sciences, Saveetha University, Chennai, IND

Corresponding author: Geetha R V Sr., geetha@saveetha.com

## Abstract

### Introduction

One of the most common health issues that the global population is dealing with is the associated complications of diabetes, which encompasses cataracts, peripheral neuropathy, vascular damage, impaired wound healing, retinal issues, and arterial wall stiffening. The present study is aimed to evaluate the effect of dark chocolate and its assisted zinc oxide nanoparticles against diabetes-associated complications.

#### Materials and methods

Zinc oxide nanoparticles were synthesized using commercially dark chocolate (DC-ZnO NP). The synthesized DC-ZnO NPs were evaluated against recombinant aldose reductase (AR) activity and the formation of advanced glycation end products (AGEs). Aminoguanidine and gallic acid were used as reference standards for AGE assay and sorbitol accumulation inhibition, respectively.

#### Results

The results of the present study showed that green synthesized DC-ZnO NP had a significant dosedependent inhibitory activity on both AR and AGEs. The inhibitory activity was compared to that of quercetin and aminoguanidine, respectively.

#### Conclusion

Targeting the endogenous antioxidant systems like AGEs and AR enzymes seems to provide a promising therapeutic approach, thus concluding that ZnO-NP could be a promising agent for treating diabetes-related complications such as diabetic retinopathy, diabetic nephropathy, and diabetic neuropathy that provide grounds for further clinical investigations and trials.

**Categories:** Endocrinology/Diabetes/Metabolism, Integrative/Complementary Medicine, Therapeutics **Keywords:** zinc oxide, green synthesis, nanoparticles, protein glycation, aldose reductase, diabetes mellitus

## Introduction

Diabetes mellitus is a broad term encompassing various metabolic conditions, with chronic high blood sugar levels being the most prevalent manifestation. The primary culprits behind diabetes are either insufficient insulin production, impaired insulin function, or a combination of both. In its initial phases, diabetes poses a substantial risk for atherosclerotic vascular disease and coronary heart disease [1]. Diabetes patients have twice the rate of heart disease as the general population. Other associated complications of diabetes are retinopathy, neuropathy, etc. [2]. The global prevalence of diabetes has risen considerably in recent decades, and it is anticipated to continue to rise in the future. Obesity and advancing age are both linked to an increased incidence of diabetes, regardless of race or ethnicity [3]. The stimulation of the sorbitol tract induces non-enzymatic protein glycation, which leads to basement membrane thickening and endothelial cell proliferation. This membrane thickening leads to increased vascular resistance hence reducing tissue perfusion and causing nerve hypoxia [4]. The basis for aldose reductase (AR) inhibition in diabetes is the high AR pathway activity in the peripheral nerve and other tissues that are susceptible to complications related to diabetes, and its activation by hyperglycemia [5].

With a high flavonoid concentration, cocoa beans and derivatives, such as cocoa powder and chocolate, are one of the greatest sources of antioxidants. Several studies have shown that flavonoids present in cocoa have antioxidant and anti-diabetic characteristics that affect glucose metabolism [6]. Many studies have found that flavanol-rich cocoa products can enhance endothelial function, decrease platelet reactivity, increase insulin sensitivity, and lower systolic and diastolic blood pressure whether consumed acutely or on a regular basis [7,8].

#### How to cite this article

Nair V R, R V G, R P P (November 17, 2023) Aldose Reductase and Protein Glycation Inhibitory Activity of Dark Chocolate-Assisted Zinc Oxide Nanoparticles. Cureus 15(11): e48953. DOI 10.7759/cureus.48953

Zinc plays a crucial role in various physiological processes, particularly in the regulation of carbohydrate metabolism [9]. It is highly concentrated in the beta cells of the pancreas, where it is involved in the synthesis, storage, crystallization, and release of insulin [10]. A deficiency in zinc is known to exacerbate kidney damage caused by diabetes. Several clinical observations suggest that zinc may have a preventive effect on the development of diabetic heart and kidney problems [11]. Zinc oxide nanoparticles (ZnONPs) represent a novel method for delivering zinc and hold promise for treating various diseases, including diabetes [12]. The development of a zinc-based medication could be beneficial for managing diabetes and its complications, as preclinical trials have shown positive effects of zinc supplementation [13]. Therefore, the aim of this study is to evaluate the AR and protein glycation inhibitory activity of dark chocolate and its assisted zinc oxide nanoparticles.

## **Materials And Methods**

### Preparation of dark chocolate extract

About 100 mg of dark chocolate, obtained from a commercial source, was dissolved in 100 ml of distilled water and heated to  $50^{\circ}$ C. Subsequently, the resulting mixture underwent filtration, initially using Whatman filter paper no. 1 and then through a vacuum filter with a pore size of 0.2 µm. The final filtrate was preserved in a cool, dry location for future utilization.

### Synthesis of zinc oxide nanoparticles using dark chocolate

About 25 ml of zinc acetate dihydrate (Zn(NO  $_3$ ) $_2$ ·2H $_2$ O) were combined with 4 ml of dark chocolate extract and stirred on a magnetic stirrer at 60°C for two hours. After the reaction was complete, the mixture was cooled to 25°C and then centrifuged at 10,000 rpm for 10 minutes. The liquid portion was discarded, and the remaining solid was washed three times with distilled water. It was then transferred to a clean Petri dish, dried in an oven at 90°C, and subsequently ground into a fine powder using a mortar and pestle. This powder was heated at 500°C for two hours to eliminate any impurities through a process called calcination. The resulting annealed powder was stored in a sealed glass container and was utilized for various biological applications.

### **Determination of AR inhibition**

A total of 531 µL of 0.1 M potassium buffer (pH 7.0), 90 µL of nicotinamide adenine dinucleotide phosphate hydrogen (NADPH) solution (1.6 mM in potassium buffer), 90 µL of recombinant human AR (6.5 U/mg) (SRP6371-100UG, Sigma-Aldrich, St. Louis, United States), 90 µL of ammonium sulfate solution (4 M in potassium buffer), and 90 µL of DL-glyceraldehyde (25 mM in potassium buffer) were mixed with 9 µL of different concentrations of ZnO nanoparticles (5, 10, 20, 40, 80, and 160 µL) in a cuvette, and the activity of AR was assessed spectrophotometrically by measuring the decrease in NADPH absorbance at 340 nm for three minutes using a spectrophotometer (Biotek Synergy H4 multimode reader, Thermo Fisher Scientific Inc., Waltham, United States). Quercetin was used as a positive control. The inhibition of AR (%) was calculated using the following equation:  $(1 - (\triangle A sample/min) - (\triangle A blank/min))/(\triangle A control/min) - (\triangle A blank/min)) \times 100\%$ , where  $\triangle A$  sample/min is the decrease in absorbance over three minutes with reaction solution, test sample, and substrate, and  $\triangle A$  control/min without the test sample [14].

### Advanced glycation end-product assay

Advanced glycation end products (AGEs) [15] are formed by non-enzymatic glycosylation of proteins that enhance vascular permeability in both micro and macro vascular structures by binding to specific macrophage receptors. The ZnO nanoparticles were evaluated for their activity on AGEs formation. AGE reaction mixture was constituted as follows; 1 mg/mL bovine serum albumin (BSA) in 50 mM sodium phosphate buffer (pH 7.4) and 0.02% sodium benzoate into 0.2 M fructose and 0.2 M glucose. The reaction mixture (2.75 mL) was treated with different volumes of ZnO nanoparticles (5, 10, 20, 40, 80, and 160 µg/ml). Amino guanidine was used as a positive control. After incubating at 37°C for three days, the fluorescence intensity of the reaction was determined at excitation and emission wavelengths of 350 nm and 450 nm, respectively, using a Biotek synergy multi-mode reader (Agilent, Santa Clara, United States). The percentage activity was calculated with respect to solvent control.

## **Results**

### Sorbitol accumulation inhibition activity of DC-ZnO NP

In the pathogenesis of diabetic complications, it is thought that an important role is played by increased oxidative stress, as supported by the increased levels of oxidized DNA, lipids, and proteins [16]. Increased AR activity has been linked to the development of intracellular sorbitol accumulation, which was linked to a number of secondary complications associated with diabetes. Inhibiting aldolase reductase may be a useful tactic for delaying or preventing some diabetic problems [17]. Figure *1* shows that there was a dose-dependent increase in the AR inhibition by dark chocolate-mediated ZnONP which was comparable to the standard quercetin.

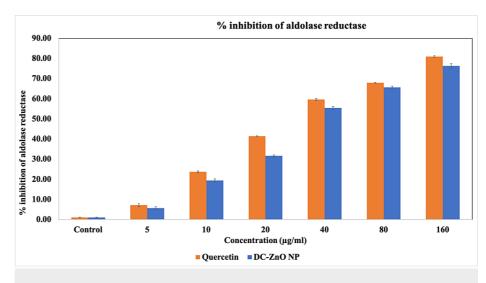


FIGURE 1: Effect of dark chocolate-mediated zinc oxide nanoparticles (DC-ZnO NP) on Aldolase reductase enzyme activity

#### Inhibitory activity of DC-ZnO NPs on advanced glycation end products

The formation of AGEs is a process that can result in the production of reactive oxygen species (ROS). Additionally, when AGEs interact with their receptors, known as receptor for AGEs, it can also lead to the generation of ROS. These AGEs tend to accumulate in various tissues affected by diabetic complications, such as the kidneys, retina, and atherosclerotic plaques. The glycation of proteins disrupts their normal functions by affecting molecular interactions, altering enzymatic activity, reducing their degradation capacity, and interfering with receptor recognition [15]. Figure 2 in the study indicates that the presence of dark chocolate-mediated zinc oxide nanoparticles led to a dose-dependent reduction in the formation of AGEs. This suggests that these nanoparticles have the potential to effectively mitigate complications associated with diabetes.

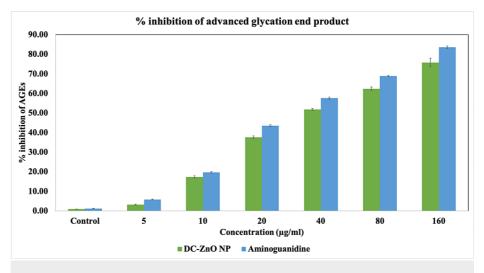


FIGURE 2: Inhibitory effect of dark chocolate mediated zinc oxide nanoparticles (DC-ZnO NP) on advanced glycation end product (AGE)

## **Discussion**

The current study's findings suggest that inhibiting the AR enzyme and AGEs can decrease the risk of diabetes-related complications. In vivo experiments provide strong evidence that high blood sugar levels contribute to oxidative stress by generating ROS, leading to acute dysfunction of the blood vessel endothelium in diabetic patients [16]. The research proposes that excessive production of superoxide due to mitochondrial electron transport triggered by hyperglycemia plays a central role in initiating various pathways responsible for the development of complications associated with diabetes. These pathways include non-enzymatic glycation, hexosamine pathway activation, protein kinase activity, and

mitochondrial respiratory chain [17-19].

Excessive generation of ROS disrupts cellular signaling pathways and overall cellular balance, leading to instability in pancreatic beta cells and increased insulin resistance. This resistance to insulin is a crucial factor in the development of elevated blood sugar levels, contributing to complications in both small and large blood vessels associated with diabetes [20, 21]. In a study conducted by Asri and colleagues, administering zinc oxide nanoparticles (ZnO-NP) at a dose of 3 mg/kg/day for eight weeks resulted in significant reductions. Specifically, it halved brain natriuretic peptide levels, decreased the atherogenic index by 80%, and lowered serum cholesterol levels in diabetic rats [22]. Another study by Nazarizadeh and his team concluded that ZnO-NP played a role in restoring catalase activity in both serum and erythrocytes of diabetic rats [23]. Furthermore, results from oral glucose tolerance tests suggested that ZnO-NP could potentially improve glucose tolerance in experimental diabetes [24]. The extent of this effect varied in a dose-dependent manner, with concentrations of 10 mg/kg/day reducing it to approximately 40% to 70% compared to the diabetic control group [25].

Abd El-Khalik et al. conducted a study proposing the potential of ZnO-NPs to mitigate diabetic nephropathy progression by influencing the interplay between autophagy and the Nrf2/TXNIP/NLRP3 inflammasome signaling pathways [26]. In an earlier investigation, Ian et al. explored the therapeutic effects of ZnO-NPs derived from Aquilegia pubiflora. Administered at a dosage of 200mg/ml, these nanoparticles exhibited significant inhibition of alpha-glucosidase and alpha-amylase, underscoring their anti-diabetic properties [27]. Another study examined the use of green-synthesized ZnO-NPs, combined with natural polymers and other therapeutic agents, to address delayed wound healing in diabetic individuals. This approach demonstrated efficient antibacterial activity against diabetic wound infections, accelerating the woundhealing process [28]. Additionally, ZnO-NPs synthesized from Morus indica significantly inhibited methylglyoxal-mediated glycation of BSA, showing a dose-dependent inhibition of AGEs formation [29]. Recent findings have also shown that ZnO-NPs markedly prevent AGE formation in diabetic mice by acting on hypoglycemic and antihyperlipidemic pathways [30,31]. Furthermore, ZnO-NPs not only prevent AGE formation but also inhibit protein structure changes, thereby hindering AGE formation. Moreover, the capacity of ZnO-NPs to mitigate oxidative stress, particularly by targeting advanced glycation end products, not only holds potential benefits for managing diabetes-related complications but also shows promise in addressing neurodegenerative disorders and other degenerative conditions [32].

The glycation reaction sets off a harmful cycle as it interacts with free radicals and carbonyl species, ultimately resulting in the generation of AGEs. This process significantly contributes to the onset and progression of various diseases. In light of the detrimental impact of AGEs, numerous anti-glycating agents have been identified [32]. Nevertheless, there is an urgent need to explore more effective agents to address this serious clinical issue. According to the given findings, zinc oxide, renowned for its antioxidant properties, has demonstrated potential in inhibiting AGE formation [26-31]. However, it is crucial to adopt a systematic and mechanistic approach before the significant role of ZnO-NPs can be effectively harnessed in the medical field.

### Limitations

The initial preclinical evaluation provides preliminary data indicating the impact of synthesized nanoparticles on complications related to diabetes. Nevertheless, additional research is essential, involving both in vitro studies using cell lines and in vivo investigations focused on assessing toxicity and efficacy. Further studies are required to establish and validate the potential of the synthesized nanoparticles for the treatment of complications associated with diabetes.

### Conclusions

The increasing global prevalence of diabetes mellitus and its associated complications necessitates the search for effective treatments. Nanotechnology and nanomedicine advancements have opened up possibilities for biomedical applications and disease management, including the use of materials like ZnO-NP. In this study, we found that dark chocolate ZnO-NP, synthesized through a green process, exhibited a dose-dependent inhibitory effect on both AR and the formation of protein glycation end products. By targeting the body's natural antioxidant systems, such as AGEs and the AR enzyme, this approach appears to offer a promising therapeutic avenue. In conclusion, ZnO-NP shows great promise as a treatment agent for diabetes and its associated complications, warranting further investigation through clinical trials.

## **Additional Information**

### **Author Contributions**

All authors have reviewed the final version to be published and agreed to be accountable for all aspects of the work.

Concept and design: Geetha R V Sr., Parameswari R P

Acquisition, analysis, or interpretation of data: Geetha R V Sr., Vedha R. Nair, Parameswari R P

Drafting of the manuscript: Geetha R V Sr., Vedha R. Nair, Parameswari R P

**Critical review of the manuscript for important intellectual content:** Geetha R V Sr., Vedha R. Nair, Parameswari R P

Supervision: Geetha R V Sr., Parameswari R P

#### Disclosures

Human subjects: All authors have confirmed that this study did not involve human participants or tissue. Animal subjects: All authors have confirmed that this study did not involve animal subjects or tissue. Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: Payment/services info: All authors have declared that no financial support was received from any organization for the submitted work. Financial relationships: All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. Other relationships: All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

#### Acknowledgements

The authors would like to thank Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Saveetha University for providing research laboratory facilities to carry out the study.

## References

- Kerner W, Brückel J: Definition, classification and diagnosis of diabetes mellitus. Exp Clin Endocrinol Diabetes. 2014, 122:384-6. 10.1055/s-0034-1366278
- Bailes BK: Diabetes mellitus and its chronic complications. AORN J. 2002, 76:266-76. 10.1016/s0001-2092(06)61065-x
- Deshpande AD, Harris-Hayes M, Schootman M: Epidemiology of diabetes and diabetes-related complications. Phys Ther. 2008, 88:1254-64. 10.2522/ptj.20080020
- Pierzchala K: Contemporary views on the pathogenesis of diabetic neuropathy (Article in Polish). Wiad Lek. 1998, 51 Suppl 2:30-4.
- Greene DA, Arezzo JC, Brown MB: Effect of aldose reductase inhibition on nerve conduction and morphometry in diabetic neuropathy. Zenarestat Study Group. Neurology. 1999, 53:580-91. 10.1212/wnl.53.3.580
- Shah SR, Alweis R, Najim NI, et al.: Use of dark chocolate for diabetic patients: a review of the literature and current evidence. J Community Hosp Intern Med Perspect. 2017, 7:218-21. 10.1080/20009666.2017.1361293
- Erdman JW Jr, Carson L, Kwik-Uribe C, Evans EM, Allen RR: Effects of cocoa flavanols on risk factors for cardiovascular disease. Asia Pac J Clin Nutr. 2008, 17:284-7.
- Grassi D, Necozione S, Lippi C, et al.: Cocoa reduces blood pressure and insulin resistance and improves endothelium-dependent vasodilation in hypertensives. Hypertension. 2005, 46:398-405. 10.1161/01.HYP.0000174990.46027.70
- Chimienti F, Devergnas S, Favier A, Seve M: Identification and cloning of a beta-cell-specific zinc transporter, ZnT-8, localized into insulin secretory granules. Diabetes. 2004, 53:2330-7. 10.2337/diabetes.53.9.2330
- Capdor J, Foster M, Petocz P, Samman S: Zinc and glycemic control: a meta-analysis of randomised placebo controlled supplementation trials in humans. J Trace Elem Med Biol. 2013, 27:137-42. 10.1016/j.jtemb.2012.08.001
- 11. Li B, Tan Y, Sun W, Fu Y, Miao L, Cai L: The role of zinc in the prevention of diabetic cardiomyopathy and nephropathy. Toxicol Mech Methods. 2013, 23:27-33. 10.3109/15376516.2012.735277
- Siddiqui SA, Or Rashid MM, Uddin MG, Robel FN, Hossain MS, Haque MA, Jakaria M: Biological efficacy of zinc oxide nanoparticles against diabetes: a preliminary study conducted in mice. Biosci Rep. 2020, 40:BSR20193972. 10.1042/BSR20193972
- Tang KS: The current and future perspectives of zinc oxide nanoparticles in the treatment of diabetes mellitus. Life Sci. 2019, 239:117011. 10.1016/j.lfs.2019.117011
- Akileshwari C, Muthenna P, Nastasijević B, Joksić G, Petrash JM, Reddy GB: Inhibition of Aldose Reductase by Gentiana lutea extracts. Exp Diabetes Res. 2012, 2012:147965. 10.1155/2012/147965
- Harris CS, Beaulieu LP, Fraser MH, et al.: Inhibition of advanced glycation end product formation by medicinal plant extracts correlates with phenolic metabolites and antioxidant activity. Planta Med. 2011, 77:196-204. 10.1055/s-0030-1250161
- Ceriello A: Oxidative stress and diabetes-associated complications. Endocr Pract. 2006, 12 Suppl 1:60-2. 10.4158/EP.12.S1.60
- 17. Brownlee M: Biochemistry and molecular cell biology of diabetic complications . Nature. 2001, 414:813-20. 10.1038/414813a
- Genuth S, Eastman R, Kahn R, et al.: Implications of the United kingdom prospective diabetes study . Diabetes Care. 2003, 26 Suppl 1:S28-32. 10.2337/diacare.26.2007.s28
- Vinik AI, Richardson DW: Implications of the diabetes control and complications trial for persons with noninsulin-dependent diabetes mellitus. South Med J. 1997, 90:268-82.
- 20. Andersen K, Hurlen M, Arnesen H, Seljeflot I: Aspirin non-responsiveness as measured by PFA-100 in

patients with coronary artery disease. Thromb Res. 2002, 108:37-42. 10.1016/s0049-3848(02)00405-x

- Bhatti JS, Sehrawat A, Mishra J, et al.: Oxidative stress in the pathophysiology of type 2 diabetes and related complications: current therapeutics strategies and future perspectives. Free Radic Biol Med. 2022, 184:114-34. 10.1016/j.freeradbiomed.2022.03.019
- Asri-Rezaei S, Dalir-Naghadeh B, Nazarizadeh A, Noori-Sabzikar Z: Comparative study of cardio-protective effects of zinc oxide nanoparticles and zinc sulfate in streptozotocin-induced diabetic rats. J Trace Elem Med Biol. 2017, 42:129-41. 10.1016/j.jtemb.2017.04.013
- Nazarizadeh A, Asri-Rezaie S: Comparative study of antidiabetic activity and oxidative stress induced by zinc oxide nanoparticles and zinc sulfate in diabetic rats. AAPS PharmSciTech. 2016, 17:834-43. 10.1208/s12249-015-0405-y
- Alkaladi A, Abdelazim AM, Afifi M: Antidiabetic activity of zinc oxide and silver nanoparticles on streptozotocin-induced diabetic rats. Int J Mol Sci. 2014, 15:2015-23. 10.3390/ijms15022015
- El-Gharbawy RM, Emara AM, Abu-Risha SE: Zinc oxide nanoparticles and a standard antidiabetic drug restore the function and structure of beta cells in type-2 diabetes. Biomed Pharmacother. 2016, 84:810-20. 10.1016/j.biopha.2016.09.068
- Abd El-Khalik SR, Nasif E, Arakeep HM, Rabah H: The prospective ameliorative role of zinc oxide nanoparticles in STZ-induced diabetic nephropathy in rats: mechanistic targeting of autophagy and regulating Nrf2/TXNIP/NLRP3 inflammasome signaling. Biol Trace Elem Res. 2022, 200:1677-87. 10.1007/s12011-021-02773-4
- Jan H, Shah M, Andleeb A, et al.: Plant-based synthesis of zinc oxide nanoparticles (ZnO-NPS) using aqueous leaf extract of Aquilegia pubiflora: their antiproliferative activity against HepG2 cells inducing reactive oxygen species and other in vitro properties. Oxid Med Cell Longev. 2021, 2021;4786227.
  10.1155/2021/4786227
- Steffy K, Shanthi G, Maroky AS, Selvakumar S: Enhanced antibacterial effects of green synthesized ZnO NPs using Aristolochia indica against multi-drug resistant bacterial pathogens from diabetic foot ulcer. J Infect Public Health. 2018, 11:463-71. 10.1016/j.jiph.2017.10.006
- Ashraf JM, Ansari MA, Fatma S, et al.: Inhibiting effect of zinc oxide nanoparticles on advanced glycation products and oxidative modifications: a potential tool to counteract oxidative stress in neurodegenerative diseases. Mol Neurobiol. 2018, 55:7438-52. 10.1007/s12035-018-0935-x
- Anandan S, Mahadevamurthy M, Ansari MA, et al.: Biosynthesized ZnO-NPs from Morus indica attenuates methylglyoxal-induced protein glycation and RBC damage: in-vitro, in-vivo and molecular docking study. Biomolecules. 2019, 9:882. 10.3390/biom9120882
- Meer B, Andleeb A, Iqbal J, et al.: Bio-assisted synthesis and characterization of zinc oxide nanoparticles from Lepidium sativum and their potent antioxidant, antibacterial and anticancer activities. Biomolecules. 2022, 12:855. 10.3390/biom12060855
- Sengani M, Chakraborty S, Balaji MP, et al.: Anti-diabetic efficacy and selective inhibition of methyl glyoxal, intervention with biogenic zinc oxide nanoparticle. Environ Res. 2023, 216:114475. 10.1016/j.envres.2022.114475