

A Case of New Delhi Metallo- β -Lactamase-Producing Enterobacter and a Review of Cases in the United States From January 2009 to September 2022

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Abstract

Antimicrobial resistance is a growing problem. Novel resistance mechanisms continue to emerge, and the pipeline of antimicrobial development struggles to keep up. Antimicrobial stewardship and proper infection control are key in preventing the spread of these infections. A case of a carbapenem-resistant *Enterobacter cloacae* complex urinary isolate was identified in an 81-year-old male patient at the San Antonio Veterans Affairs hospital, Texas, USA. The patient was placed on isolation, and further testing of the isolate to other antibiotics requested. The purpose of this study is to analyze the details of reports of such cases and to review at-risk populations and appropriate treatment for resistant organisms.

Categories: Epidemiology/Public Health, Internal Medicine, Infectious Disease

Keywords: new delhi metallo- β lactamases (ndm), united states of america (usa), enterobacter cloacae, carbapenamase, antimicrobial resistance

Introduction

New Delhi Metallo- β -lactamase (NDM) producers are known to be resistant to multiple antimicrobial drug classes, thus making treatment difficult [1]. Infection control measures and antimicrobial stewardship (ASP) interventions have important roles to prevent spread in NDM [1]. Detection of these organisms initially occurred in individuals with travel to endemic locations where carbapenemases with the blaNDM gene were prevalent [2]. Plasmid-mediated transfer of these genes eventually resulted in spread across Enterobacterales, and the returning traveler is known to carry these isolates, resulting in imported infections in various parts of the world [2]. We report a case of the NDM isolate in an individual with no prior travel outside of San Antonio, Texas, USA, without any identifiable risk factors. We also perform a review of all cases reported in the USA from January 2009 to September 2022 to determine baseline characteristics, risks, and treatment.

Case Presentation

A 81-year-old male, with no relevant travel history, not of Southeast Asian origin, and with multiple comorbidities, including coronary artery disease, pulmonary fibrosis, prior stroke, obstructive sleep apnea, type 2 diabetes, and dyslipidemia, was hospitalized at the Veterans Affairs at San Antonio, Texas, USA, in 2021. Prior to this hospitalization, he was not on any antibiotics in the past year, nor did he have a history of multi-drug resistant organisms. He had a 5 mm left distal ureteral calculus and was not a candidate for general anesthesia given his cardiac risk. Therefore, he previously had a percutaneous nephrostomy tube placed two months prior to admission. After cardiac clearance for surgery, he had a pre-operative urine culture performed prior to planned cystoscopy, ureteroscopy, laser lithotripsy, and ureteral stent placement. The urine culture grew carbapenem-resistant *Enterobacter cloacae* complex and *Enterococcus faecalis*. The *E. cloacae* complex was a carbapenemase producer, detected by the Cepheid Carba-R, carbapenem resistance molecular test, and the antibiogram indicated resistance to all beta-lactams and carbapenems, sensitivity to tigecycline, and intermediate sensitivity to tobramycin. Cefiderocol susceptibilities were requested thereafter, and the isolate noted to be susceptible. Of the genotypes tested, blaNDM was detected. blaIMP, blaVIM, blaOXA-48, and blaKPC were not detected in this isolate. The patient was admitted five days later for the urologic procedure and was not initially placed on contact precautions. Contact precautions were initiated within 24 hours of his admission. The in-patient unit and the operating room (OR) staff were notified, and the case reported to Texas Department of State Health Services. The OR was terminally cleaned, and the patient was educated on precautions by the Infection Prevention and Control team. There were no secondary cases infected from this patient. In terms of treatment, the patient was empirically treated for his urine cultures prior to susceptibilities, with levofloxacin. Since the urologic procedure had already been performed when susceptibilities (Table 1) were reviewed, and this case was asymptomatic bacteriuria rather than a symptomatic urinary tract infection, treatment with other broader antibiotics was not pursued. He returned to urology follow-up a month later with no complaints.

How to cite this article

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Antibiotic	MIC (µg/mL)	Interpretation
Amikacin	<=4	Susceptible
Aztreonam	<=2	Susceptible
Cefepime	>16	Resistant
Cefiderocol	0.12	Susceptible
Cefotaxime	>32	Resistant
Ciprofloxacin	>2	Resistant
Colistin	<0.25	Susceptible
Doxycycline	4	Susceptible
Ertapenem	>4	Resistant
Gentamicin	2	Susceptible
Imipenem	8	Resistant
Meropenem	>8	Resistant
Minocycline	4	Susceptible
Piperacillin/Tazobactam	>64	Resistant
Polymyxin B	<=0.25	Susceptible
Ticaracillin/Clavulanic acid	>128	Resistant
Tigecycline	0.5	Susceptible
Tobramycin	8	Intermediate
Trimethoprim/Sulfamethoxazole	>4	Resistant

TABLE 1: Antibiotic susceptibility profile of NDM-producing Enterobacter cloacae complex of a case patient, with interpretation of breakpoints per the CLSI, 31st edition
CLSI: Clinical and Laboratory Standards Institute, MIC: Minimum Inhibitory Concentration

We conducted a review of NDM cases in the literature from January 2009 to September 2022, and we have summarized individual characteristics of cases identified in the United States. Google Scholar and PubMed were search engines used to search for articles, and the keywords, "New-Delhi Metallo-β-Lactamase", "NDM", "United States", "USA", "carbapenem resistant", "multi-drug resistant", and "CRE" were used to locate articles pertaining to this review. Duplicates of cases summarized in other articles as part of another author's literature review process were encountered. These were carefully identified and eliminated. To our knowledge, this is a complete review of all cases reported in the United States within this timeframe.

Discussion

The study of carbapenemase-producing strains and knowledge of their epidemiological distribution has tremendous clinical significance. As we combat various bacterial strains with antibiotics, there is a slow growth in innumerable resistance mechanisms that bacteria exhibit to circumvent the deleterious effects of antibiotics [1]. Identification of these resistant isolates early is crucial to institute contact precautions and prevent further spread [2]. We present one of the few cases of NDM in E. cloacae complex.

A review of NDM cases in the literature from January 2009 to September 2022 showed that exposure risks included travel to endemic places, as well as acquisition through inadequately sterilized equipments. Individuals with prior travel to these countries, should prompt early consideration of NDM presence, in the setting of clinical non-improvement while on standard broad-spectrum therapy. As of January 2013, the Center for Disease Control and Prevention (CDC) reported 69 patients in the United States with NDM strains, out of which 44 were from Northeastern Illinois. Twenty-three of these cases were noted to be acquired through an exposure involving culture-positive endoscope during endoscopic retrograde cholangiopancreatography (ERCP). Subsequent to this outbreak, successful gas sterilization with ethylene oxide was achieved [3]. A retrospective cohort details a total of eight patients identified on review of cases from January 2012 to October 2012 in Colorado [4]. Following these CDC reports, there have been additional cases, with one such report arising from University of Texas MD Anderson Cancer Center at Houston in 2015. All the Enterobacterales isolates that were resistant to meropenem in this hospital were tested for susceptibility to CAZ-AVI via the E test technique, or disk diffusion, confirmed by the broth microdilution method. PCR testing of various genetic sequences expressed in CREs was undertaken. Surprisingly high rates of six out of the 11 isolates with CREs were identified as NDM, out of which four had a history of foreign travel within the previous year [5].

Of the 85 patients reviewed in Tables 2-3, *E. coli* predominated as the cause, with 53 cases, or 62.4% of total infections due to this organism. With the available data, it can be ascertained that upwards of 14 cases (16.5%) were secondary to *Klebsiella*, and nine cases (10.6%) secondary to blaNDM-1 carrying *Enterobacter*. *Enterobacter* spp., as is seen in our patient, is rarely seen as a causative organism, of which *E. cloacae* complex was implicated in eight total infections in the Unites States, nine including our case patient. Infections have occurred in all (reported) age groups, varying from 13 months to 81 years, with 14 out of 23 (60.8%) cases known to occur in older individuals over 50 years of age, likely since there is a correlation between long exposure to antibiotics, resulting in increasing resistant mutations. Our patient who carried an autochthonous isolate is stipulated to have developed genetic mutations, resulting in NDM resistance due to prior exposure, since he did not have any prior travel history. Cluster of cases are known to occur with secondary infection from an index case, the largest of which is the Illinois outbreak with infection from duodenoscopes [3]. Of the dozen cases that reported all resistant genes identified, six of the isolates possessed blaCTX-M and blaTEM-1; five isolates possessed blaOXA [5-10]. Infections are identified in the USA starting from January 2011 [11].

Report	Number of cases reported	Organism	Risk factors	Genotypes reported	Patient age/gender	Isolation date	Isolation body site	State	Sensitivity profile (MIC, E test reported as µg/ml)	Treatment	C
Frias et al. [3]	44	<i>Escherichia coli</i>	Prior ERCP	blaNDM	NR	Jan to Dec 2013	Clinical culture unclassified; rectal surveillance culture	Illinois	NR	NR	N
Epson et al. [4]	8	Unknown case number of <i>Klebsiella pneumoniae</i> ; <i>Enterobacter</i> ; <i>Citrobacter</i>	NR	blaNDM	NR	Jan-Oct 2012	Urine; respiratory; rectal surveillance; other unclassified	Colorado	NR	NR	N
Pecora et al. [6]	4	<i>Escherichia coli</i>	Travel to India	blaNDM-5; blaCTX-M-15; blaTEM-1; blaOXA-1	NR	2015	Urine	Massachusetts	Tigecycline (Etest 0.25); intermediate to chloramphenicol (KB); colistin (MIC 0.06)	NR	N
Pecora et al. [6]	As above	<i>Escherichia coli</i>	Travel to China	blaNDM-5; blaTEM-1	NR	2015	Abscess	Massachusetts	Tigecycline (Etest 0.75); colistin (MIC 0.06)	NR	N
Pecora et al. [6]	As above	<i>Escherichia coli</i>	No known risk factors	blaNDM-1; blaCTX-M-15; blaTEM-1b; blaOXA-1; blaCMY-6	NR	2016	Urine	Massachusetts	Tigecycline (Etest 0.125); sensitive to tetracycline (MIC 2); colistin (MIC 0.12)	NR	N
Pecora et al. [6]	As above	<i>Escherichia coli</i>	Travel to India	blaNDM-5; blaCTX-M-15; blaTEM-1; blaOXA-1	NR	2016	Blood	Massachusetts	Tigecycline (Etest 0.38); sensitive to tetracycline (KB); intermediate to chloramphenicol (KB); colistin (MIC 0.12)	As above	A
Mediavilla et al. [10]	1	<i>Escherichia coli</i>	Travel to India	blaNDM-5; blaOXA-1; mcr1, strA, strB, aac(6')-Ib-cr, catB3, floR, arr-3, sul1, sul2, tet(A)	76/M	Aug 2014	Urine	New Jersey	Sensitive to gentamicin, TMP-SMX	NR	N
Green et al. [12]	2 in case patient	<i>Escherichia coli</i>	Travel to India	blaNDM-1	7/F	Nov 2012	Urine	California	Susceptible to tigecycline, fosfomycin Susceptible to fosfomycin,	5 days of tigecycline, then a single dose of fosfomycin	C

									intermediate to tigecycline		
Green et al. [12]	As above	<i>Klebsiella pneumoniae</i>	As above	bla _{NDM-1}	As above	Dec 2012	As above	As above	NR	6 days of tigecycline, concomitant with single dose of fosfomycin on day 1	C
Lee et al. [9]	2 in case patient	<i>Klebsiella pneumoniae</i>	Travel to India	bla _{NDM-7} , bla _{CTX-M-15} , rmtF	69/M	2012	Intra-abdominal abscess	Minnesota	Tigecycline (MIC 1); colistin (MIC <=0.25); polymyxin B (MIC <=0.25)	NR	N
Lee et al. [9]	As above	<i>Escherichia coli</i>	As above	bla _{NDM-7} , bla _{CTX-M-15}	As above	As above	As above	As above	Tigecycline (MIC <=0.25); colistin (MIC <=0.25); polymyxin B (MIC <=0.25)	NR	N
Aitken et al. [5]	5	<i>Escherichia coli</i>	Travel to Venezuela	bla _{NDM-1}	22/M	2015	Scrotal abscess	Texas	NR	Tigecycline+ aztreonam+ meropenem+ colistin	C
Aitken et al. [5]	As above	<i>Escherichia coli</i>	Travel to China	bla _{NDM-9}	31/M	2015	Stool	Texas	NR	Aztreonam+CAZ-AVI+tigecycline+TMP-SMX	C
Aitken et al. [5]	As above	<i>Klebsiella pneumoniae</i>	Travel to Mexico	bla _{NDM-1}	27/M	2015	Sputum	Texas	NR	Imipenem+ cilastatin+ ertapenem+ TMP-SMX+ tigecycline+ colistin+ amikacin	C
Aitken et al. [5]	As above	<i>Klebsiella pneumoniae</i>	Travel to India	bla _{NDM-1}	68/F	2015	Urine	Texas	NR	Colistin	C
Aitken et al. [5]	As above	<i>Klebsiella oxytoca</i>	No known risk factors	bla _{NDM-1}	3/F	2015	Stool	Texas	NR	Colistin+ tigecycline+ cefepime	C
Mochon et al. [13]	2 in case patient	<i>Klebsiella pneumoniae</i>	Travel to Pakistan	bla _{NDM-1}	13 mo/M	NR	Nasal wash	California	Colistin (MIC 0.25); sensitive to tigecycline (MIC 1); minocycline (MIC 4)	Colistin	C
Mochon [13]	As above	<i>Klebsiella pneumoniae</i>	As above	bla _{NDM-1}	As above	As above	Sputum	As above	Sensitive to tigecycline (MIC <=0.5); minocycline (MIC <=4)	As above	A
Li et al. [7]	1	<i>Klebsiella pneumoniae</i>	Travel to Iran	bla _{NDM-1} , bla _{CTX-M-15} , bla _{SHV-12} , bla _{TEM-1} , rmtC, armA	72/F	Feb 2014	Hip wound culture	Florida	Susceptible to tigecycline, colistin	NR	N
Mittal et al. [14]	1	<i>Klebsiella pneumoniae</i>	Travel to Bangladesh	bla _{NDM-1} , bla _{OXA-48}	42/M	NR	Elbow wound culture	New York	Intermediate to tigecycline (MIC 3); polymyxin B (MIC 0.05); synergy to CAZ-AVI, aztreonam (FIC index 0.11)	Meropenem+ tigecycline on day 1, day 2, then meropenem+ polymyxin B day 3, day 4, then polymyxin B day 5 to 15, then CAZ-AVI+ aztreonam days 16 to 21, then polymyxin B on days 22 to 29, then CAZ-AVI+ aztreonam until day 62	C
Hardy et al. [2]	2	<i>Klebsiella pneumoniae</i>	Travel to Vietnam	bla _{NDM}	NR	Feb; Mar 2012	Urine	Rhode island	Susceptible to tigecycline (MIC 2), colistin, polymyxin B	NR	N

									(MIC 1) NR		
Hardy et al. [2]	As above	<i>Klebsiella pneumoniae</i>	exposure to index case	bla _{NDM}	NR	Mar 2012	Rectal swab	Rhode island	As above	NR	N
Chen et al. [15]	1	<i>Klebsiella pneumoniae</i>	Travel to India	bla _{NDM}	70/F	Aug 2016	Wound culture	Nevada	Tigecycline intermediate; fosfomycin (Etest 16)	NR	C
Toomer et al. [16]	1	<i>Klebsiella pneumoniae</i>	No known risk factors	bla _{NDM}	53/M	NR	Blood	Florida	Sensitive to tetracycline, tigecycline; synergy to CAZ-AVI, aztreonam (FIC 0.375)	Tigecycline+ CAZ-AVI+ aztreonam from day 1 to 8, then CAZ-AVI+ aztreonam, unknown duration	N
Savard et al. [11]	2 in case patient	<i>Klebsiella pneumoniae</i>	Travel to India	bla _{NDM}	60/M	Jan 2011	Sputum	Maryland	Sensitive to colistin (MIC 0.12)	NR	N
Savard et al. [11]	As above	<i>Salmonella enterica subspecies enterica serovar Seftenberg</i>	As above	bla _{NDM-1}	As above	As above	Perirectal culture	As above	Susceptible to tetracycline, tigecycline, TMP-SMX	NR	N

TABLE 2: Summary of cases with NDM isolated in the United States (Jan 2009-Sept 2022) secondary to species other than Enterobacter

ERCP: Endoscopic Retrograde Cholangiopancreatography; NR: Not Reported; KB: Kirby-Bauer Disk Diffusion Testing; MIC: Minimum Inhibitory Concentration; TMP-SMX: Trimethoprim-Sulfamethoxazole; CAZ-AVI: Ceftazidime-Avibactam

Report	Number of cases reported	Organism	Risk factors	Genotype	Patient age/gender	Isolation date	Isolation body site	State	Sensitivity profile (MIC, Etest reported as µg/ml)	Treatment	Out
Pecora et al. [6]	1	<i>Enterobacter cloacae</i> complex	No known risk factors	bla _{NDM-1} , bla _{SHV-7} , bla _{SHV-12} , bla _{TEM-1} , bla _{ACT25}	NR	2015	Blood	Massachusetts	Tigecycline (Etest 0.75); sensitive to tetracycline (MIC 4); colistin (MIC 0.06)	NR	NR
Aitken et al. [5]	1	<i>Enterobacter cloacae</i>	Travel to Mexico	bla _{NDM-1} , bla _{KPC-17}	27/M	2015	Respiratory	Texas	NR	Imipenem+ cilastatin+ ertapenem+ TMP-SMX+ tigecycline+ colistin+ amikacin	Dea
Yasmin et al. [8]	1	<i>Enterobacter hormaechei subsp.hoffmannii strain Eh</i>	NR	bla _{NDM} , bla _{KPC}	4/M	NR	Blood, perianal swab	Ohio	Synergy to CAZ-AVI, aztreonam	CAZ-AVI+ aztreonam for 14 days	Cure
Siddamreddy et al. [17]	1	<i>Enterobacter cloacae</i>	NR	bla _{NDM-1}	75/F	NR	Leg wound	Not reported	Sensitive to amikacin (MIC <=8), aztreonam (MIC <=4), gentamicin (MIC <=1), tigecycline (MIC <=1), tobramycin, intermediate to ciprofloxacin (MIC 0.5), levofloxacin	Tigecycline	Cure

									(MIC 1)		
Nori [18]	5	Enterobacter cloacae	No known risk factors	bla _{NDM}	68/F	Apr 2020	Blood, respiratory	New York	Sensitive to Colistin (MIC <=0.25); gentamicin (MIC <=2); tigecycline (MIC <=1)	Tigecycline+CAZ-AVI+aztreonam	NR
Nori et al. [18]	As above	Enterobacter cloacae	No known risk factors	bla _{NDM}	57/M	Apr 2020	Urine, respiratory	New York	Sensitive to Colistin (MIC 0.5); gentamicin (MIC <=2); tigecycline (MIC <=1)	Tigecycline	NR
Nori et al. [18]	As above	Enterobacter cloacae	No known risk factors	bla _{NDM}	63/M	Apr 2020	Blood, respiratory	New York	Sensitive to colistin (MIC 0.5); gentamicin (MIC 4); tigecycline (MIC <=1)	Tigecycline+ gentamicin	NR
Nori et al. [18]	As above	Enterobacter cloacae	No known risk factors	bla _{NDM}	63/F	Apr 2020	Urine, respiratory	New York	Intermediate to colistin (MIC <=0.25); sensitive to gentamicin (MIC <=2); tigecycline (MIC <=1)	CAZ-AVI+ aztreonam	NR
Nori et al. [18]	As above	Enterobacter cloacae	No known risk factors	bla _{NDM}	54/M	May 2020	Respiratory	New York	Intermediate to colistin (MIC <=0.25); sensitive to gentamicin (MIC <=2); tigecycline (MIC <=1)	Tigecycline,gentamicin,CAZ-AVI,aztreonam	NR
This case	1	Enterobacter cloacae	No known risk factors	bla _{NDM}	81/M	Apr 2021	Urine	Texas	Sensitive to tigecycline, ceftiderocol and intermediate to tobramycin	None	Clin no con

TABLE 3: Summary of case reports of NDM cases in the United States (Jan 2009-Sept 2022) due to Enterobacter spp.

KB: Kirby-Bauer Disk Diffusion Testing; MIC: Minimum Inhibitory Concentration; TMP-SMX: Trimethoprim-Sulfamethoxazole; CAZ-AVI: Ceftazidime-Avibactam; FIC: Fractional Inhibitory Concentration Index

Of the *E. coli* isolates with reported sensitivities, six out of seven (85.7%) was sensitive to tigecycline, followed by three out of seven (42.9%) sensitive to colistin, one each sensitive to trimethoprim-sulfamethoxazole (TMP-SMX), tetracyclines, polymyxin B, and fosfomycin. Of the *K. pneumoniae* isolates with reported sensitivities, five out of 10 (50%) showed sensitivity each to tigecycline and colistin; two isolates (20%) showed sensitivity to fosfomycin, minocycline, and polymyxin B; and one isolate was sensitive to tetracycline. Of the *E. cloacae* complex reported, all isolates, including our patient, were sensitive to tigecycline; five out of eight isolates (62.5%) were sensitive to gentamicin; four isolates (50%) to colistin; and one isolate to tetracycline and tobramycin. Our case patient had a sensitive isolate to ceftiderocol, although it is uncertain whether the other reported isolates were tested to this antibiotic. This highlights empiric use of tigecycline, colistin, and gentamicin for blaNDM carrying isolates, with additional sensitivities to be requested to ceftiderocol. With higher minimum inhibitory concentration (MIC) to CAZ-AVI seen in isolates, and clinical decline in several cases, the fractional inhibitory concentration (FIC) index becomes necessary. Out of the cases reviewed, only three noted synergy by this testing method. Validation of

this test will be helpful in the care of patients.

Plasmid-transfer of the NDM gene from other Enterobacterales likely resulted in *Enterobacter* spp. acquiring this resistance pattern. The first ever case of *Enterobacter* with blaNDM was seen in 2015 [14]. In our case, there was no secondary infection in the seven month follow-up period after terminal disinfection. CAZ-AVI and aztreonam, tigecycline, and gentamicin have been used successfully to eradicate the infection [8,12,17]. More studies are required to assess difference in clinical outcome, eradication of the isolate via infection control practices, and different susceptibilities to antimicrobials in *Enterobacter* spp. compared to other Enterobacterales. Caution should be undertaken when reviewing clinical data from individuals with history of extensive antibiotic exposure, as well as resistance prevalent countries, with low threshold in suspecting NDM infections. To accurately identify every single case, it is also prudent for Infection control in a hospital to perform routine checks on all patients, as individuals like our patient could rarely present with no risk factors. System errors that occurred in our case can be curbed with alerts indicating buzzwords such as “highly resistant”, “isolation mandated immediately”, and “New-Delhi metallo- β -lactamase”, with explanation requested in case the provider defers isolation.

In our review, we identified that *K. pneumoniae* and *E. coli* were predominantly implicated in carrying the NDM gene in the United States. Exposure risks to acquiring an NDM isolate are travel to endemic places and the use of inadequately sterilized equipments. Cluster of cases have been reported with secondary infection from an index case. Of the 85 patients we reviewed carrying an NDM isolate in the United States, *E. coli* predominated as the cause, with 53 cases, or 62.4% of total infections due to this organism. Meanwhile, 14 cases (16.5%) were secondary to *Klebsiella*, and nine cases (10.6%) secondary to blaNDM-1 carrying *Enterobacter*. Our case patient is the only individual case reported in the prior year in the United States with no relevant travel history and no immediately prior antibiotic exposure who was identified to carry a resistant isolate. Of the *E. cloacae* complex reported, all isolates, including our patient were sensitive to tigecycline; five out of eight isolates (62.5%) were sensitive to gentamicin; four isolates (50%) to colistin; and one isolate to tetracycline and tobramycin. Susceptibility testing in our case patient indicated sensitivity to cefiderocol, although this antibiotic was not tested against most of the isolates captured in this review. This highlights empiric use of tigecycline, colistin, and gentamicin for blaNDM carrying isolates, with additional sensitivities to be requested to cefiderocol.

Conclusions

NDM-producing Enterobacterales are a growing concern with pan-resistance being reported in the past and concerns of loss of synergy to CAZ-AVI and aztreonam in the future. Worldwide spread is known to occur with multiple areas of the United States affected to date as a result of travel from a prevalent location. The plasmid-mediated transfer results in fast transmission of resistance among Enterobacterales. NDM-producing strains are being isolated in individuals with no travel history as well, given community acquisition of resistance in the recent past. This requires increased awareness for adherence to applied infection prevention practices. Overall, the importance of appropriate contact precautions, hand hygiene, antimicrobial stewardship, and rapid laboratory detection has been crucial in the prevention of highly resistant organism transmission. We must all continue to be vigilant and cautious as these resistant organisms continue to be reported. Special importance to members from prevalent countries can help with early identification, and appropriate Infection control department routine screening is necessary to identify individuals with no risk factors.

Additional Information

Disclosures

Human subjects: Consent was obtained or waived by all participants in this study. **Conflicts of interest:** In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

References

1. Reygaert WC: An overview of the antimicrobial resistance mechanisms of bacteria. *AIMS Microbiol.* 2018, 4:482-501. [10.3934/microbiol.2018.3.482](https://doi.org/10.3934/microbiol.2018.3.482)
2. Hardy EJ, Mermel LA, Chapin KC, et al.: Carbapenem-resistant Enterobacteriaceae containing New Delhi metallo- β -lactamase in two patients — Rhode Island, March 2012. *MMWR Morb Mortal Wkly Rep.* 2012, 61:446-8.
3. Frias M, Tsai V, Moulton-Meissner H: New Delhi metallo- β -lactamase-producing *Escherichia coli* associated with endoscopic retrograde cholangiopancreatography — Illinois, 2013. *MMWR Morb Mortal Wkly Rep.* 2014, 62:1051.
4. Epson EE, Pisney LM, Wendt JM, et al.: Carbapenem-resistant *Klebsiella pneumoniae* producing New Delhi metallo- β -lactamase at an acute care hospital, Colorado, 2012. *Infect Control Hosp Epidemiol.* 2014, 35:390-7. [10.1086/675607](https://doi.org/10.1086/675607)
5. Aitken SL, Tarrand JJ, Deshpande LM, et al.: High rates of nonsusceptibility to ceftazidime-avibactam and identification of New Delhi metallo- β -lactamase production in Enterobacteriaceae bloodstream infections at a major cancer center. *Clin Infect Dis.* 2016, 63:954-8. [10.1093/cid/ciw398](https://doi.org/10.1093/cid/ciw398)
6. Pecora N, Zhao X, Nudel K, et al.: Diverse vectors and mechanisms spread New Delhi metallo- β -lactamases among carbapenem-resistant Enterobacteriaceae in the Greater Boston area. *Antimicrob Agents Chemother.* 2019, 63:10.1128/AAC.02040-18
7. Li JJ, Munoz-Price LS, Spykhal CN, DePascale D, Doi Y: New Delhi metallo- β -lactamase-1-producing *Klebsiella pneumoniae*, Florida, USA. *Emerg Infect Dis.* 2016, 22:744-6. [10.3201/eid2204.151176](https://doi.org/10.3201/eid2204.151176)

8. Yasmin M, Fouts DE, Jacobs MR, et al.: Monitoring ceftazidime-avibactam and aztreonam concentrations in the treatment of a bloodstream infection caused by a multidrug-resistant *Enterobacter* sp. carrying both *Klebsiella pneumoniae* carbapenemase-4 and New Delhi metallo- β -lactamase-1. *Clin Infect Dis*. 2020, 71:1095-8. [10.1093/cid/ciz1155](https://doi.org/10.1093/cid/ciz1155)
9. Lee CS, Vasoo S, Hu F, Patel R, Doi Y: *Klebsiella pneumoniae* ST147 coproducing NDM-7 carbapenemase and RmtF 16S rRNA methyltransferase in Minnesota. *J Clin Microbiol*. 2014, 52:4109-10. [10.1128/JCM.01404-14](https://doi.org/10.1128/JCM.01404-14)
10. Mediavilla JR, Patrawalla A, Chen L, et al.: Colistin- and carbapenem-resistant *Escherichia coli* harboring MCR-1 and blaNDM-5, causing a complicated urinary tract infection in a patient from the United States. *mBio*. 2016, 7:[10.1128/mBio.01191-16](https://doi.org/10.1128/mBio.01191-16)
11. Savard P, Gopinath R, Zhu W, et al.: First NDM-positive *Salmonella* sp. strain identified in the United States. *Antimicrob Agents Chemother*. 2011, 55:5957-8. [10.1128/AAC.05719-11](https://doi.org/10.1128/AAC.05719-11)
12. Green DA, Srinivas N, Watz N, Tenover FC, Amieva M, Banaei N: A pediatric case of New Delhi metallo- β -lactamase-1-producing *Enterobacteriaceae* in the United States. *Pediatr Infect Dis J*. 2013, 32:1291-4. [10.1097/INF.0b013e31829eca34](https://doi.org/10.1097/INF.0b013e31829eca34)
13. Mochon AB, Garner OB, Hindler JA, et al.: New Delhi metallo- β -lactamase (NDM-1)-producing *Klebsiella pneumoniae*: case report and laboratory detection strategies. *J Clin Microbiol*. 2011, 49:1667-70. [10.1128/JCM.00183-11](https://doi.org/10.1128/JCM.00183-11)
14. Mittal J, Szymczak WA, Guo Y, et al.: Two for the price of one: emerging carbapenemases in a returning traveller to New York City. *BMJ Case Rep*. 2018, 2018: [10.1136/bcr-2018-225440](https://doi.org/10.1136/bcr-2018-225440)
15. Chen L, Todd R, Kiehlbauch J, Walters M, Kallen A: Notes from the field: pan-resistant New Delhi metallo-beta-lactamase-producing *Klebsiella pneumoniae* — Washoe County, Nevada, 2016. *MMWR Morb Mortal Wkly Rep*. 2017, 66:33.
16. Toomer KH, de Lima Corvino D, McCrink KA, Gonzales Zamora JA: A New Delhi metallo- β -lactamase (NDM)-positive isolate of *Klebsiella pneumoniae* causing catheter-related bloodstream infection in an ambulatory hemodialysis patient. *IDCases*. 2020, 21:e00816. [10.1016/j.idcr.2020.e00816](https://doi.org/10.1016/j.idcr.2020.e00816)
17. Siddamreddy S, Meegada S, Dandu V, Muppidi V, Bachu R: New Delhi metallo-beta-lactamase-producing *Enterobacter cloacae* - a rare multidrug resistance strain in a Caucasian woman. *Cureus*. 2020, 12:e7177. [10.7759/cureus.7177](https://doi.org/10.7759/cureus.7177)
18. Nori P, Szymczak W, Puius Y, et al.: Emerging co-pathogens: New Delhi metallo-beta-lactamase producing *Enterobacterales* infections in New York City COVID-19 patients. *Int J Antimicrob Agents*. 2020, 56:106179. [10.1016/j.ijantimicag.2020.106179](https://doi.org/10.1016/j.ijantimicag.2020.106179)