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# A Fatal Case of Necrotizing Soft Tissue Infection Caused by Aeromonas hydrophila Starting in the Thigh

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## Abstract

Necrotizing soft tissue infection (NSTI) remains a highly fatal disease. Among its causes, *Aeromonas hydrophila* is a Gram-negative bacillus endemic in freshwater environments that can cause fatal systemic infections in compromised hosts. The most important treatment of NSTI is source control done as soon as possible.

In this report, we describe a case of right femoral NSTI caused by *Aeromonas hydrophila* that was diagnosed based on minor physical findings, in which the patient died, despite early hip dissection. An 82-year-old man was admitted because of right femoral NSTI. Thirty hours after admission, we performed a right hip disarticulation for source control, and *Aeromonas hydrophila* was detected in the wound culture on the same day. Despite continuous treatment, he developed fungemia due to candida, eventually leading to his death.

In general, infection in a typical NSTI spreads from the extremities to the trunk, and appropriate debridement can save the patient's life. In the present case, the infection started from the thigh and may have progressed more rapidly than in other cases of NSTI.

It is important to assume *Aeromonas hydrophila* as the causative organism of NSTI in a compromised host, regardless of the history of exposure to a freshwater environment, depending on the patient's background and Gram staining results. Even if physical findings appear mild, a decision to perform hip dissection or pelvic hemisection for quick and aggressive source control may save the lives of similar patients.

**Categories:** Emergency Medicine, Infectious Disease, Orthopedics **Keywords:** aeromonas hydrophila, hip disarticulation, immunocompromised host, nsti, thigh

# Introduction

The median fatality rate of necrotizing soft tissue infection (NSTI) remains high, at 32.2%, and its prevalence globally has been reported to be 0.40 cases per 100,000 population. The disease affects all age groups, although middle-aged and elderly patients (over 50 years of age) are more likely to be infected. The most common risk factor for the development of NSTI is diabetes mellitus. The other comorbidities include obesity, hypertension, alcohol abuse, liver cirrhosis, chronic renal failure, immunodeficiency, and peripheral vascular disease. Infection begins in the hypodermis or superficial fascia. Invasive bacteria cause thrombosis of the nutrient vessels in the subcutis. Necrosis of hypodermis and superficial fascia is directly related to bacterial enzymes that destroy fascia and fat and is secondarily of vascular origin. Tissue ischemia promotes infectious dissemination, later leading to skin necrosis and, when nerve branches are involved, severe pain. Gas formed by anaerobic bacteria may cause crepitus [1]. In the treatment of NTSI, it is essential to perform debridement as early and reliably as possible [2]. Wong et al. retrospectively reviewed the medical records of 89 consecutive patients who had been admitted to their institution for NSTI from January 1997 to August 2002. It reported that the mortality rate of patients treated surgically more than 24 hours after the initial diagnosis is 9.4 times higher than that of patients treated within 24 hours, and the presence or absence of surgical treatment is directly related to saving lives [3].

Aeromonas hydrophila is a facultative anaerobic Gram-negative rod that is endemic in freshwater environments such as sewage and wastewater and soil. It is non-spore-forming with rounded ends that measure 1-3.5 µm across. It can thrive at temperatures ranging from 0°C to 45°C with an optimum temperature of 22°C-32°C [4,5]. It can cause systemic infections such as NSTI in compromised hosts and requires prompt treatment because of its fatal course.

In this report, we describe a case of right femoral NSTI caused by *Aeromonas hydrophila* that was diagnosed from minor physical findings and treated with hip dissection at an early stage but resulted in the patient's death after a dramatic course. This paper does not require approval by an ethics committee as it is a case report. The patient's family has given their consent for publication.

#### How to cite this article

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# **Case Presentation**

The patient was an 82-year-old man with a history of castration-resistant prostate cancer with liver metastasis (stage IV, clinical TNM classification: tumor, 4; nodes, 1; metastasis, 1). The patient had been treated with docetaxel for 10 months about two years prior to admission. After local radiotherapy and antihormonal therapy, he had been treated with cabazitaxel therapy for about six months prior to admission, and the disease onset was during the seventh course of cabazitaxel therapy. He had been taking prednisolone 10 mg per day as an oral anticancer drug combination since the beginning of the chemotherapy period.

The patient had been feeling unwell and aware of right lower extremity pain since the day before admission to the hospital, but he had not sought medical assistance. About 24 hours later, on the morning of the day of admission, he fell into a coma, and an emergency medical service was called. According to the patient and his family, there was no history of exposure to a freshwater environment.

The initial vital signs were as follows: Glasgow Coma Scale, 15 (eye-opening: 4, best verbal response: 5, best motor response: 6); respiratory rate, 24 beats/minute; saturation of percutaneous oxygen, unmeasurable; pulse rate, 158 beats/minute; systolic blood pressure, 69 mmHg; diastolic pressure, unmeasurable; and body temperature, 39°C. The patient was in a marked state of shock. Physical examination revealed mild erythema of 10 cm diameter long on the distal skin of the right medial thigh, but no other findings such as swelling and crepitus (Figure 1).



#### FIGURE 1: Initial skin inspection at initial presentation

There is mild erythema of 10 cm diameter long on the distal skin of the right medial thigh (black arrow).

The pain was severe and deviated from the impression of skin findings. Blood tests on admission showed marked myelosuppression, elevated CRP, acute kidney injury (AKI) (KDIGO stage 1), and elevated creatine kinase (CK) and transaminases (Table 1).

Parameter	Reference range	Day 1	Day 24	Unit of measure
Hematology				
White blood cell	3.3-8.6 × 10 <sup>3</sup>	0.9 × 10 <sup>3</sup>	0.7 × 10 <sup>3</sup>	/µL
Neutrophils	42.4-75.0	36.8	57.8	%
Lymphocytes	18.2-47.7	50	19.7	%
Monocytes	3.3-9.0	12	19.7	%
Hematocrit	40.7-50.1	32.5	25.4	%
Hemoglobin	13.7-16.8	10.7	8.6	g/dL
Red blood cell	4.35-5.55 × 10 <sup>4</sup>	332 × 10 <sup>4</sup>	291 × 10 <sup>4</sup>	/µL
Mean corpuscular volume	83.6-98.2	95.4	87.3	fL
Platelet count	15.8-34.8 × 10 <sup>4</sup>	4.2 × 10 <sup>4</sup>	1.5 × 10 <sup>4</sup>	/µL
Activated partial thromboplastin time	24.1-31.7	20	46.5	Second
Prothrombin time %	74.4-120.0	88.3	51.1	%
Blood chemistry				
Total bilirubin	0.4-1.5	2.2	22.9	mg/dL
Direct bilirubin	0.0-0.2	1.3	18.9	mg/dL
Aspartate transaminase	13-30	62	247	U/L
Alanine transaminase	10-42	105	115	U/L
Total protein	6.6-8.1	5.2	3.5	g/dL
Serum albumin	4.1-5.1	3	1.6	g/dL
Sodium	138-145	137	136	mEq/L
Potassium	3.6-4.8	3.5	4.7	mEq/L
Chloride	101-108	100	106	mEq/L
Urea nitrogen	8-20	26	28	mg/dL
Creatinine	0.65-1.07	1.72	1.15	mg/dL
Glucose	73-109	93	210	mg/dL
СРК	59-248	103	42	U/L
C-reactive protein	0.00-0.14	11.34	12.15	mg/dL

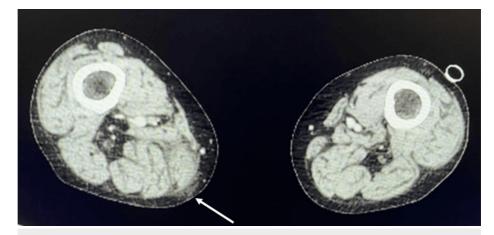
### TABLE 1: Blood tests on day 1 and day 24

The results indicate marked myelosuppression.

CPK: creatine phosphokinase

Contrast-enhanced computed tomography (CT) showed edematous changes around the gracilis muscle on the medial side of the right thigh, but no other findings (Figure 2).





#### FIGURE 2: Contrast-enhanced CT at initial presentation

There is edema around the gracilis muscle on the medial side of the right thigh (white arrow).

CT: computed tomography

However, based on the patient's marked shock and the results of the physical examination, blood tests, and CT, the decision was made to treat the patient diagnostically as having NSTI of the right thigh. A test incision made in the erythematous area of the right thigh revealed the presence of turbid exudate and a dark red myoid body that was suspicious for insufficiency of blood flow (Figure 3).



#### FIGURE 3: Findings at the time of the test incision

Incising down to the deep fascia and probing of the index finger, dark exudate is seen along with a dark red myoid body of the gracilis muscle, suspicious for insufficiency of blood flow (black arrow).

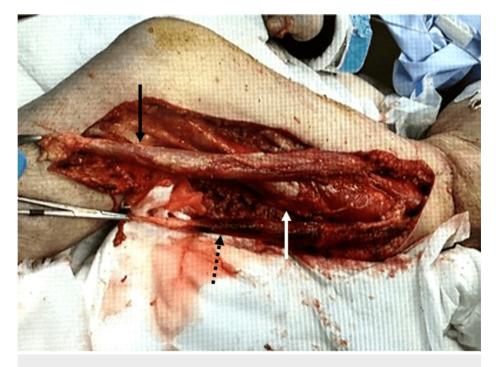
There was no bleeding associated with the incision.

Gram staining of the exudate identified Gram-negative bacillus. The rapid group A streptococcal antigen test was negative.

Based on the results of the test incision and Gram staining, we diagnosed septic shock due to right femoral



NSTI caused by a Gram-negative bacillus. For septic shock, 3 liters of extracellular fluid was administered via a peripheral venous line according to the Surviving Sepsis Campaign Guidelines. For local treatment, after cleaning the wound and submitting wound culture and blood culture, treatment was started immediately with meropenem (MEPM) 1 g/day, vancomycin (VCM) 1.5 g/day, and clindamycin (CLDM) 2700 mg/day. A central venous catheter was inserted through the right internal jugular vein, and noradrenaline and vasopressin were administered. Hydrocortisone 200 mg/day was also started for critical illness-related corticosteroid insufficiency. The patient required intubation and ventilatory management for marked shock. After admission to the intensive care unit, the right thigh was debrided, and the sartorius, gracilis, and vastus medialis muscles were resected, but the lactic acidosis rapidly progressed, and purpura appeared on the thigh (Figure 4).



#### FIGURE 4: Intraoperative photograph during debridement

After admission, the thigh was debrided, and the sartorius muscle (black arrow), gracilis muscle (black dotted arrow), and vastus medialis muscle (white arrow) were resected.

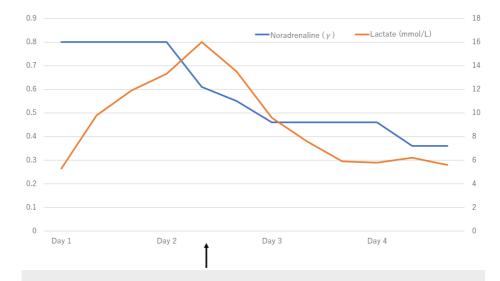
For source control, right hip disarticulation was performed 30 hours after admission (Figures 5, 6).





# FIGURE 5: Intraoperative photograph at the time of hip disarticulation

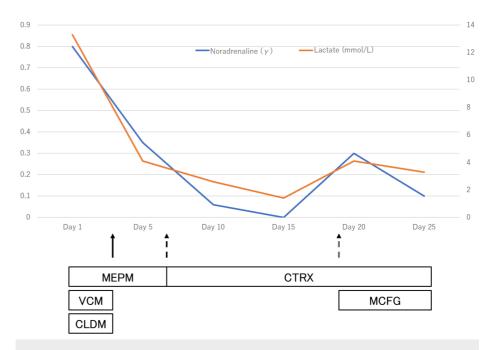
Right hip disarticulation was performed 30 hours after admission. There is an acetabulum (black arrow).



# FIGURE 6: Noradrenaline dosage and lactate: right hip disarticulation course from day 1 to day 4

The lactic acidosis rapidly progressed, and purpura appeared on the thigh. Thirty hours after admission, right hip disarticulation was performed for source control (black arrow).

Although the lactic acidosis improved, multiple organ failure was observed, including AKI, coagulopathy, and thrombocytopenia, requiring high doses of vasopressors. On the same day, *Aeromonas hydrophila* was detected in the wound culture on admission, and VCM and CLDM were terminated for de-escalation of antimicrobial agents. On the fourth day after admission, renal replacement therapy was started for metabolic acidosis. On the seventh day, sensitivity of *Aeromonas hydrophila* was found, and the antimicrobial agent was de-escalated from MEPM to ceftriaxone 2 g/day. The patient's condition began to improve. On the 15th day, the patient was weaned from ventilator management, and the demand for vasopressors was eliminated. However, progression of soft tissue necrosis in the wound was observed, the patient's blood pressure dropped on the 17th day, and the demand for vasopressors increased once more (Figure 7).



#### FIGURE 7: Noradrenaline dosage and lactate: antimicrobial course

On the third day, Aeromonas hydrophila was detected in the wound culture (black arrow). On the seventh day, the sensitivity of Aeromonas hydrophila was found (black dotted arrow). On the 19th day, Candida parapsilosis was detected in the blood culture (black double double arrow). On the 17th day, the demand for vasopressors increased once more.

MEPM: meropenem, VCM: vancomycin, CLDM: clindamycin, MCFG: micafungin, CTRX: ceftriaxone

On the 19th day, *Candida parapsilosis* was detected in wound culture, and the blood culture was submitted. On the 21st day, *Candida parapsilosis* was detected in the blood culture. Micafungin 100 mg/day was started as a treatment for deep mycosis, but *Candida parapsilosis* continued to be detected in subsequent retests of blood and wound cultures. On the 24th day, blood test results were suggestive of significant myelosuppression (Table 1). We judged that it would be difficult to save the patient's life, and the family was informed of the patient's condition on the 25th day. With their consent, we made the decision to withhold further treatment. Despite continuous multidisciplinary treatment from the day of admission, the patient died on the 26th day.

## Discussion

*Aeromonas hydrophila* is a facultative anaerobic Gram-negative bacillus that is endemic in freshwater environments such as sewage and wastewater and soil [4,5]. It is a common cause of diarrhea, wound infection, and bacteremia [6] but can also cause soft tissue infections and meningitis. Because of its low pathogenicity, it is usually a problem when it is a causative agent of opportunistic infections. In compromised hosts, systemic infections such as NSTI may occur that require prompt treatment because of the fatal course of the disease [7,8]. The mortality rate of NSTI caused by *Aeromonas hydrophila* is 80%, and it has been reported that the mortality rate increases to almost 100% in the presence of bacteremia or multiple organ failure complicating NSTI that is caused by this organism [9]. In the present case, *Aeromonas hydrophila* was detected in the wound culture, and multiple organ failure was observed, including AKI, coagulopathy, and thrombocytopenia, indicating a high likelihood of mortality.

The initial skin features of NSTI are erythema and calor. As the infection progresses, blistering, purpura, and dysesthesia are seen [10], of which blistering was a helpful clinical feature in the early diagnosis [11]. The most important treatment for NSTI is source control done as soon as possible. Specifically, it is recommended that within 24 hours of the initial debridement, a re-evaluation and debridement of the wound should be continued daily until all necrotic tissue has been removed and only healthy tissue remains [10,12,13]. It has also been reported that performing the first fasciotomy and radical debridement within 24 hours of symptom onset significantly improves survival [2,14], suggesting the need for a more rapid response in the present patient.

The above indicates that survival is remarkably low in compromised hosts with NSTI caused by *Aeromonas hydrophila* and that debridement is essential. Most reported cases of NSTI caused by *Aeromonas hydrophila* have resulted in death [15-22]. However, another report describes a case that was saved by appropriate debridement [9]. Our case was undergoing chemotherapy for carcinoma and was a long-term steroid user. He

had bone marrow suppression that may have been caused by chemotherapy or severe sepsis on blood test at the time of transport, so it was known immediately after treatment that the patient was a compromised host.

The course of the present case is characterized by the fact that although there was no history of exposure to a freshwater environment and the initial physical examination was relatively unremarkable, with only erythema of the skin, the treatment and hip disarticulation performed 30 hours after admission was not effective, and the patient developed multiple organ failure. Although the hip joint was dissected at a very early stage, the infection was still not under control, leading to multiple organ failure and deep mycosis.

A typical NSTI starts at the distal end of an extremity and spreads to the trunk. In the present case, the infection originated in the thigh, and it is possible that the infection progressed to the trunk more rapidly than in typical cases. Infection typically spreads from the toes and other parts of the body, and below-knee amputation or above-knee amputation (AKA) with appropriate debridement can be performed to save the patient's life, as has been reported in many cases [23-28]. More extensive debridement than AKA includes hip dissection and pelvic hemiarthroplasty. As these are extremely invasive for the patient, it is not easy for the emergency physician or orthopedic surgeon in charge to decide on these procedures immediately after diagnosis [9,29-31].

It is important to assume *Aeromonas hydrophila* as the causative organism in NSTI of compromised hosts, regardless of whether the patient has a history of exposure to a freshwater environment, depending on the patient background and the results of Gram staining [32]. Even if the physical findings are minor, the decision to perform a hip dissection or pelvic hemisection can be made more confidently based on the results of Gram staining, sometimes simultaneously with diagnosis. More rapid source control along with sufficient debridement may save the lives of those such as the present patient, who could not be saved this time.

# Conclusions

NSTI caused by *Aeromonas hydrophila* in a compromised host has a significantly poor prognosis. It is important to estimate the causative organism by Gram staining to make an early diagnosis. Even if the physical findings are slight, it may be necessary to be willing to perform debridement earlier and more extensively, depending on the site of onset of the disease.

# **Additional Information**

#### **Author Contributions**

All authors have reviewed the final version to be published and agreed to be accountable for all aspects of the work.

Concept and design: Masayuki Yonezu, Toshiro Imamoto

Acquisition, analysis, or interpretation of data: Masayuki Yonezu, Toshiro Imamoto

Drafting of the manuscript: Masayuki Yonezu, Toshiro Imamoto

Critical review of the manuscript for important intellectual content: Toshiro Imamoto

Supervision: Toshiro Imamoto

#### **Disclosures**

Human subjects: Consent for treatment and open access publication was obtained or waived by all participants in this study. **Conflicts of interest:** In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

# References

- 1. Misiakos EP, Bagias G, Patapis P, Sotiropoulos D, Kanavidis P, Machairas A: Current concepts in the management of necrotizing fasciitis. Front Surg. 2014, 1:36. 10.3389/fsurg.2014.00036
- Roje Z, Roje Z, Matić D, Librenjak D, Dokuzović S, Varvodić J: Necrotizing fasciitis: literature review of contemporary strategies for diagnosing and management with three case reports: torso, abdominal wall, upper and lower limbs. World J Emerg Surg. 2011, 6:46. 10.1186/1749-7922-6-46

- Wong CH, Chang HC, Pasupathy S, Khin LW, Tan JL, Low CO: Necrotizing fasciitis: clinical presentation, microbiology, and determinants of mortality. J Bone Joint Surg Am. 2003 Aug, 85(8):1454-60.
- 4. Semwal A, Kumar A, Kumar N: A review on pathogenicity of Aeromonas hydrophila and their mitigation through medicinal herbs in aquaculture. Heliyon. 2023, 9:e14088. 10.1016/j.heliyon.2023.e14088
- Stratev D, Odeyemi OA: Antimicrobial resistance of Aeromonas hydrophila isolated from different food sources: a mini-review. J Infect Public Health. 2016, 9:535-44. 10.1016/j.jiph.2015.10.006
- Fernández-Bravo A, Figueras MJ: An update on the genus Aeromonas: taxonomy, epidemiology, and pathogenicity. Microorganisms. 2020, 8:10.3390/microorganisms8010129
- Pessoa RB, de Oliveira WF, Correia MT, Fontes A, Coelho LC: Aeromonas and human health disorders: clinical approaches. Front Microbiol. 2022, 13:868890. 10.3389/fmicb.2022.868890
- Abd El-Ghany WA: A review on aeromoniasis in poultry: a bacterial disease of zoonotic nature . J Infect Dev Ctries. 2023, 17:1-9. 10.3855/jidc.17186
- Monaghan SF, Anjaria D, Mohr A, Livingston DH: Necrotizing fasciitis and sepsis caused by Aeromonas hydrophila after crush injury of the lower extremity. Surg Infect (Larchmt). 2008, 9:459-67. 10.1089/sur.2007.028
- 10. Salati SA: Necrotizing fasciitis a review. Pol Przegl Chir. 2022, 95:1-8. 10.5604/01.3001.0015.7676
- 11. Wei XK, Huo JY, Yang Q, Li J: Early diagnosis of necrotizing fasciitis: imaging techniques and their combined application. Int Wound J. 2024, 21:e14379. 10.1111/iwj.14379
- Gundersen IM, Bruun T, Almeland SK, et al.: Necrotising soft tissue infections. Tidsskr Nor Laegeforen. 2024, 144:10.4045/tidsskr.23.0720
- Chen LL, Fasolka B, Treacy C: Necrotizing fasciitis: a comprehensive review. Nursing. 2020, 50:34-40. 10.1097/01.NURSE.0000694752.85118.62
- 14. Cheung JP, Fung B, Tang WM, Ip WY: A review of necrotising fasciitis in the extremities . Hong Kong Med J. 2009, 15:44-52.
- 15. Lin CS, Cheng SH: Aeromonas hydrophila sepsis presenting as meningitis and necrotizing fasciitis in a man with alcoholic liver cirrhosis. J Formos Med Assoc. 1998, 97:498-502.
- Moses AE, Leibergal M, Rahav G, Perouansky M, Or R, Shapiro M: Aeromonas hydrophila myonecrosis accompanying mucormycosis five years after bone marrow transplantation. Eur J Clin Microbiol Infect Dis. 1995, 14:237-40. 10.1007/BF02310364
- González-Barca E, Ardanuy C, Carratalá J, Sanchez A, Fernández-Sevilla A, Grañena A: Fatal myofascial necrosis due to imipenem-resistant Aeromonas hydrophila. Scand J Infect Dis. 1997, 29:91-2. 10.3109/00365549709008672
- Huang DW, Liu HH, Dai NT, Chen SG, Tzeng YS: Necrotizing fasciitis caused by Aeromonas hydrophila with catastrophic progression. Int J Low Extrem Wounds. 2021, 20:379-83. 10.1177/1534734620973490
- 19. Janjua TK, Siddique S, Ibrahim MF, Khurshaidi MN: Aeromonas hydrophila induced necrotizing fasciitis following laparoscopic cholecystectomy. J Pak Med Assoc. 2024, 74:576-9. 10.47391/JPMA.9344
- Ugarte-Torres A, Perry S, Franko A, Church DL: Multidrug-resistant Aeromonas hydrophila causing fatal bilateral necrotizing fasciitis in an immunocompromised patient: a case report. J Med Case Rep. 2018, 12:326. 10.1186/s13256-018-1854-1
- Hong CK, Chen HC, Su WR, Lee PY: Rapid progression and mortality of necrotizing fasciitis caused by Aeromonas hydrophila in a patient with type I open tibial fracture. Kaohsiung J Med Sci. 2018, 34:307-9. 10.1016/j.kjms.2017.12.001
- 22. Hoa DT, Duc VA, Ghi NH, Duong Le X, Anh DD: Fatal septic shock due to Aeromonas hydrophila in a cirrhotic patient; a case report. Arch Acad Emerg Med. 2024, 12:e37. 10.22037/aaem.v12i1.2270
- Herzwurm ZP, Murphy CS, Griswold BG, Webber CR, Mahoney KM: From lateral ankle sprain to above-knee amputation: a unique case report of group A Streptococcus necrotizing soft-tissue infection. J Orthop Case Rep. 2020, 10:52-5. 10.13107/jocr.2020.v10.i09.1900
- 24. Abdulkareem A, D'Souza RS, Shogbesan O, Donato A: A case of rituximab-induced necrotizing fasciitis and a review of the literature. Case Rep Hematol. 2017, 2017;6971027. 10.1155/2017/6971027
- Roberts M, Crasto D, Roy D: A case of subacute necrotizing fasciitis due to Serratia marcescens. J Clin Aesthet Dermatol. 2021, 14:55-8.
- 26. Kay TJ, Wallace L, Bennett L, Gallagher PJ: Necrotising fasciitis: a case series set in rural Australia . J Surg Case Rep. 2023, 2023:rjad031. 10.1093/jscr/rjad031
- Fukuda K, Ryujin M, Sakio R, Fukuzumi S, Omae T, Hayakawa K: Bilateral necrotizing fasciitis of the foot associated with group B Streptococcus. Case Rep Dermatol. 2016, 8:243-9. 10.1159/000448163
- Tsuruta K, Ueyama T, Watanabe T, Nakano K, Uno K, Fukushima H: Intensive care management of a patient with necrotizing fasciitis due to non-O1/O139 Vibrio cholerae after traveling to Taiwan: a case report. BMC Infect Dis. 2020, 20:618. 10.1186/s12879-020-05343-6
- Brown TS, Salib CG, Rose PS, Sim FH, Lewallen DG, Abdel MP: Reconstruction of the hip after resection of periacetabular oncological lesions: a systematic review. Bone Joint J. 2018, 100-B:22-30. 10.1302/0301-620X.100B1.BII-2017-0548.R1
- Han Q, Zhang K, Zhang Y, et al.: Individual resection and reconstruction of pelvic tumor with threedimensional printed customized hemi-pelvic prosthesis: a case report. Medicine (Baltimore). 2019, 98:e16658. 10.1097/MD.00000000016658
- Cernat E, Docquier PL, Paul L, Banse X, Codorean IB: Patient specific instruments for complex tumor resection-reconstruction surgery within the pelvis: a series of 4 cases. Chirurgia (Bucur). 2016, 111:439-44. 10.21614/chirurgia.111.5.439
- 32. Tsujimoto Y, Kanzawa Y, Seto H, Nakajima T, Ishimaru N, Waki T, Kinami S: Necrotizing fasciitis and sepsis caused by Aeromonas hydrophila. Infez Med. 2019, 27:429-35.