

Exploring the Therapeutic Potential of Apigenin in Obesity-Associated Fibrinolytic Dysfunction: Insights From an Animal Study

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Abstract

Introduction: Obesity (Obe) is a chronic metabolic disorder usually complicated by impaired fibrinolytic activity. Apigenin (Api) is one of the flavonoids that have anti-adiposity effects. This study aimed to explore the therapeutic potential of Api in high-fat diet (HFD)-induced obese rats.

Methods: Twenty-four Wistar adult male rats were randomly allocated into control group, supplemented with a normal pellet diet (NPD); Api group, supplemented with Api (10 mg/kg) for eight weeks; Obe group, obesity was induced by feeding HFD for eight weeks; and Obe/Api group, obese rats supplemented with Api for eight weeks. Body mass index (BMI), homeostatic model assessment of insulin resistance (HOMA-IR), tumor necrosis factor- α (TNF- α), malondialdehyde (MDA), total superoxide dismutase (t-SOD) activity, and plasminogen activator inhibitor-1 (PAI-1) were measured.

Results: Compared to the control group, Obe group exhibited a significant increase in BMI, HOMA-IR, TNF- α , MDA, and PAI-1. These results were also associated with a significant decrease in serum t-SOD activity. Supplementation of Api alleviated the measured deteriorated parameters and ameliorated visceral adiposity in obese rats.

Conclusion: This study provides compelling evidence regarding a promising role for Api in ameliorating the impairment of fibrinolytic activity in an Obe animal model. The observed effects are likely mediated through Api's anti-obesity properties, as well as its indirect modulation of PAI-1, oxidative stress, and inflammation. Future clinical studies are recommended that may make benefit of the preclinical therapeutic use of apigenin in obesity-associated fibrinolytic dysfunctions.

Categories: Therapeutics

Keywords: apigenin, plasminogen activator inhibitor-1, fibrinolysis, inflammation, oxidative stress, obesity

Introduction

The global prevalence of obesity (Obe) has increased dramatically over the past few decades and has become a pandemic in not only developed but also developing countries [1]. Furthermore, Obe is usually associated with impaired fibrinolytic activity that is attributed mainly to increased plasminogen activator inhibitor-1 (PAI-1), which is the major inhibitor of the fibrinolytic system [2]. Also, PAI-1 inhibits tissue-type plasminogen activator (t-PA) and urokinase-type plasminogen activator (u-PA) [3,4].

There is much-emerging evidence that Obe is associated with increased blood PAI-1 levels which is a risk factor for higher incidence of coronary heart disease and cardiovascular thrombotic events. Also, with obesity, the higher level of PAI-1 hinders the breakdown of clots and indirectly contributes to their formation [5-7]. The pathogenesis of fibrinolytic dysfunction in obesity is correlated to body mass index (BMI), insulin resistance (IR), oxidative stress, and low-grade inflammation [8,9].

Recently, attention was paid to the prevention of obesity-associated fibrinolytic dysfunction by using natural compounds, such as "flavonoids" [10,11]. Apigenin (Api) is one of the flavonoids that is present in vegetables and fruits, e.g., parsley, celery, onions, and oranges [12]. It has been shown that Api exhibits anti-inflammatory, antioxidant, and anti-adipogenic effects [13-15]. Therefore, by curbing the accumulation of adipose tissue, Api could contribute to a decrease of PAI-1 and hence indirectly lessen the likelihood of clot formation. Moreover, Api inhibits platelet adhesion and thrombus formation [16]. Therefore, this work was designed to elaborate on the potential role of this flavonoid in ameliorating Obe-associated fibrinolytic dysfunction in an animal model.

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Materials And Methods

Diet and treatment

Apigenin, a light yellow powder with purity >98% (CAS no: 520-46-5) was purchased from Sigma-Aldrich Co. (St. Louis, MO). High-fat diet (HFD) was freshly prepared every three days. It consisted of 43% carbohydrates, 17% protein, and 40% fat and provides 414 kcal/100 g. HFD was reconstituted in a mixture of 68% normal pellet diet (NPD), 6% corn oil, 6% ghee, and 20% instant milk powder [17].

Experimental protocol

Twenty-four adult male Wistar rats weighing 175 ± 25 g and aged three months were left to acclimatize for one week before the start of the experiments. They were housed at an ambient temperature of $23 \pm 2^\circ\text{C}$, under the natural 12-hour day/night cycle with free access to food and tap water. The experimental protocol of this study was approved by the local Ethical Committee at the Faculty of Pharmacy, King Abdulaziz University, Jeddah, Saudi Arabia. The local institutional rules were in strict accordance with the international guiding principles for the care and use of laboratory animals.

The rats were divided into four following groups: control group, rats in this group were supplemented with NPD for eight weeks. Api group, rats were administered orally with Api (10 mg/kg) dissolved in 0.1% dimethyl sulfoxide (DMSO) for eight weeks. Obe group, rats supplemented with HFD for eight weeks [18]. Obe/Api group, obese rats were administered with Api (10 mg/kg) for eight weeks. Body mass index was calculated following this formula; $\text{BMI} = \text{body weight (g)} / \text{length (cm)}^2$ [19].

Serum sample collection

Following the diet and treatment period morning fasting blood samples were withdrawn from the retro-orbital venous sinuses and the blood was centrifuged at $7,000 \times g$ for five minutes in a microcentrifuge (1-15P microfuge, Sigma Microfuge, Shropshire, UK: SciQuip Ltd) and serum was separated and stored at -20°C until biochemical analysis.

Biochemical analysis

Serum glucose measurement was performed using a commercial glucose assay kit following the manufacturer's instructions. Serum insulin and TNF- α measurement were determined using rat-specific ELISA kits (San Diego, CA: MyBioSource), according to the provided protocol. Serum PAI-1 measurement was determined using a rat-specific ELISA kit, following the manufacturer's instructions. Serum MDA and t-SOD measurement - MDA, a marker of oxidative stress, and t-SOD, an indicator of the antioxidant defense system, were measured using colorimetric assay kits, following the respective protocols provided by the manufacturer. All measurements were performed in triplicate to ensure accuracy and reliability of results.

Measurement of insulin resistance

The gathered data of glucose and insulin were used to evaluate insulin resistance using the homeostatic model assessment of insulin resistance (HOMA-IR) that was calculated using the following formula: $\text{HOMA-IR} = (\text{fasting glucose in mg/dL} \times \text{fasting insulin in } \mu\text{IU/mL}) / 405$ [20].

Statistical study

The results were analyzed using the SPSS software (Chicago, IL: SPSS 23 Inc.). The numerical variables were expressed as mean \pm SD and were checked for normality using the Shapiro-Wilk test. The statistical differences between groups were determined by one-way analysis of variance (ANOVA) followed by post-hoc test of least significant difference (LSD). P-value \leq 0.05 was considered statistically significant.

Results

Effect of Api on BMI

After ingestion of HFD in the Obe group, there was a significant increase in the final BMI compared to the control and Api groups. In Obe/Api group, the final BMI was significantly lower compared to Obe group (Figure 1).

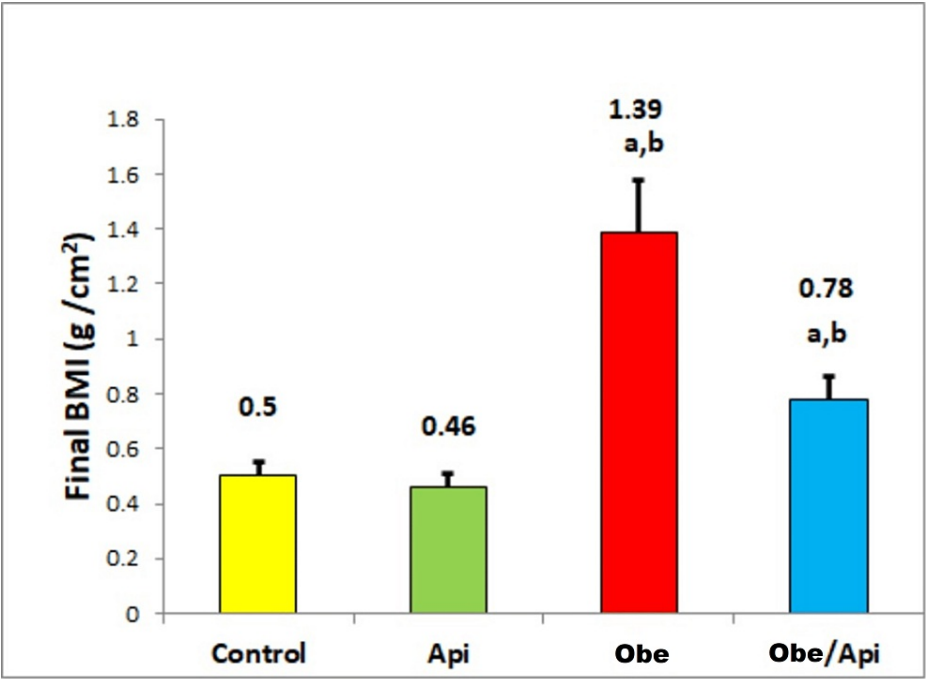


FIGURE 1: Final BMI in the control and experimental groups.

^aP-value<0.05 when compared to control and Api groups.

^bP-value<0.05 when compared to Obe group.

Api: apigenin; Obe: obesity; BMI: body mass index

The number of animals was six per group. Data are expressed as mean±SD.

Effect of Api on IR, inflammation, and oxidative stress

Ingestion of HFD resulted in significant increases in HOMA-IR, serum TNF-α, and MDA in the Obe group as compared to the control and Api groups. Also, Obe group demonstrated significant decrease in serum t-SOD activity. The addition of Api in the diet resulted in significant decrease in IR, serum TNF-α, and MDA. Furthermore, there was significant increase in serum t-SOD level in Obe/Api group as compared to Obe group (Table 1).

	Control	Api	Obe	Obe/Api	F-value	p-Value
HOMA-IR	4.95±0.53	4.13±0.42	13.60±1.39 ^a	7.36±1.42 ^{a,b}	1.93	0.015
TNF-a (pg/mL)	25.45±2.21	23.83±1.85	31.31±2.45 ^a	26.56±4.68 ^a	6.83	0.002
MDA (nmol/mL)	6.25±1.08	6.08±0.86	21.96±3.87 ^a	13.78±1.82 ^{a,b}	67.39	0.001
t-SOD (U/L)	165.00±10.41	163.66±5.92	133.83±8.61 ^a	142.16±6.55 ^a	22.381	0.02

TABLE 1: HOMA-IR and serum TNF-α, MDA, and t-SOD in the control and experimental groups.

^aP-value<0.05 when compared to control and Api groups.

^bP-value<0.05 when compared to Obe group.

Api: apigenin; Obe: obesity; HOMA-IR: homeostatic model assessment of insulin resistance; TNF-α: tumor necrosis factor-alpha; MDA: malondialdehyde; t-SOD: total superoxide dismutase, LSD: least significance difference

Values are expressed as mean±SD. Statistical analysis was carried out using one-way ANOVA followed by LSD. The number of animals was six per group.

Effect of Api on PAI-1

Obe group showed a significant increase in mean serum PAI-1 as compared to the control and Api groups. Supplementation with Api significantly decreased the mean PAI-1 in Obe/Api group when compared with Obe group (Figure 2).

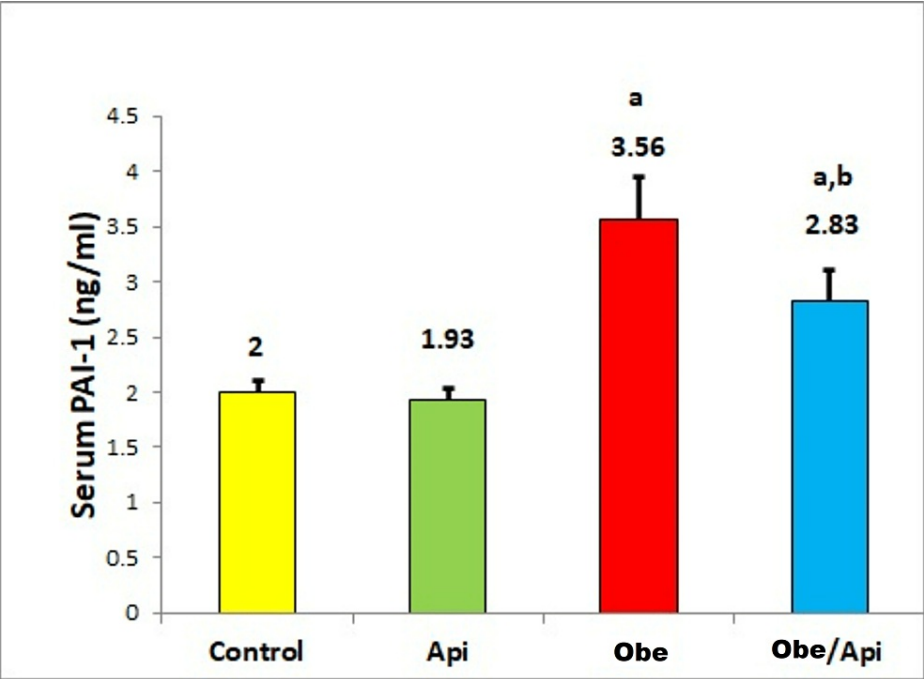


FIGURE 2: Serum PAI-1 in the control and experimental groups.

^aP-value<0.05 when compared to control and Api groups.

^bP-value<0.05 when compared to Obe group.

Api: apigenin; Obe: obesity; PAI-1: plasminogen activator inhibitor one

The number of animals was six per group. Data are expressed as mean±SD.

Discussion

Obesity is a serious metabolic disorder that is usually complicated in the long term with hemostatic complications, particularly fibrinolytic disorders. This work was settled down to elaborate on the potential therapeutic role of the flavonoid apigenin in ameliorating the fibrinolytic dysfunction in HFD-obese animal model.

In this study, Obe group exhibited a significant increase in BMI. This was concomitantly associated with a significant increase in IR. The later condition usually occurs when peripheral tissues like muscles, fat, and liver don't respond well to insulin hormone and hyperinsulinemia is a common sequel. Consequently, more lipogenesis is encountered by insulin that ultimately ends with elevations in BMI with a switch of cellular adiposity [21]. Adipocyte hypertrophy and hyperplasia are pathological features of obesity [22,23].

Obesity-induced IR is correlated positively with most of the obesity-associated complications, particularly, prothrombotic and hypofibrinolytic ones [24]. In that concern, in our results increased PAI-1 was associated with increased IR. A finding implied that the hypofibrinolytic milieu in obesity is related to IR.

The increase in BMI observed in the Obe group is commonly used as a diagnostic tool for characterizing generalized obesity. However, waist circumference (WC) represents a crude estimate of visceral obesity, which is associated with a higher risk of cardiometabolic complications [25,26]. In our study, we observed a significant decline in BMI following Api supplementation, suggesting an anti-obesity effect of this flavonoid. Moreover, Su et al. supported this finding by demonstrating the anti-visceral adiposity effect of Api through its impact on adipogenesis [15].

In this work, there was a significant increase in IR in the Obe group compared to the control groups. In obesity, various adipocyte-derived pro-inflammatory cytokines, including TNF- α , interleukin-1 β , interleukin-6, monocyte chemoattractant protein-1, leptin, and resistin are overproduced, contributing to IR [27,28]. Consistent with this, our study found a significant increase in serum TNF- α levels in the Obe group. This implied that obesity is associated with a state of systemic inflammation [29,30].

Also, Obe group displayed an increase in MDA and a decrease in t-SOD. Therefore, being a strong antioxidant flavonoid, Api ameliorated Obe-associated redox imbalance in Obe/Api group. Scientists have perpetuated that inflammation and oxidative stress are risk factors for Obe-associated disturbed fibrinolytic activity [31]. In this study, obese rats have encountered a significant elevation of serum PAI-1. This could be attributed to the concomitant rise of serum TNF- α in these rats. In alignment with this assumption, it was cited that PAI-1 synthesis is upregulated by TNF- α [32].

A previous study established a prothrombotic link between obesity and impaired fibrinolytic system in obese mice [4]. The high adipocyte-derived PAI-1 was documented as a risk factor for reduced fibrinolytic capacity, cardiovascular thrombotic events, higher incidence of coronary heart disease, and myocardial infarction [5-7]. In that regard, via a significant decrease in serum PAI-1 in Obe group, Api supplementation could indirectly prevent the previously mentioned obesity-associated thrombotic complications. Additionally, it was reported that Api, by unsettled mechanism, has the ability to inhibit platelet adhesion and thrombus formation [16].

Conclusions

The findings of this study are unique in showing the ability of Api to alleviate fibrinolytic dysfunction associated with HFD-induced obesity. The confirmed mechanisms included decreased IR, inflammation, and oxidative stress associated with obesity. Enhanced fibrinolytic activity was depicted by the decrease in serum levels of PAI-1. Also, these findings confirmed that Api has anti-adipogenic, anti-inflammatory, and antioxidant properties. Further research is warranted to explore more underlying mechanisms and fully harness the therapeutic potential of Api in managing obesity-associated complications.

Additional Information

Disclosures

Human subjects: All authors have confirmed that this study did not involve human participants or tissue.

Animal subjects: Research Ethics Committee of the Faculty of Pharmacy at King Abdulaziz University Issued protocol number PH-1443-18. **Conflicts of interest:** In compliance with the ICMJE uniform

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References

1. Boutari C, Mantzoros CS: A 2022 update on the epidemiology of obesity and a call to action: as its twin COVID-19 pandemic appears to be receding, the obesity and dysmetabolism pandemic continues to rage on. *Metabolism*. 2022, 133:10.1016/j.metabol.2022.155217
2. Skurk T, Hauner H: Obesity and impaired fibrinolysis: role of adipose production of plasminogen activator inhibitor-1. *Int J Obes Relat Metab Disord*. 2004, 28:1357-64. 10.1038/sj.ijo.0802778
3. Correia ML, Haynes WG: A role for plasminogen activator inhibitor-1 in obesity: from pie to PAI? . *Arterioscler Thromb Vasc Biol*. 2006, 26:2183-5. 10.1161/01.ATV.0000244018.24120.70
4. Lijnen HR: Role of fibrinolysis in obesity and thrombosis . *Thromb Res*. 2009, 123:46-9. 10.1016/S0049-3848(09)70143-4
5. Song C, Burgess S, Eicher JD, O'Donnell CJ, Johnson AD: Causal effect of plasminogen activator inhibitor type 1 on coronary heart disease. *J Am Heart Assoc*. 2017, 6:10.1161/JAHA.116.004918
6. Vilahur G, Ben-Aicha S, Badimon L: New insights into the role of adipose tissue in thrombosis . *Cardiovasc Res*. 2017, 113:1046-54. 10.1093/cvr/cvx086
7. Jung RG, Motazedian P, Ramirez FD, et al.: Association between plasminogen activator inhibitor-1 and cardiovascular events: a systematic review and meta-analysis. *Thromb J*. 2018, 16:10.1186/s12959-018-0166-4
8. Juhan-Vague I, Alessi MC, Mavri A, Morange PE: Plasminogen activator inhibitor-1, inflammation, obesity, insulin resistance and vascular risk. *J Thromb Haemost*. 2003, 1:1575-9. 10.1046/j.1538-7836.2003.00279.x
9. Ma LJ, Mao SL, Taylor KL, et al.: Prevention of obesity and insulin resistance in mice lacking plasminogen activator inhibitor 1. *Diabetes*. 2004, 53:336-46. 10.2337/diabetes.53.2.336
10. Vazhappilly CG, Ansari SA, Al-Jaleeli R, et al.: Role of flavonoids in thrombotic, cardiovascular, and inflammatory diseases. *Inflammopharmacology*. 2019, 27:863-9. 10.1007/s10787-019-00612-6
11. García-Barrado MJ, Iglesias-Osma MC, Pérez-García E, Carrero S, Blanco EJ, Carretero-Hernández M, Carretero J: Role of flavonoids in the interactions among obesity, inflammation, and autophagy . *Pharmaceuticals (Basel)*. 2020, 13:10.3390/ph13110342
12. Hostetler GL, Ralston RA, Schwartz SJ: Flavones: food sources, bioavailability, metabolism, and bioactivity . *Adv Nutr*. 2017, 8:423-35. 10.3945/an.116.012948
13. Madunić J, Madunić IV, Gajski G, Popić J, Garaj-Vrhovac V: Apigenin: a dietary flavonoid with diverse anticancer properties. *Cancer Lett*. 2018, 413:11-22. 10.1016/j.canlet.2017.10.041
14. Ono M, Fujimori K: Antiadipogenic effect of dietary apigenin through activation of AMPK in 3T3-L1 cells . *J Agric Food Chem*. 2011, 59:13346-52. 10.1021/jf203490a
15. Su T, Huang C, Yang C, et al.: Apigenin inhibits STAT3/CD36 signaling axis and reduces visceral obesity . *Pharmacol Res*. 2020, 152:10.1016/j.phrs.2019.104586
16. Navarro-Núñez L, Lozano ML, Palomo M, et al.: Apigenin inhibits platelet adhesion and thrombus formation and synergizes with aspirin in the suppression of the arachidonic acid pathway. *J Agric Food Chem*. 2008, 56:2970-6. 10.1021/jf0723209
17. Abdul Kadir NA, Rahmat A, Jaafar HZ: Protective effects of tamarillo (*Cyphomandra betacea*) extract against high fat diet induced obesity in Sprague-Dawley rats. *J Obes*. 2015, 2015:10.1155/2015/846041
18. Gentile D, Fornai M, Colucci R, et al.: The flavonoid compound apigenin prevents colonic inflammation and motor dysfunctions associated with high fat diet-induced obesity. *PLoS One*. 2018, 13:10.1371/journal.pone.0195502
19. Novelli EL, Diniz YS, Galhardi CM, et al.: Anthropometrical parameters and markers of obesity in rats . *Lab Anim*. 2007, 41:111-9. 10.1258/002367707779399518
20. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC: Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*. 1985, 28:412-9. 10.1007/BF00280883
21. Nishino N, Tamori Y, Kasuga M: Insulin efficiently stores triglycerides in adipocytes by inhibiting lipolysis and repressing PGC-1 α induction. *Kobe J Med Sci*. 2007, 53:99-106.
22. Sun K, Kusminski CM, Scherer PE: Adipose tissue remodeling and obesity . *J Clin Invest*. 2011, 121:2094-101. 10.1172/JCI45887
23. Ochner CN, Barrios DM, Lee CD, Pi-Sunyer FX: Biological mechanisms that promote weight regain following weight loss in obese humans. *Physiol Behav*. 2013, 120:106-13. 10.1016/j.physbeh.2013.07.009
24. Kaye SM, Pietiläinen KH, Kotronen A, et al.: Obesity-related derangements of coagulation and fibrinolysis: a study of obesity-discordant monozygotic twin pairs. *Obesity (Silver Spring)*. 2012, 20:88-94. 10.1038/oby.2011.287
25. Ross R, Neeland JJ, Yamashita S, et al.: Waist circumference as a vital sign in clinical practice: a consensus statement from the IAS and ICCR Working Group on Visceral Obesity. *Nat Rev Endocrinol*. 2020, 16:177-89. 10.1038/s41574-019-0310-7
26. Lopes HF, Corrêa-Giannella ML, Consolim-Colombo FM, Egan BM: Visceral adiposity syndrome. *Diabetol Metab Syndr*. 2016, 8:10.1186/s13098-016-0156-2
27. Lumeng CN, Saltiel AR: Inflammatory links between obesity and metabolic disease . *J Clin Invest*. 2011,

- 121:2111-7. [10.1172/JCI57132](https://doi.org/10.1172/JCI57132)
28. Jung UJ, Choi MS: Obesity and its metabolic complications: the role of adipokines and the relationship between obesity, inflammation, insulin resistance, dyslipidemia and nonalcoholic fatty liver disease. *Int J Mol Sci.* 2014, 15:6184-223. [10.3390/ijms15046184](https://doi.org/10.3390/ijms15046184)
29. de Luca C, Olefsky JM: Inflammation and insulin resistance. *FEBS Lett.* 2008, 582:97-105. [10.1016/j.febslet.2007.11.057](https://doi.org/10.1016/j.febslet.2007.11.057)
30. Suganami T, Ogawa Y: Adipose tissue macrophages: their role in adipose tissue remodeling . *J Leukoc Biol.* 2010, 88:33-9. [10.1189/jlb.0210072](https://doi.org/10.1189/jlb.0210072)
31. Mertens I, Van Gaal LF: Obesity, haemostasis and the fibrinolytic system. *Obes Rev.* 2002, 3:85-101. [10.1046/j.1467-789x.2002.00056.x](https://doi.org/10.1046/j.1467-789x.2002.00056.x)
32. Alessi MC, Bastelica D, Morange P, et al.: Plasminogen activator inhibitor 1, transforming growth factor-beta1, and BMI are closely associated in human adipose tissue during morbid obesity. *Diabetes.* 2000, 49:1374-80. [10.2337/diabetes.49.8.1374](https://doi.org/10.2337/diabetes.49.8.1374)