DOI: 10.7759/cureus.38183

Review began 04/01/2023 Review ended 04/20/2023 Published 04/27/2023

© Copyright 2023

Dalyan Cilo. This is an open access article distributed under the terms of the Creative Commons Attribution License CC-BY 4.0., which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Species Distribution and Antifungal Susceptibilities of Candida Species Isolated From Blood Culture

Burcu Dalyan Cilo 1

1. Section of Medical Mycology, University of Health Sciences, Bursa Yuksek Ihtisas Training & Research Hospital, Bursa, TUR

Corresponding author: Burcu Dalyan Cilo, bdalyan@yahoo.com

Abstract

Introduction

Candida species (spp.) are among the leading agents of bloodstream infections. Candidemias are a major cause of morbidity and mortality. Having an understanding of Candida epidemiology and antifungal susceptibility patterns in each center is crucial in guiding the management of candidemia. In this study, the species distribution and antifungal susceptibility of Candida spp. isolated from blood culture at the University of Health Sciences, Bursa Yuksek Ihtisas Training & Research Hospital were examined and the first data on the epidemiology of candidemia in our center were presented.

Methods

A total of 236 *Candida* strains isolated from blood cultures in our hospital over a four-year period were analyzed and their antifungal susceptibilities were studied retrospectively. Strains were identified at the species complex (SC) level by the germ tube test, morphology in cornmeal-tween 80 medium, and the automated VITEK 2 Compact (bioMérieux, Marcy-l'Étoile, France) system. Antifungal susceptibility tests were performed on VITEK 2 Compact (bioMérieux, Marcy-l'Étoile, France) system. The susceptibilities of the strains to fluconazole, voriconazole, micafungin, and amphotericin B were determined according to Clinical and Laboratory Standards Institute (CLSI) guidelines and epidemiologic cut-off values.

Results

Of the *Candida (C.)* strains, 131 were *C. albicans* (55.5%), 40 were *C. parapsilosis* SC (16.9%), 21 were *C. tropicalis* (8.9%), 19 were *C. glabrata* SC (8.1%), eight were *C. lusitaniae* (3.4%), seven were *C. kefyr* (3.0%), six were *C. krusei* (2.6%), two were *C. guilliermondii* (0.8%) and two were *C. dubliniensis* (0.8%).

Amphotericin B resistance was not detected in *Candida* strains. Micafungin susceptibility was 98.3%, and four *C. parapsilosis* SC strains (10%) were intermediate (I) to micafungin. Fluconazole susceptibility was 87.2%. Apart from *C. krusei* strains which intrinsically resistant to fluconazole, three *C. parapsilosis* (7.5%), one *C. glabrata* SC (5.3%) strain were resistant (R) to fluconazole, and one *C. lusitaniae* (12.5%) strain was wild-type (WT). Voriconazole susceptibility of *Candida* strains was 98.6%. Two *C. parapsilosis* SC strains were I to voriconazole, while one strain was R.

Conclusion

In this study, the first epidemiological data of candidemia agents in our hospital were presented. It was determined that rare and naturally resistant species did not cause any problem in our center yet. *C. parapsilosis* SC strains showed decreased susceptibility to fluconazole, whereas *Candida* strains were highly susceptible to the four antifungals tested. Close monitoring of these data will help guide the treatment of candidemia.

Categories: Infectious Disease

 $\textbf{Keywords:} \ \text{fungal infections, candidemia, epidemiology, candida, blood culture}$

Introduction

Candida spp. are the most common cause of fungal infections [1]. Candidemias account for 50-70% of invasive Candida infections and Candida spp. rank fourth among the agents of bloodstream infections [2]. Particularly in hospitalized, immunosuppressed and critically ill patients, candidemia is a significant cause of morbidity and mortality [3,4]. Early diagnosis is important in reducing mortality and morbidity, but since it is not easy, empirical and preemptive treatment is often planned in cases with underlying risk factors and clinical findings [2,5,6]. It is important to monitor epidemiological data and antifungal susceptibility patterns at each center to initiate appropriate therapy.

C. albicans is the most common cause of candidemia, but an increase in infections caused by non-albicans *Candida* spp. has been observed in recent years [7-10]. In many studies, it has been reported that the spectrum of agents in candidemia varies from country to country, between years and between hospitals in the same country. For this reason, it is important to conduct surveillance studies at regular intervals in terms of the management of fungal infections in hospitals [11].

The aim of this study is to determine the species distribution and antifungal susceptibilities of Candida spp. isolated from blood samples in our hospital and to present local epidemiological data.

Materials And Methods

A total of 236 *Candida* strains isolated from blood cultures at the University of Health Sciences, Bursa Yuksek Ihtisas Training & Research Hospital over a four-year period (2018-2021) and their antifungal susceptibilities were retrospectively analyzed. The bottles incubated in a fully automated blood culture system (BACTEC FX-40, Becton Dickinson, MD, USA). Yeast cells were seen in Gram-stained microscopic examination prepared from blood culture bottles with positive signals and were inoculated onto sheep blood agar (RTA Laboratories, Kocaeli, Turkey), Eosin methylene blue (EMB) agar (RTA Laboratories, Kocaeli, Turkey), and chocolate agar (RTA Laboratories, Kocaeli, Turkey). After incubation at 37°C for 24 hours, pure culture was obtained by passaging the plates with yeast growth on Sabouraud dextrose agar (SDA) (RTA Laboratories, Kocaeli, Turkey). Strains were identified to SC level by germ tube test, morphology on cornmeal-tween 80 medium and VITEK 2 YST ID (bioMérieux, Marcy-l'Étoile, France) card on VITEK 2 Compact system (bioMérieux, Marcy-l'Étoile, France). Quality control was achieved with *Candida parapsilosis* ATCC 22019 and *Candida krusei* ATCC 6258 strains.

In the determination of antifungal susceptibility, the turbidity of $\it Candida$ strains was set to 2.0 McFarland (1.8-2.2; DensiCheck, BioMérieux) with 0.45% sterile NaCl according to the manufacturer's instructions. They were loaded to the VITEK 2 AST YS08 fungal susceptibility card (BioMérieux) and the cards were placed into the instrument. This card includes amphotericin B (\leq 0.25->16 µg/ml), flucytosine (\leq 1->64µg/ml), fluconazole (\leq 0.5->64µg/ml), voriconazole (\leq 0.125->8µg/ml), caspofungin (\leq 0.125->8µg/ml) [12]. In this study, flucytosine which is not available in our country and caspofungin which produced variable results in vitro susceptibility were excluded.

In the determination of antifungal susceptibility, clinical breakpoints (CBs) in Clinical and Laboratory Standards Institute (CLSI) guideline and epidemiologic cut-off values (ECOFFs) were used (Table 1) [13-14].

| Organism | Antifungal agent | ECOFFs | CBs: (μ/ml) | | | | |
|--------------------|------------------|--------|-------------|-------|-----|----------|-------|
| | Anthungaragent | WT | Non-WT | S | SDD | I | R |
| C. albicans | Amphotericin B | ≤2 | >2 | | | | |
| | Fluconazole | | | ≤2 | 4 | | ≥8 |
| | Voriconazole | | | ≤0.12 | | 0.25-0.5 | ≥1 |
| | Micafungin | | | ≤0.25 | | 0.5 | ≥1 |
| C. glabrata SC | Amphotericin B | ≤2 | >2 | | | | |
| | Fluconazole | | | | ≤32 | | ≥64 |
| | Voriconazole | ≤0.5 | >0.5 | | | | |
| | Micafungin | | | ≤0.06 | | 0.12 | ≥0.25 |
| C. parapsilosis SC | Amphotericin B | ≤2 | >2 | | | | |
| | Fluconazole | | | ≤2 | 4 | | ≥8 |
| | Voriconazole | | | ≤0.12 | | 0.25-0.5 | ≥1 |
| | Micafungin | | | ≤2 | | 4 | ≥8 |
| C. tropicalis | Amphotericin B | ≤2 | >2 | | | | |
| | Fluconazole | | | ≤2 | 4 | | ≥8 |
| | Voriconazole | | | ≤0.12 | | 0.25-0.5 | ≥1 |
| | Micafungin | | | ≤0.25 | | 0.5 | ≥1 |
| C. krusei | Amphotericin B | ≤2 | >2 | | | | |

| | Fluconazole | | | | | |
|-------------------|----------------|--------|--------|-------|-----|----|
| | Voriconazole | | | ≤0.5 | 1 | ≥2 |
| | Micafungin | | | ≤0.25 | 0.5 | ≥1 |
| C. lusitaniae | Amphotericin B | ≤2 | >2 | | | |
| | Fluconazole | ≤2 | >2 | | | |
| | Voriconazole | ≤0.03 | >0.03 | | | |
| | Micafungin | ≤0.5 | >0.5 | | | |
| C. guilliermondii | Amphotericin B | ≤2 | >2 | | | |
| | Fluconazole | ≤8 | >8 | | | |
| | Voriconazole | ≤0.25 | >0.25 | | | |
| | Micafungin | ≤2 | >2 | | | |
| C. dubliniensis | Amphotericin B | ≤2 | >2 | | | |
| | Fluconazole | ≤0.5 | >0.5 | | | |
| | Voriconazole | ≤0.03 | >0.03 | | | |
| | Micafungin | ≤2 | >2 | | | |
| C. kefyr | Fluconazole | ≤1 | >1 | | | |
| | Voriconazole | ≤0.015 | >0.015 | | | |
| | Micafungin | ≤0.12 | >0.12 | | | |

TABLE 1: Clinical breakpoints and epidemiologic cut-off values for Candida species

ECOFFs: Epidemiologic cut-off values, CBs: Clinical breakpoints, S: Susceptible, SDD: Susceptible-dose-dependent, I: Intermediate, R: Resistant, WT: Wild-Type, NWT: Non-Wild-Type

Amphotericin B susceptibility could not be evaluated since there were no established CBs and ECOFFs for *C. kefyr*. Since the voriconazole MIC range detected by the automated antifungal susceptibility testing system we used was $\leq 0.125 - 8 \mu g/ml$, the voriconazole susceptibility of *C. kefyr* with an ECOFFs of $\leq 0.015 \mu g/ml$ and *C. lusitaniae* and *C. dubliniensis* strains with an ECOFFs of $\leq 0.03 \mu g/ml$ could not be evaluated.

Results

When the species distribution of 236 *Candida* strains isolated from blood culture in our center over four years was examined, 131 of the isolates were *C. albicans* (55.5%), 40 of them were *C. parapsilosis* SC (16.9%), and 21 of them were *C. tropicalis* (8.9%), 19 of them were *C. glabrata* SC (8.1%), eight of them were *C. lusitaniae* (3.4%), seven of them were *C. kefyr* (3.0%), six of them were *C. krusei* (2.6%), two each of them were identified as *C. guilliermondii* (0.8%) and *C. dubliniensis* (0.8%) (Table 2).

| Candida species | 2018 | 2019 | 2020 | 2021 | TOTAL |
|--------------------|------|------|------|------|-------------|
| C. albicans | 29 | 44 | 22 | 36 | 131 (55.5%) |
| C. parapsilosis SC | 10 | 13 | 8 | 9 | 40 (16.9%) |
| C. tropicalis | 3 | 8 | 5 | 5 | 21 (8.9%) |
| C. glabrata SC | 3 | 3 | 4 | 9 | 19 (8.1%) |
| C. kefyr | 2 | 2 | 1 | 2 | 7 (3.0%) |
| C. lusitaniae | 3 | 3 | 1 | 1 | 8 (3.4%) |
| C. krusei | 1 | 3 | 2 | | 6 (2.6%) |
| C. guilliermondii | 1 | - | - | 1 | 2 (0.8%) |
| C. dubliniensis | - | 2 | - | - | 2 (0.8%) |
| TOTAL | 52 | 78 | 43 | 63 | 236 (100%) |

TABLE 2: Species distribution of Candida strains

The susceptibilities of the strains to fluconazole, voriconazole, micafungin and amphotericin B are presented in Table *3. Candida* spp. were 100% WT for amphotericin B, 87.2% susceptible (S) to fluconazole, 98.6% susceptible to voriconazole and 98.3% susceptible to micafungin.

| Candida species | Amphotericin B | | | Fluconazole | | | Micafungin | | | Voriconazole | | |
|------------------------------|----------------|-------|-------|----------------|---------------|--------------|----------------|-------------|-------|----------------|----------|------------|
| | S/WT | I/SDD | R/NWT | S/WT | I/SDD | R/NWT | S/WT | I/SDD | R/NWT | S/WT | I/SDD | R/NW |
| C. albicans (n=131) | 131 (100%) | 0 | 0 | 130 (99.2%) | 1 (0.8%) | 0 | 131 (100%) | 0 | 0 | 131 (100%) | 0 | 0 |
| C. parapsilosis SC (n=40) | 40 (100%) | 0 | 0 | 37 (92.5%) | 0 | 3 (7.5%) | 36 (90%) | 4 (10%) | 0 | 37 (92.5%) | 1 (2.5%) | 2 (5.0% |
| C. tropicalis (n=21) | 21 (100%) | 0 | 0 | 21 (100%) | 0 | 0 | 21 (100%) | 0 | 0 | 21 (100%) | 0 | 0 |
| C. glabrata SC (n=19) | 19 (100%) | 0 | 0 | 0 | 18 (94.7%) | 1 (5.3%) | 19 (100%) | 0 | 0 | 19 (100%) | 0 | 0 |
| C. krusei (n=6) | 6 (100%) | 0 | 0 | 0 | 0 | 6 (100%) | 6 (100%) | 0 | 0 | 6 (100%) | 0 | 0 |
| C. lusitaniae (n=8) | 8 (100%) | 0 | 0 | 7 (87.5%) | 0 | 1 (12.5%) | 8 (100%) | 0 | 0 | * | * | * |
| C. kefyr (n=7) | ** | ** | ** | 7 (100%) | 0 | 0 | 7 (100%) | 0 | 0 | * | * | * |
| C. guilliermondii (n=2) | 2 (100%) | 0 | 0 | 2 (100%) | 0 | 0 | 2 (100%) | 0 | 0 | 2 (100%) | 0 | 0 |
| C. dubliniensis (n=2) | 2 (100%) | 0 | 0 | 2 (100%) | 0 | 0 | 2 (100%) | 0 | 0 | * | * | * |
| TOTAL (n=236) | 229 (100%) | 0 | 0 | 206 (87.2%) | 19 (8.1%) | 11 (4.7%) | 232 (98.3%) | 4 (1.7%) | 0 | 216 (98.6%) | 1 (0.5%) | 2 (0.9% |

TABLE 3: Antifungal susceptibilities of Candida strains

S: Susceptible, I: Intermediate, SDD: Susceptible-dose-dependent, R: Resistant, WT: Wild-Type, NWT: Non-Wild-Type

When the antifungal susceptibilities of the species were analyzed, it was determined that all $\it C. albicans$ strains (100%) were WT for amphotericin B and susceptible to micafungin and voriconazole. Fluconazole susceptibility was 99.2%, while one strain (0.8%) was susceptible-dose-dependent (SDD) (Table $\it 3$).

All *C. parapsilosis* SC strains were WT for amphotericin B, while micafungin susceptibility was 90% and four strains (10%) were SDD. Fluconazole susceptibility was 92.5%, three strains (7.5%) were resistant to fluconazole. Voriconazole susceptibility was 92.5%, while one strain (2.5%) was SDD and two (5.0%) were resistant (Table 3).

All *C. tropicalis* strains (100%) were susceptible to fluconazole, voriconazole, micafungin and WT for amphotericin B (Table 3).

All *C. glabrata* SC strains were susceptible to micafungin and WT for voriconazole and amphotericin B. When fluconazole susceptibility was analyzed, it was found that 94.7% of the strains were SDD to fluconazole and one strain (5.3%) was resistant (Table 3).

All *C. krusei* strains (100%) that were intrinsically resistant to fluconazole were WT for amphotericin B and susceptible to voriconazole and micafungin. All *C. kefyr* strains (100%) were WT for fluconazole and micafungin. While 87.5% of *C. lusitaniae* strains were WT for fluconazole and one strain (12.5%) was non-wild-type (NWT), 100% were WT for amphotericin B and micafungin. All *C. guilliermondii* strains (100%) were WT for fluconazole, voriconazole, amphotericin B and micafungin. All *C. dubliniensis* strains (100%) were WT for fluconazole, amphotericin B and micafungin (Table 3).

Discussion

The incidence of candidemia has increased in recent years. However, the epidemiology of *Candida* is

^{*=} Susceptibility could not be evaluated because the MIC range detected by the automated antifungal susceptibility testing system was above the epidemiological cut-off values.

^{**=} Could not be evaluated due to lack of established clinical breakpoint values and epidemiological cut-off values.

changing and the widespread use of antifungal agents for therapeutic and prophylactic purposes leads to an increase in the prevalence of infections caused by non-albicans species all over the world [15-18].

Among about 15 *Candida* species known to cause disease in humans, five species, *C. albicans, C. glabrata* SC, *C. parapsilosis* SC, *C. tropicalis* and *C. krusei* are responsible for 90% of infections [6].

C. albicans is the most commonly isolated species among candidemia agents with varying rates (37.9%-76.3%) [19-23]. Similarly, *C. albicans* (55.5%) was found to be the most common *Candida* spp. isolated in our hospital.

Among non-albicans *Candida* spp., *C. glabrata* SC is the most commonly isolated species in North America, Northern China, and Northern Europe, while *C. parapsilosis* SC ranks first in Southern Europe and Latin America [24-25]. In studies conducted in our country, *C. parapsilosis* SC has been reported as the most common species among non-albicans species with different rates (6-66%) [11, 26-32]. Consistent with this, *C. parapsilosis* SC (16.9%) was the second most frequently isolated species in our center, following *C. albicans*.

In studies from different centers, *C. tropicalis* or *C. glabrata* SC ranks second among non-albicans species [11, 26-32]. In recent years, infections caused by *C. glabrata* SC have been increasing [33]. This increase is associated with the increasing use of azoles to which *C. glabrata* SC is less susceptible [33-37]. In our study, *C. tropicalis* ranked second among non-albicans species, followed by *C. glabrata* SC.

Candida isolates resistant to clinically available antifungal agents are still rare, but are increasingly being reported worldwide [38-39]. Therefore, continuous monitoring of antifungal susceptibility patterns and mechanisms of resistance to clinically used agents becomes important [18].

Amphotericin B resistance is rare in *Candida* species. In a recent study, it was determined that amphotericin B susceptibility decreased over the years in *Candida* strains [30]. In another study examining 1062 *Candida* strains from different centers, amphotericin B resistance was not detected [40]. Similarly, in our study, all *Candida* strains were WT (those without mutational or acquired resistance mechanisms) for amphotericin B.

In the treatment of candidemia, echinocandins are widely used due to their broad-spectrum activity against *Candida* spp. [30]. Micafungin, one of the drugs recommended for the detection of echinocandin susceptibility, was included in our study. In the SENTRY program, low rates of echinocandin resistance were found in *Candida* strains (*C. albicans* 0.0%-0.1%, *C. parapsilosis* 0.0%-0.1%, *C. tropicalis* 0.5%-0.7%, *C. krusei* 0.0%-1.7%, *C. glabrata* 1.7%-3.5%) [18]. In a multicenter study in which antifungal susceptibility data of our country were presented, no echinocandin resistance was found in *Candida* strains [40]. Decreased susceptibility to echinocandins has been reported in *Candida* spp. in different studies from Latin America, Europe, and North America [41]. Likewise, in a recent study conducted in Turkey, it was revealed that echinocandin susceptibility decreased in *C. parapsilosis* SC strains over the years [30]. In our study, the echinocandin susceptibility to micafungin. These data emphasize the need for close monitoring of antifungal susceptibility patterns in each center to detect changes that may occur over the years.

In our study, we evaluated the susceptibility of fluconazole and voriconazole among azole group antifungal agents. Fluconazole is one of the drugs commonly used in the treatment of candidemia [42]. The fluconazole susceptibility of *Candida* strains was 87.3%. In addition to intrinsically fluconazole resistant *C. krusei* strains, three *C. parapsilosis* SC (7.5%) and one *C. glabrata* SC (5.3%) strains were resistant to fluconazole and one *C. lusitaniae* (12.5%) strain was WT.

C. glabrata SC is known to be less sensitive to azole drugs [42]. In studies from different countries, fluconazole resistance rates (2.8%-36.4%) are different in *C. glabrata* SC strains [43-50]. In a 20-year study involving 12 centers in our country, fluconazole resistance was found to be 0.9% in *C. glabrata* SC strains [40]. Consistent with this, only one *C. glabrata* SC strain (5.3%) was R to fluconazole in our hospital.

Fluconazole resistance in *C. parapsilosis* SC is one of the remarkable issues regarding antifungal resistance in recent years [42]. While no significant resistance was observed in strains isolated in Europe, Latin America, and Asia Pacific regions (4.6%, 4.3%, 0.6%, respectively) [18], a study conducted in South Africa found that only 37% of *C. parapsilosis* SC strains were susceptible to fluconazole [51]. In a study from our country, it was found that the susceptibility to fluconazole in *C. parapsilosis* SC strains decreased over the years (from 94% to 49%) [30]. In a study conducted in a different center in our region, fluconazole resistance was found 18.2% in *C. parapsilosis* SC [29]. In our study, fluconazole resistance was 7.5% in *C. parapsilosis* SC. Similarly, the difference in fluconazole resistance rates of *C. parapsilosis* SC strains isolated in different centers in our country was remarkable [40]. This difference is thought to be related to the clonal distribution of *C. parapsilosis* SC in which nosocomial transmission plays an important role [40].

In our study, voriconazole susceptibility in Candida strains was 98.6%. Two C. parapsilosis SC strains were

found I to voriconazole and one strain R, these three strains were R to fluconazole. Cross-resistance to voriconazole can be seen in fluconazole-resistant *Candida* strains. In the SENTRY study, only 32.7% of fluconazole-resistant *C. parapsilosis* SC strains were susceptible to voriconazole [18].

The limitation of our study was that the voriconazole MIC range that could be determined by the antifungal susceptibility test system we used, was not suitable for *C. kefyr, C. lusitaniae, C. dubliniensis*. Therefore, voriconazole susceptibility could not be evaluated in *C. kefyr, C. lusitaniae, C. dubliniensis* strains. In addition, since this was a single-center study, the findings may not be valid for other centers and may also depend on local fungal epidemiology; *Candida auris*, an emerging fungal pathogen, was not evaluated in this study and further studies will be needed for this.

Conclusions

In conclusion, this study presents the first epidemiologic data of candidemia agents in our hospital. It was determined that rare and naturally resistant species did not cause any problem in our center yet. *C. parapsilosis* SC strains showed decreased susceptibility to fluconazole, whereas *Candida* strains were highly susceptible to the four antifungals tested. It was considered that close monitoring of these data will help guide the treatment of candidemia.

Additional Information

Disclosures

Human subjects: All authors have confirmed that this study did not involve human participants or tissue. **Animal subjects:** All authors have confirmed that this study did not involve animal subjects or tissue. **Conflicts of interest:** In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

References

- Lass-Flörl C: The changing face of epidemiology of invasive fungal disease in Europe . Mycoses. 2009, 52:197-205. 10.1111/j.1439-0507.2009.01691.x
- $2. \quad \text{Pappas PG: Invasive candidiasis. Infect Dis Clin North Am. 2006, 20:485-506.} \quad 10.1016/\text{j.idc.} \\ 2006.07.004$
- Eggiman P, Garbino J, Pittet D: Epidemiology of Candida species infections in critically ill nonimmunosuppressed patients. Lancet Infect Dis. 2003, 3:685-702. 10.1016/s1473-3099(03)00801-6
- Sellami A, Sellami H, Néji S, et al.: Antifungal susceptibility of bloodstream Candida isolates in Sfax hospital: Tunisia. Mycopathologia. 2011, 171:417-422. 10.1007/s11046-010-9388-0
- Grim SA, Berger K, Teng C, Gupta S, Layden JE, Janda WM, Clark NM: Timing of susceptibility-based antifungal drug administration in patients with Candida bloodstream infection: correlation with outcomes. J Antimicrob Chemother. 2012, 67:707-714. 10.1093/jac/dkr511
- Pappas PG, Kauffman CA, Andes DR, et al.: Clinical practice guideline for the management of Candidiasis: 2016 update by the Infectious Diseases Society of America. Clin Infect Dis. 2016, 62:e1-e50. 10.1093/cid/civ933
- 7. Tsay S, Williams S, Mu Y, et al.: 363. National Burden of Candidemia, United States, 2017. Open Forum Infect Dis. 2018, 5:142-143. 10.1093/ofid/ofy210.374
- Pfaller MA, Diekema DJ: Epidemiology of invasive candidiasis: a persistent public health problem. Clin Microbiol Rev. 2007, 20:133-163. 10.1128/CMR.00029-06
- Tan TY, Tan AL, Tee NW, Ng LS, Chee CW: The increased role of non-albicans species in candidaemia: results from a 3-year surveillance study. Mycoses. 2010, 53:515-521. 10.1111/j.1439-0507.2009.01746.x
- Dimopoulos G, Ntziora F, Rachiotis G, Armaganidis A, Falagas ME: Candida albicans versus non-albicans intensive care unit-acquired bloodstream infections: differences in risk factors and outcome. Anesth Analg. 2008, 106:523-529. 10.1213/ane.0b013e3181607262
- Atalay MA, Sav H, Demir G, Koç AN: Kan kültürlerinden izole edilen Candida türlerinin dağılımı ve amfoterisin b ve flukonazole in vitro duyarlılıkları. Selçuk Tıp Derg. 2012, 28:149-151.
- VITEK® 2 AST Cards. Cards for antimicrobial susceptibility testing. (2021). Accessed: April 15, 2023: https://www.biomerieux-diagnostics.com/vitekr-2-ast-cards-0.
- Clinical and Laboratory Standards Institute (CLSI): Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts, 4th Edition. Clinical and Laboratory Standards Institute (CLSI), 2017.
- Pfaller MA, Diekema DJ: Progress in antifungal susceptibility testing of Candida spp. by use of Clinical and Laboratory Standards Institute broth microdilution methods, 2010 to 2012. J Clin Microbiol. 2012, 50:2846-2856. 10.1128/JCM.00937-12
- Pfaller MA, Diekema DJ: Epidemiology of invasive mycoses in North America. Crit Rev Microbiol. 2010, 36:1-53. 10.3109/10408410903241444
- Nucci M, Queiroz-Telles F, Tobón AM, Restrepo A, Colombo AL: Epidemiology of opportunistic fungal infections in Latin America. Clin Infect Dis. 2010. 51:561–570. 10.1086/655683
- Wang H, Xiao M, Chen SC, et al.: In vitro susceptibilities of yeast species to fluconazole and voriconazole as determined by the 2010 National China Hospital Invasive Fungal Surveillance Net (CHIF-NET) study. J Clin Microbiol. 2012, 50:3952-3959. 10.1128/JCM.01130-12

- Pfaller MA, Diekema DJ, Turnidge JD, Castanheira M, Jones RN: Twenty years of the SENTRY antifungal surveillance program: results for Candida species from 1997-2016. Open Forum Infect Dis. 2019, 6:79-94. 10.1093/ofid/ofy358
- Bakir M, Cerikcioglu N, Barton R, Yagci A: Epidemiology of candidemia in a Turkish tertiary care hospital. APMIS. 2006, 114:601-610. 10.1111/j.1600-0463.2006.apm_359.x
- Dalyan Cilo B, Ener B: Comparison of Clinical Laboratory Standards Institute (CLSI) microdilution method and VITEK 2 automated antifungal susceptibility system for the determination of antifungal susceptibility of Candida species. Cureus. 2021, 13:e20220. 10.7759/cureus.20220
- 21. Toprak NÜ, Erdoğan S, Çelik C, Johansson C: Kan kültürlerinden soyutlanan Candida suşlarının Amfoterisin B ve Flukonazole in vitro duyarlılıkları. Türk Mikrobiyol Cem Derg. 2003, 33:252-256.
- Yapar N, Uysal U, Yucesoy M, Cakir N, Yuce A: Nosocomial bloodstream infections associated with Candida species in a Turkish University Hospital. Mycoses. 2006, 49:134-138. 10.1111/j.1439-0507.2006.01187.x
- Yücesoy M, Yuluğ N: Kan kültürlerinden soyutlanan Candida türlerinin antifungal ajanlara in vitro duyarlılıkları. ANKEM Derg. 2000, 14:71-78.
- Pfaller MA, Diekema DJ, Rinaldi MG, et al.: Results from the ARTEMIS DISK Global Antifungal Surveillance Study: a 6.5-year analysis of susceptibilities of Candida and other yeast species to fluconazole and voriconazole by standardized disk diffusion testing. J Clin Microbiol. 2005, 43:5848-5859.
 10.1128/JCM.43.12.5848-5859.2005
- Pfaller MA, Pappas PG, Wingard JR: Invasive fungal pathogens: current epidemiological trends. Clin Infect Dis. 2006, 43:3-14. 10.1086/504490
- Tukenmez Tigen E, Bilgin H, Perk Gurun H, Dogru A, Ozben B, Cerikcioglu N, Korten V: Risk factors, characteristics, and outcomes of candidemia in an adult intensive care unit in Turkey. Am J Infect Control. 2017, 45:61-63. 10.1016/j.ajic.2017.02.022
- Horasan ES, Ersöz G, Göksu M, Otag F, Kurt AO, Karaçorlu S, Kaya A: Increase in Candida parapsilosis fungemia in critical care units: a 6-years study. Mycopathologia. 2010, 170:263-268. 10.1007/s11046-010-9322-5
- Kılınçel O, Akar N, Karamurat ZD, Çalışkan E, Öksüz S, Öztürk CE, Şahin I: Kan Kültürlerinden İzole Edilen Candida Türlerinin Dağılımı ve Antifungal Duyarlılıkları. Türk Mikrobiyol Cem Derg. 2018, 48:256-263. 10.5222/TMCD.2018.256
- Kazak E, Akın H, Ener B, et al.: An investigation of Candida species isolated from blood cultures during 17 years in a university hospital. Mycoses. 2014, 57:623-629. 10.1111/myc.12209
- Mete B, Zerdali EY, Aygun G, et al.: Change in species distribution and antifungal susceptibility of candidemias in an intensive care unit of a university hospital (10-year experience). Eur J Clin Microbiol Infect Dis. 2021, 40:325-333. 10.1007/s10096-020-03994-6
- Gürcüoğlu E, Ener B, Akalin H, et al.: Epidemiology of nosocomial candidaemia in a university hospital: a 12-year study. Epidemiol Infect. 2010, 138:1328-1335. 10.1017/S0950268809991531
- Çiçek-Kolak Ç, Erman-Daloğlu A, Özhak B, Öğünç D, Günseren F: Epidemiology of candidemia, antifungal susceptibilities of Candida species and their impact on mortality in adult patients admitted to Akdeniz University Hospital [Article in Turkish]. Klimik Derg. 2019, 32:250-258. 10.5152/kd.2019.71
- Lamoth F, Lockhart SR, Berkow EL, Calandra T: Changes in the epidemiological landscape of invasive candidiasis. J Antimicrob Chemother. 2018, 73:i4-i13. 10.1093/jac/dkx444
- Astvad KM, Johansen HK, Røder BL, et al.: Update from a 12-year nationwide fungemia surveillance: increasing intrinsic and acquired resistance causes concern. J Clin Microbiol. 2018, 56: 10.1128/JCM.01564-17
- Arendrup MC, Dzajic E, Jensen RH, et al.: Epidemiological changes with potential implication for antifungal prescription recommendations for fungaemia: data from a nationwide fungaemia surveillance programme. Clin Microbiol Infect. 2013, 19:343-353. 10.1111/1469-0691.12212
- Diekema D, Arbefeville S, Boyken L, Kroeger J, Pfaller M: The changing epidemiology of healthcareassociated candidemia over three decades. Diagn Microbiol Infect Dis. 2012, 73:45-48.
 10.1016/j.diagmicrobio.2012.02.001
- Lortholary O, Desnos-Ollivier M, Sitbon K, Fontanet A, Bretagne S, Dromer F: Recent exposure to caspofungin or fluconazole influences the epidemiology of candidemia: a prospective multicenter study involving 2,441 patients. Antimicrob Agents Chemother. 2011, 55:532-538. 10.1128/AAC.01128-10
- Arendrup MC: Update on antifungal resistance in Aspergillus and Candida. Clin Microbiol Infect. 2014, 20:42-48. 10.1111/1469-0691.12513
- Verweij PE, Chowdhary A, Melchers WJ, Meis JF: Azole resistance in Aspergillus fumigatus: can we retain the clinical use of mold-active antifungal azoles?. Clin Infect Dis. 2016, 62:362-368. 10.1093/cid/civ885
- Arikan-Akdagli S, Gülmez D, Doğan Ö, et al.: First multicentre report of in vitro resistance rates in candidaemia isolates in Turkey. J Glob Antimicrob Resist. 2019, 18:230-234. 10.1016/j.jgar.2019.04.003
- Pfaller MA, Boyken L, Hollis RJ, Kroeger J, Messer SA, Tendolkar S, Diekema DJ: In vitro susceptibility of invasive isolates of Candida spp. to anidulafungin, caspofungin, and micafungin: six years of global surveillance. J Clin Microbiol. 2008, 46:150-156. 10.1128/JCM.01901-07
- Pappas PG, Kauffman CA, Andes D, et al.: Clinical practice guidelines for the management of candidiasis: 2009 update by the Infectious Diseases Society of America. Clin Infect Dis. 2009, 48:503-535.
 10.1086/596757
- Doi AM, Pignatari AC, Edmond MB, et al.: Epidemiology and microbiologic characterization of nosocomial candidemia from a Brazilian National Surveillance Program. PLoS One. 2016, 11:e0146909.
 10.1371/journal.pone.0146909
- Faria-Ramos I, Neves-Maia J, Ricardo E, et al.: Species distribution and in vitro antifungal susceptibility
 profiles of yeast isolates from invasive infections during a Portuguese multicenter survey. Eur J Clin
 Microbiol Infect Dis. 2014, 33:2241-2247. 10.1007/s10096-014-2194-8
- Jung SI, Shin JH, Song JH, et al.: Multicenter surveillance of species distribution and antifungal susceptibilities of Candida bloodstream isolates in South Korea. Med Mycol. 2010, 48:669-674. 10.3109/13693780903410386

- 46. Liu W, Tan J, Sun J, et al.: Invasive candidiasis in intensive care units in China: in vitro antifungal susceptibility in the China-SCAN study. J Antimicrob Chemother. 2014, 69:162-167. 10.1093/jac/dkt330
- Minea B, Nastasa V, Moraru RF, et al.: Species distribution and susceptibility profile to fluconazole, voriconazole and MXP-4509 of 551 clinical yeast isolates from a Romanian multi-centre study. Eur J Clin Microbiol Infect Dis. 2015, 34:367-383. 10.1007/s10096-014-2240-6
- 48. Nieto MC, Tellería O, Cisterna R: Sentinel surveillance of invasive candidiasis in Spain: epidemiology and antifungal susceptibility. Diagn Microbiol Infect Dis. 2015, 81:34-40. 10.1016/j.diagmicrobio.2014.05.021
- Pemán J, Cantón E, Linares-Sicilia MJ, et al.: Epidemiology and antifungal susceptibility of bloodstream fungal isolates in pediatric patients: a Spanish multicenter prospective survey. J Clin Microbiol. 2011, 49:4158-4163. 10.1128/JCM.05474-11
- Rodriguez L, Bustamante B, Huaroto L, et al.: A multi-centric study of Candida bloodstream infection in Lima-Callao, Peru: species distribution, antifungal resistance and clinical outcomes. PLoS One. 2017, 12:e0175172. 10.1371/journal.pone.0175172
- Govender NP, Patel J, Magobo RE, et al.: Emergence of azole-resistant Candida parapsilosis causing bloodstream infection: results from laboratory-based sentinel surveillance in South Africa. J Antimicrob Chemother. 2016, 71:1994-2004. 10.1093/jac/dkw091